

Prendre en compte les spécificités des fibres végétales pour un renforcement optimal des composites biobasés

Alain BOURMAUD

GDR FIBMAT – ENSTA Brest, le 13 novembre 2024



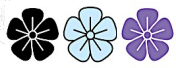


AGENDA OF THE PRESENTATION

- **Brief historical context**
- **Value chain and retting stage**
- **A large diversity of plant fibres**
- **Development and ultrastructure of plant fibres**
- **Multiscale mechanical characterization of plant fibres**
- **Structural defects: a key specificity**
- **Biobased composites processing**



BRIEF HISTORICAL CONTEXT



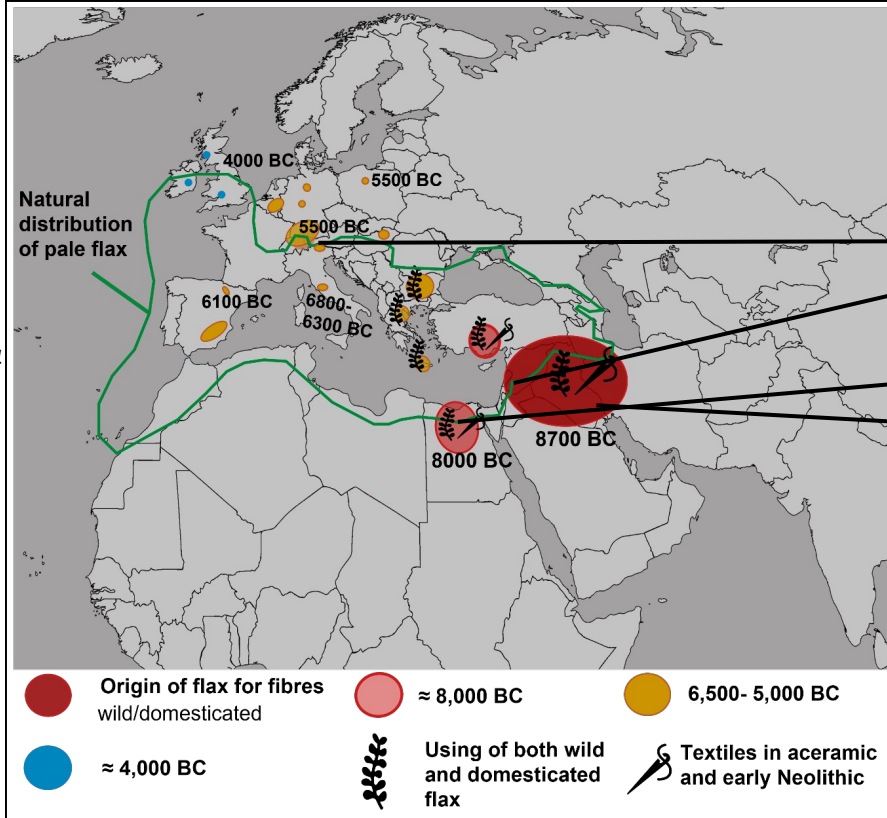
FIRST TRACES - DOMESTICATION

Melelli et al., under review

Image inspired by:

Karg, *Veg. Hist. Archaeobot.*, 2011

Harris, in: *Textile Society of America 2014 Biennial Symposium Proceedings.*, 2014



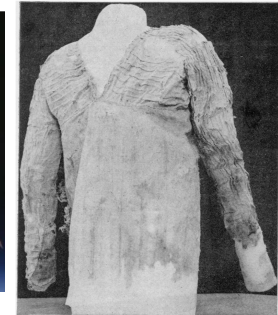
Sennedjem's tomb, Valley of the Artisans (Deir el-Medina)



© Paolo Bondielli



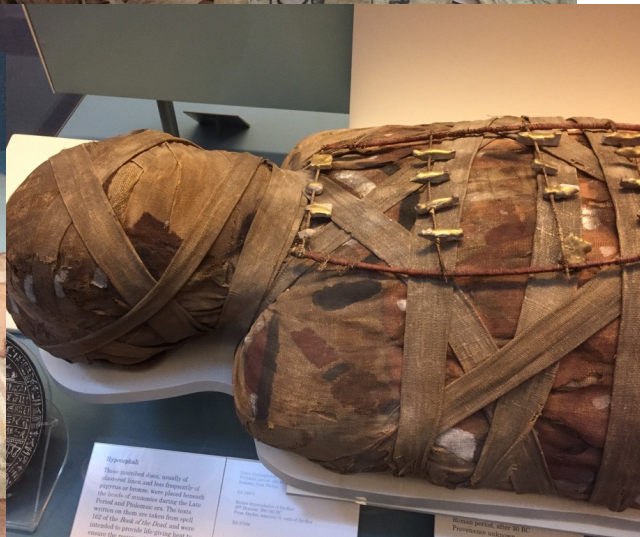
© British Museum



Landi & Hall, *Stud. Conserv.*, 1979

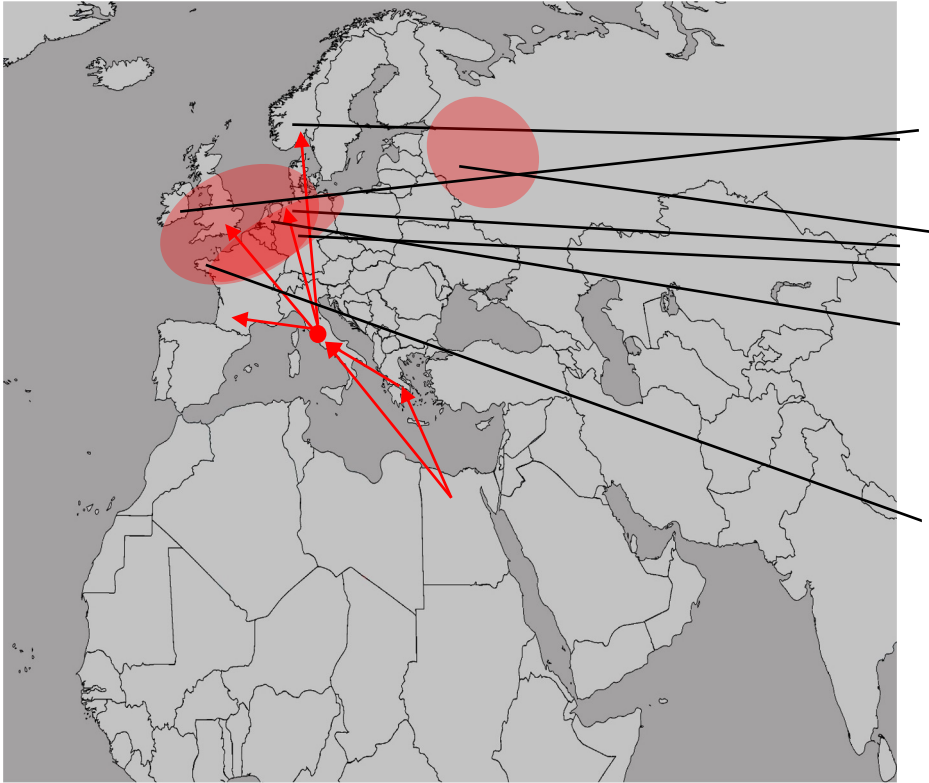


 IN EGYPT





THE MAIN CULTIVATION CENTRES



Flax Combers, Fedot Sychkov, 1905

© Mordovian Erzia Museum of Visual Arts

The processing of flax remained unchanged for centuries



IN EUROPE, MORE RECENTLY

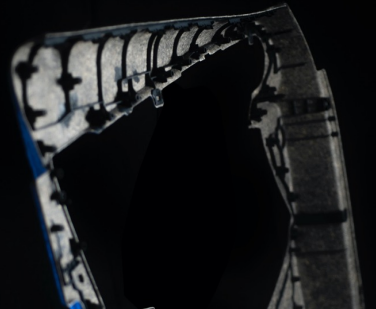


In the 17th century, about 300,000 hectares of flax were cultivated in France (today 150,000)

About 20% of textiles are made of flax (less than 0.4% today!)





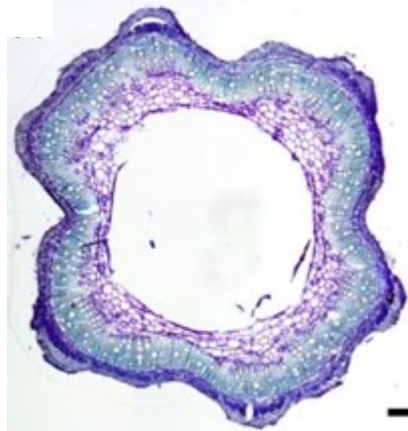






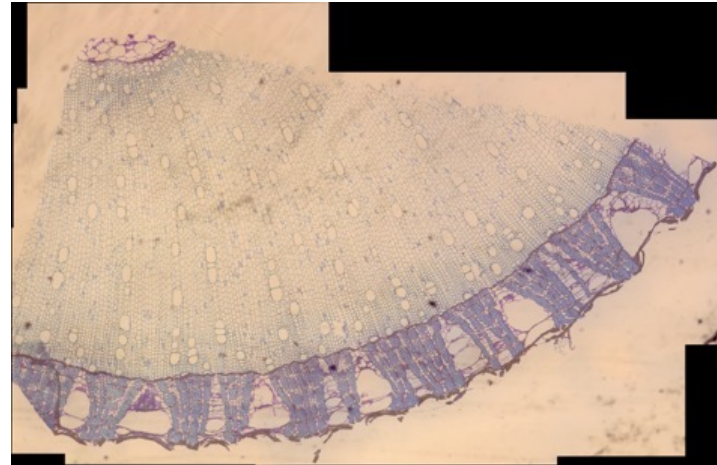
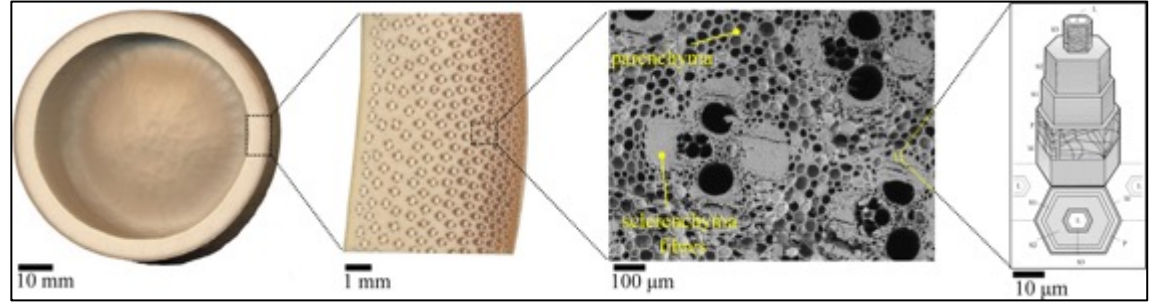
FIBRE EXTRACTION

FLAX

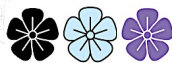


HEMP

BAMBOO

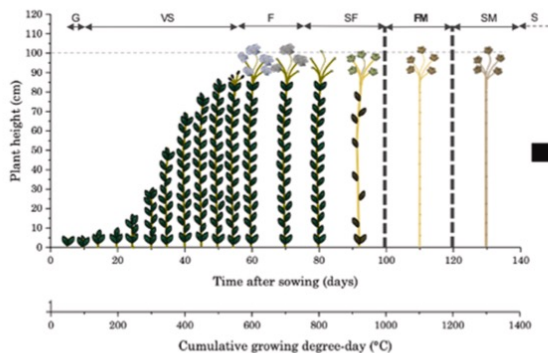


JUTE



FIBRE EXTRACTION

1. Growing stage



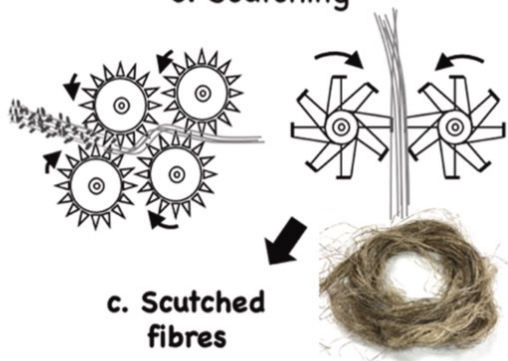
a. Green flax stems

2. Retting stage



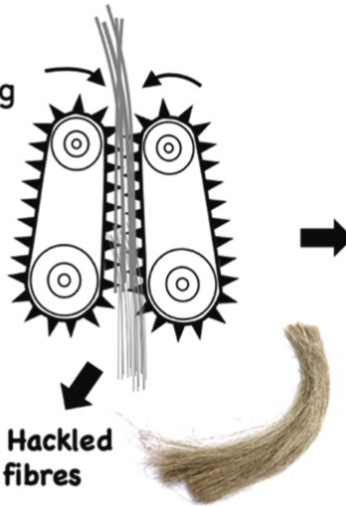
b. Retted flax stems

3. Scutching



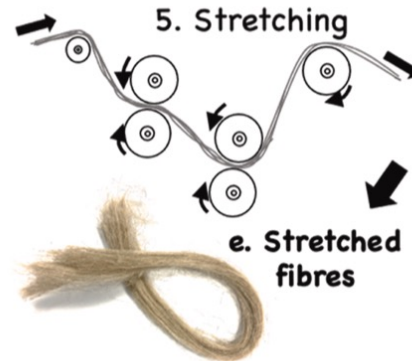
c. Scutched fibres

4. Hackling



d. Hackled fibres

5. Stretching



e. Stretched fibres



FIBRE EXTRACTION



Field Sowing Flowers



Pulling-out Retting



Fibre balls



Scutching



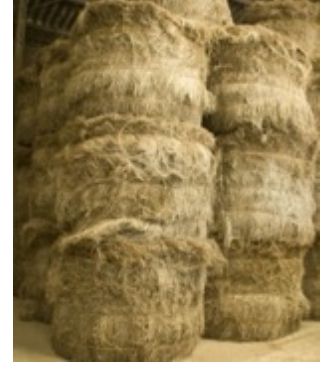
Scutched fibres

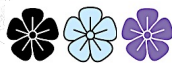


FOCUS ON RETTING



FOCUS ON RETTING – EXAMPLE OF HEMP





FOCUS ON RETTING – EXAMPLE OF HEMP

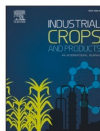


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Exploring the dew retting feasibility of hemp in very contrasting European environments: Influence on the tensile mechanical properties of fibres and composites

Samuel Rezuil^a, Brahim Mazian^{b,c}, Marie Grégoire^d, Salvatore Musio^e, Maxime Gautreau^f, Lucile Nuez^a, Arnaud Day^{g,h}, Pascal Thiébeauⁱ, Florian Philippe^j, Brigitte Chabbert^k, Anne Chamussy^l, Darshil U. Shah^k, Johnny Beaugrand^f, Vincent Placet^l, Jean-Charles Benezet^b, Antoine le Duigou^a, Mahadev Bar^d, Luc Malhautier^c, Emmanuel De Luycker^d, Stefano Amaducci^e, Christophe Baley^a, Anne Bergeret^b, Alain Bourmaud^a, Pierre Ouagne^{d,*}

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^b Polymers Composites and Hybrids (PCH), IMT Mines Ales, 6 Avenue de Clavieres, 30319, Ales Cedex, France

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^d Laboratoire Génie de Production, LGP, Université de Toulouse, INP-ENIT, Tarbes, France

^e Department of Sustainable Crop Production, Università Cattolica del Sacro Cuore, Piacenza, Italy

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^g Fibres Recherche Développement, Technopole de l'Aube en Champagne, Hotel de Bureaux 2 - 2 rue Gustave Eiffel, CS 90601-10901 Troyes Cedex 9, France

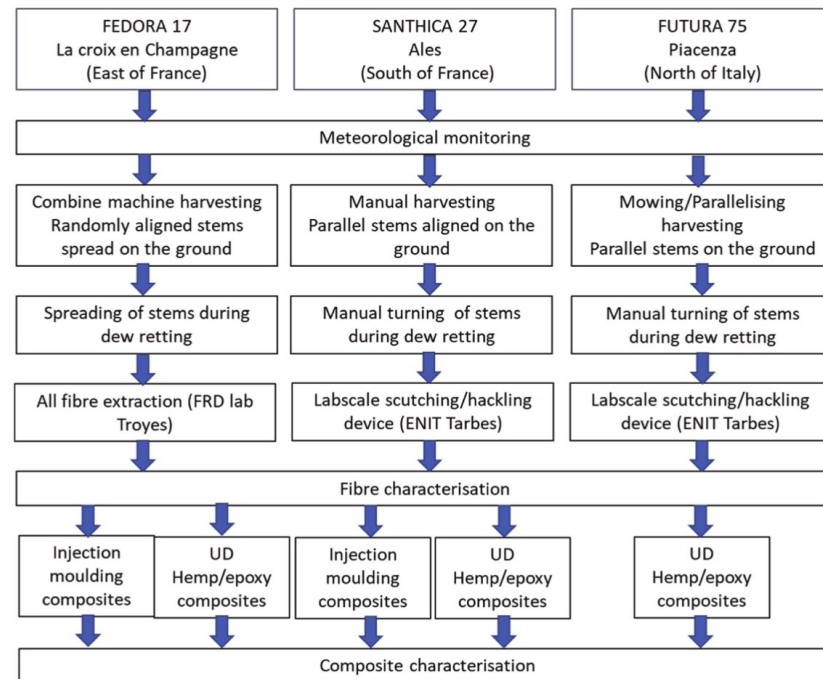
^h CNRS, UMR8576, UGSF- Unité de Glycobiologie Structurale et Fonctionnelle, Unité de Lille, 59000 Lille, France

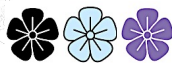
ⁱ Université de Reims Champagne Ardenne, INRAE, FARE, UMR A 614, 51097 Reims, France

^j La Chamvrière, Rue du Général de Gaulle, CS 20602, 10200, Bar-sur-Aube, France

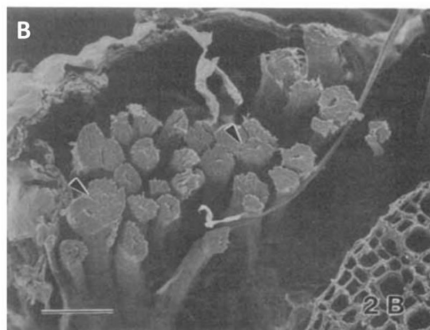
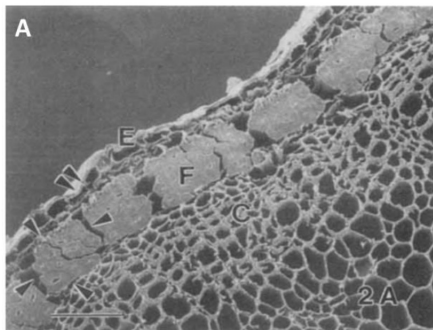
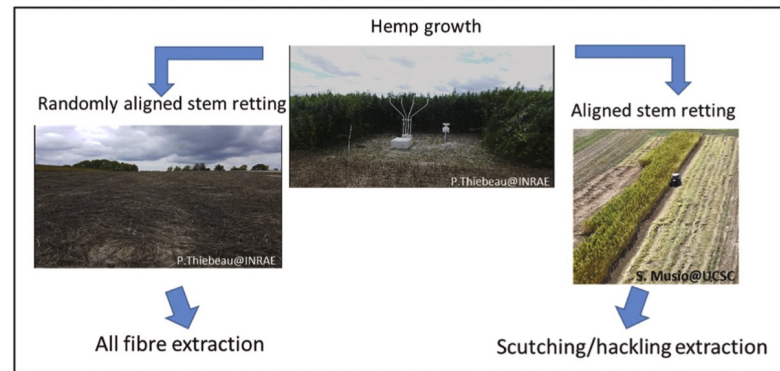
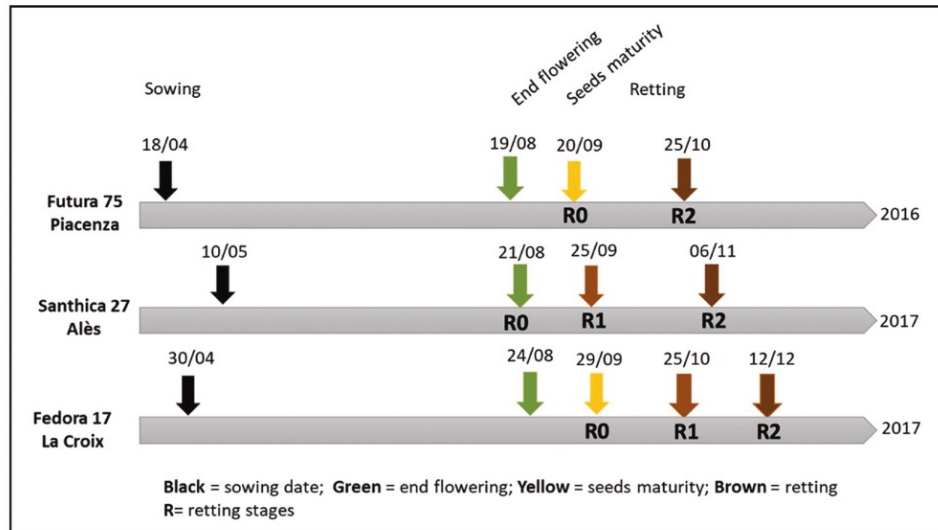
^k Centre for Natural Material Innovation, Dept. of Architecture, University of Cambridge, Cambridge CB2 1PX, United Kingdom

^l FEMTO-ST Institute, UFC/CNRS/ENSMM/UTBM, Université Bourgogne Franche-Comté, Besançon, France

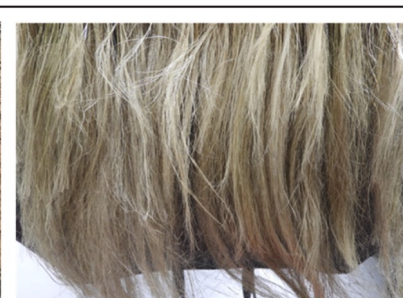




FOCUS ON RETTING – EXAMPLE OF HEMP



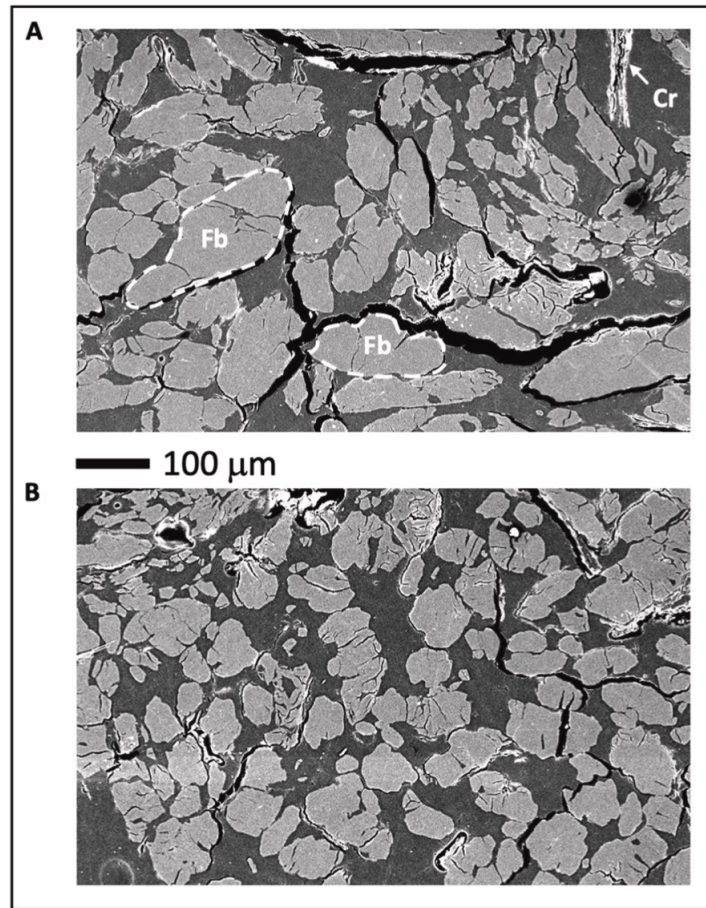
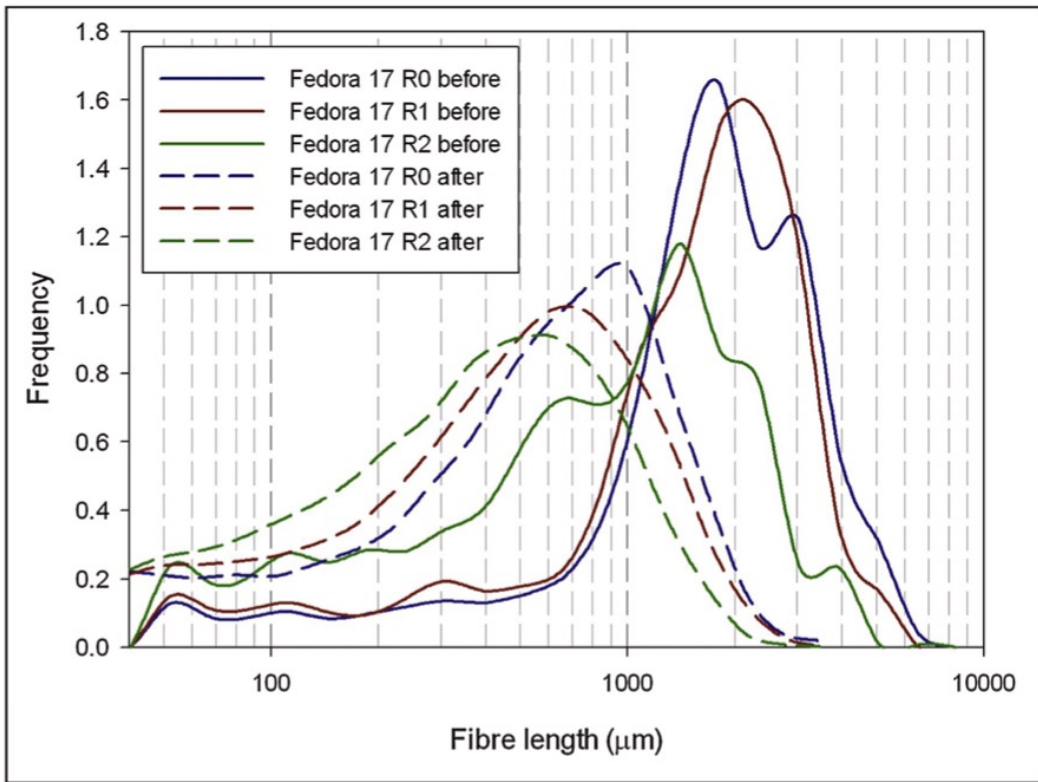
(a) Hemp extracted fibres at the Output of FRD line

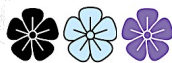


(b) Hemp extracted fibres at the output of the lab-scale scutching/hackling line

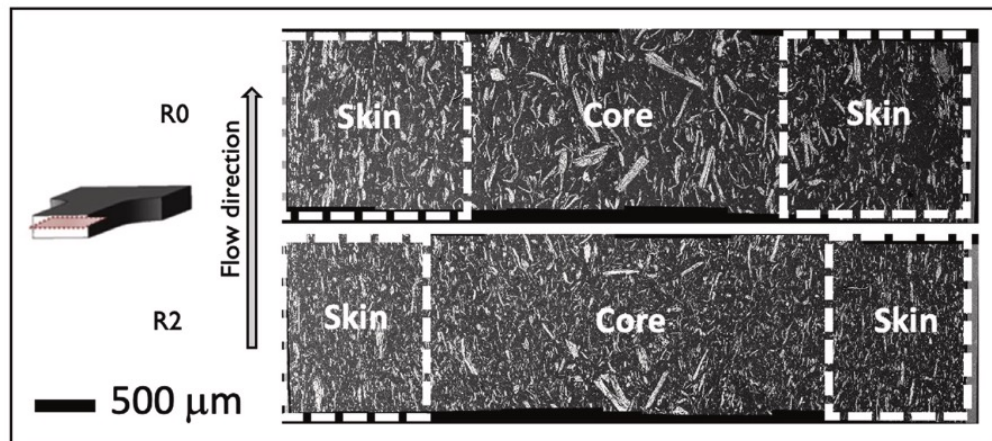
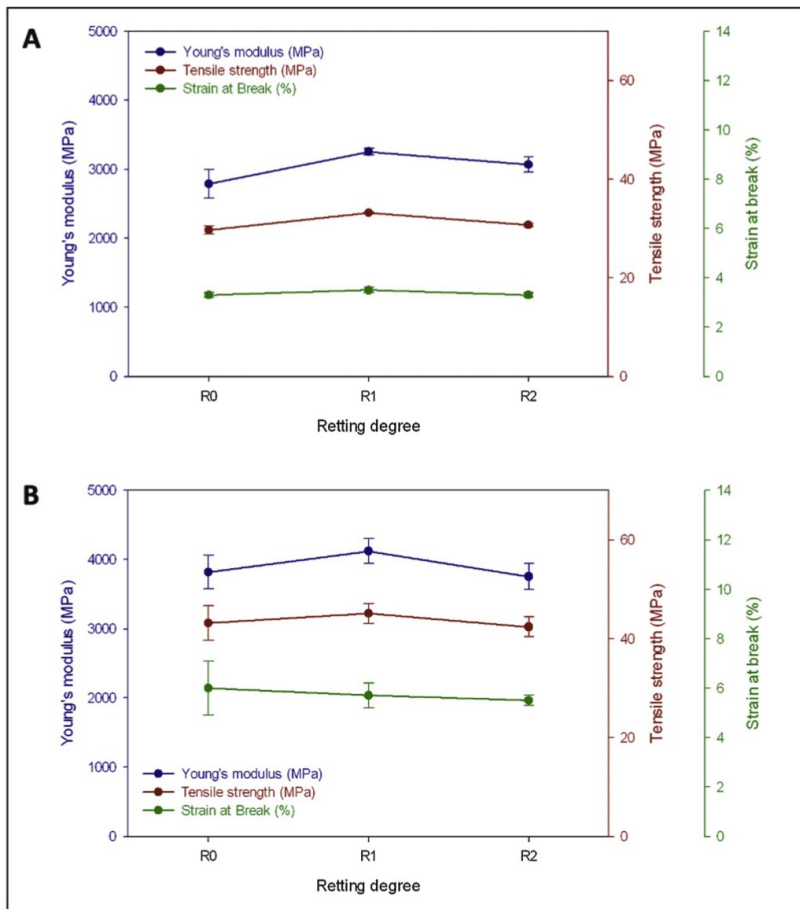


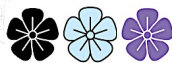
FOCUS ON RETTING – EXAMPLE OF HEMP





FOCUS ON RETTING – EXAMPLE OF HEMP



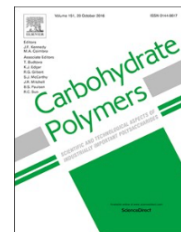


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Evolution of flax cell wall ultrastructure and mechanical properties during the retting step



Alain Bourmaud^{a,*}, David Siniscalco^a, Loïc Foucat^b, Camille Goudenhoft^a, Xavier Falourd^b, Bruno Pontoire^b, Olivier Arnould^c, Johnny Beaugrand^b, Christophe Baley^a

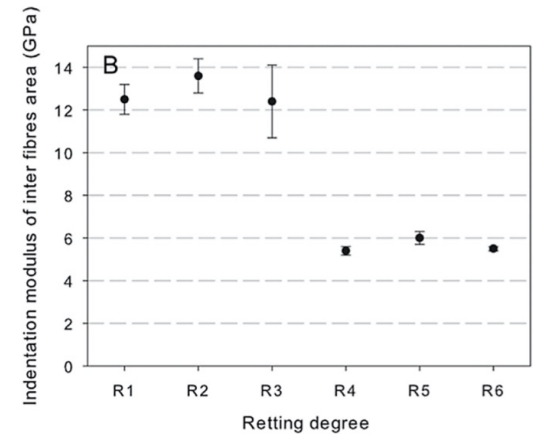
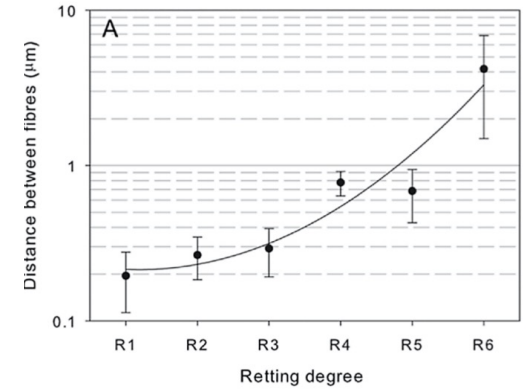
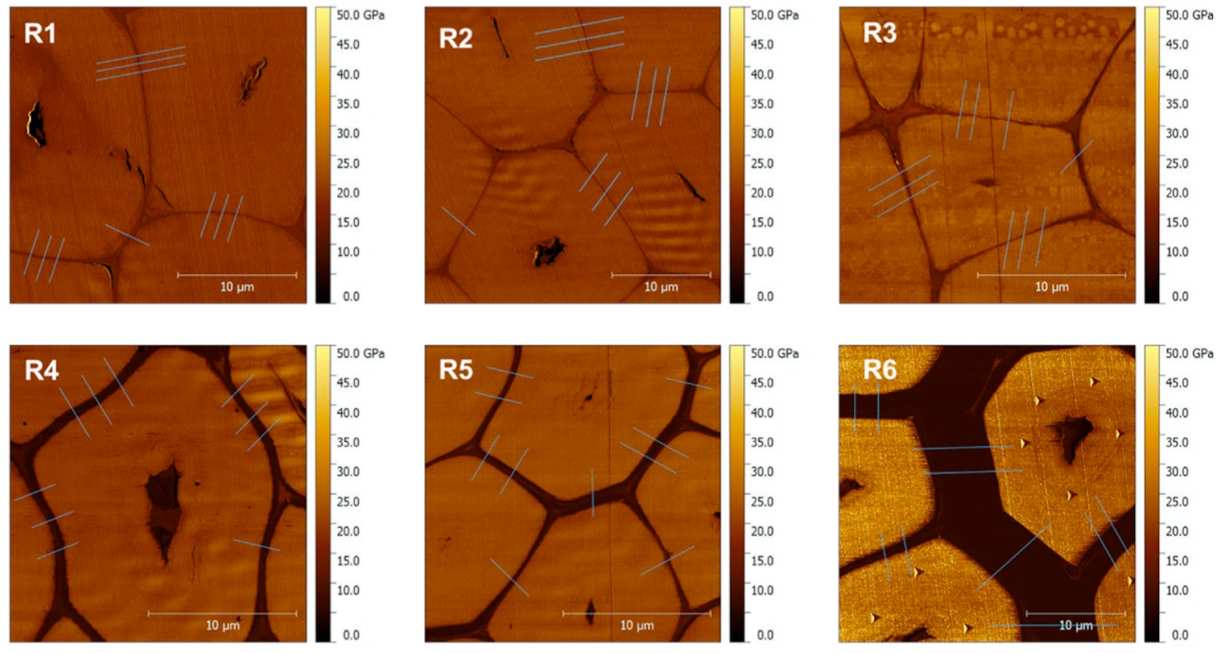
^a IRDL, Université Européenne Bretagne, CNRS, UMR 6027, Lorient, France

^b UR1268 Biopolymères Interactions Assemblages, INRA, Nantes, France

^c LMGC, Université de Montpellier, CNRS, UMR 5508, Montpellier, France

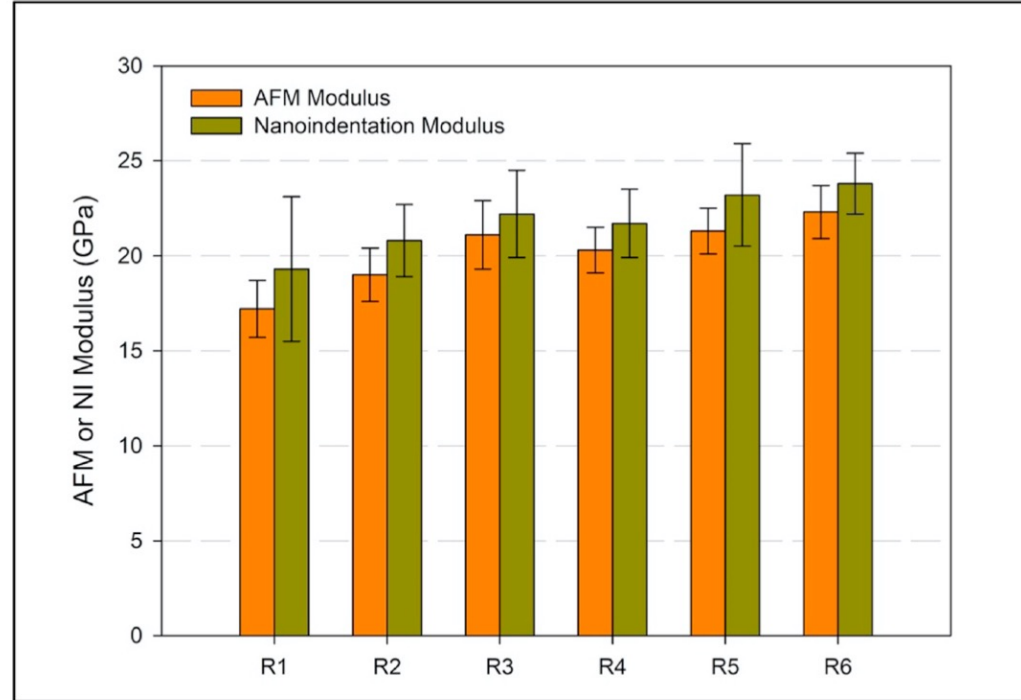
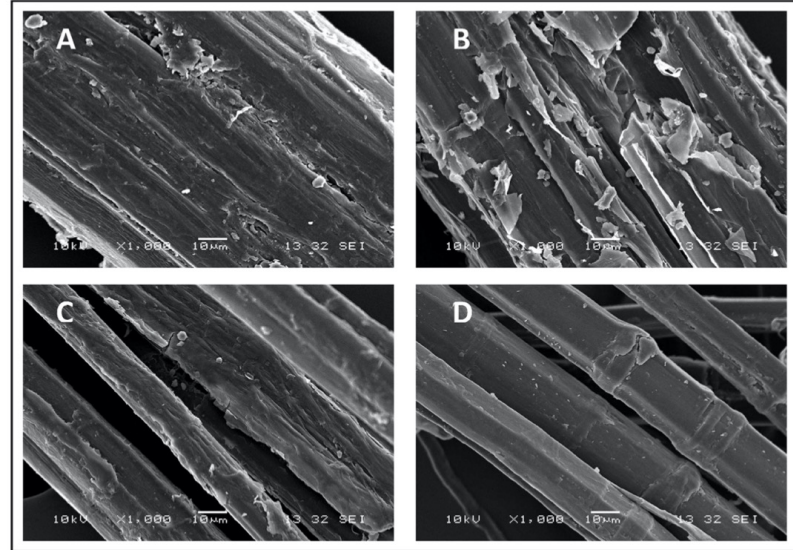


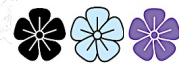
FOCUS ON RETTING – EXAMPLE OF FLAX



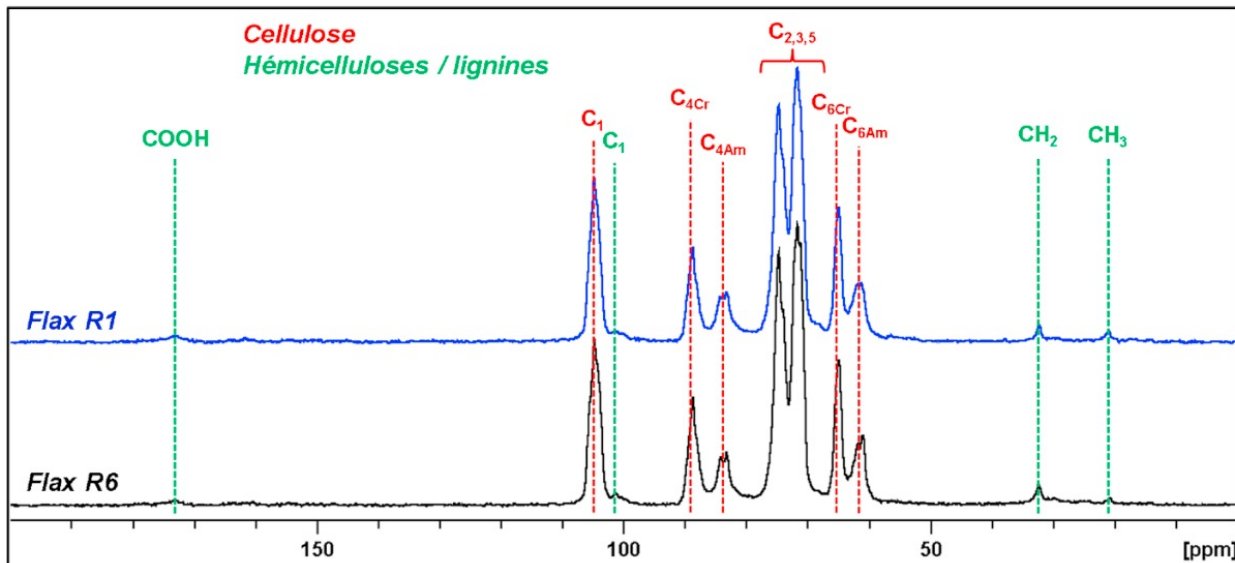


FOCUS ON RETTING – EXAMPLE OF FLAX



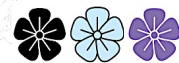


FOCUS ON RETTING – EXAMPLE OF FLAX

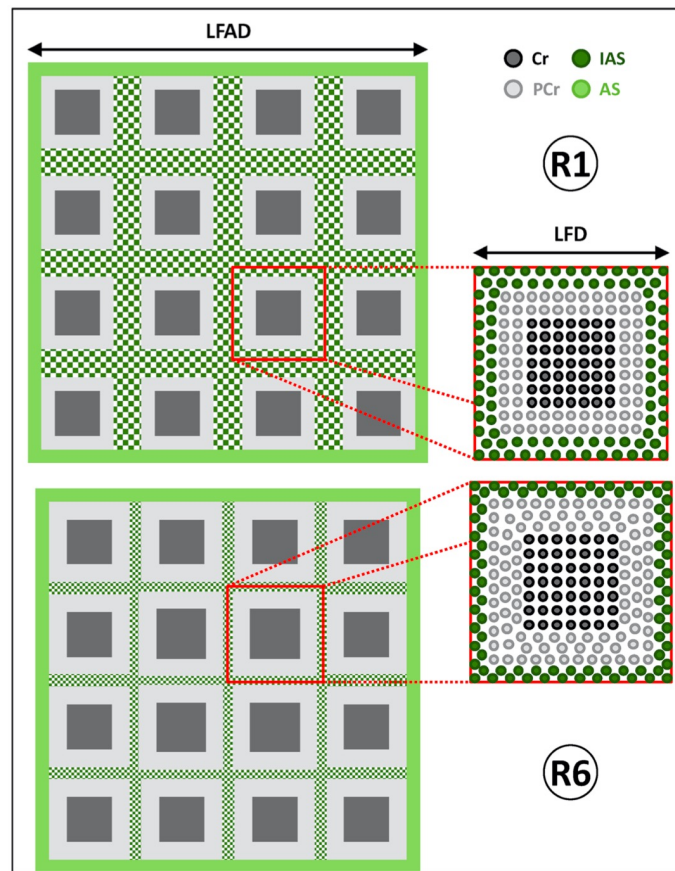
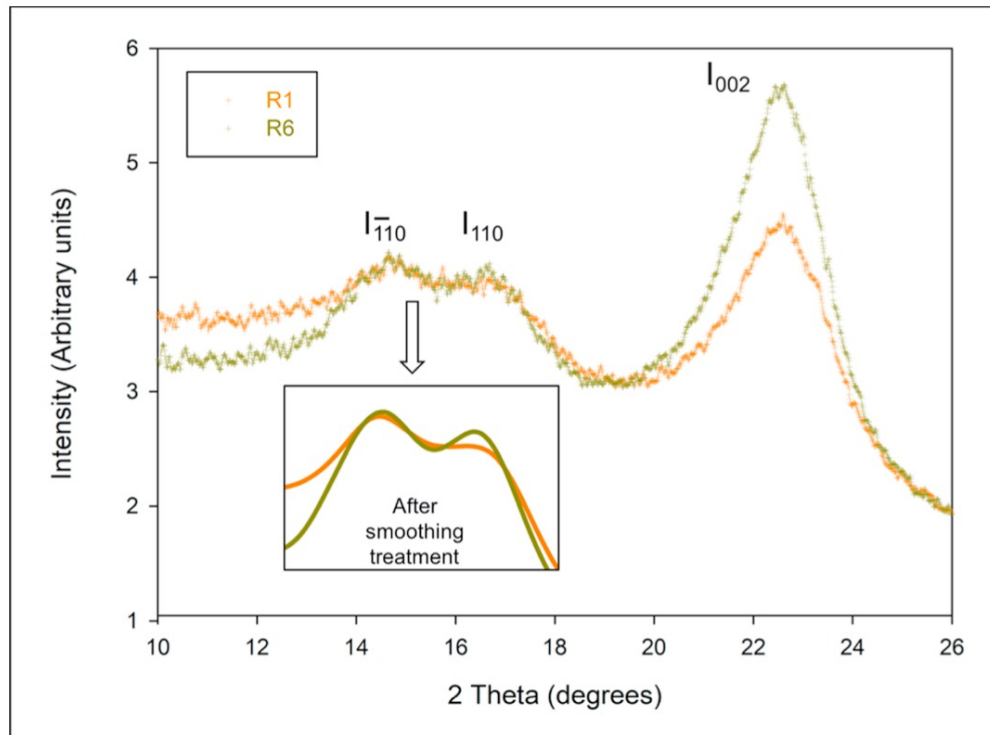


Peak assignment

	Cr(Iα)	Cr(Iα + β)	PCr	Cr(Iβ)	AS	IAS	AS	
R1	89.37	88.73	88.58	87.83	84.22	83.53	83.16	δ (ppm)
R6	89.38	88.70	88.54	87.68	84.14	83.68	83.16	
R1	81	60	199	90	98	482	86	FWHH (Hz)
R6	76	55	197	90	88	484	76	
R1	8	10	32	2	6	35	7	Normalized area (%)
R6	7	10	37	2	7	31	7	

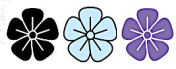


FOCUS ON RETTING – EXAMPLE OF FLAX



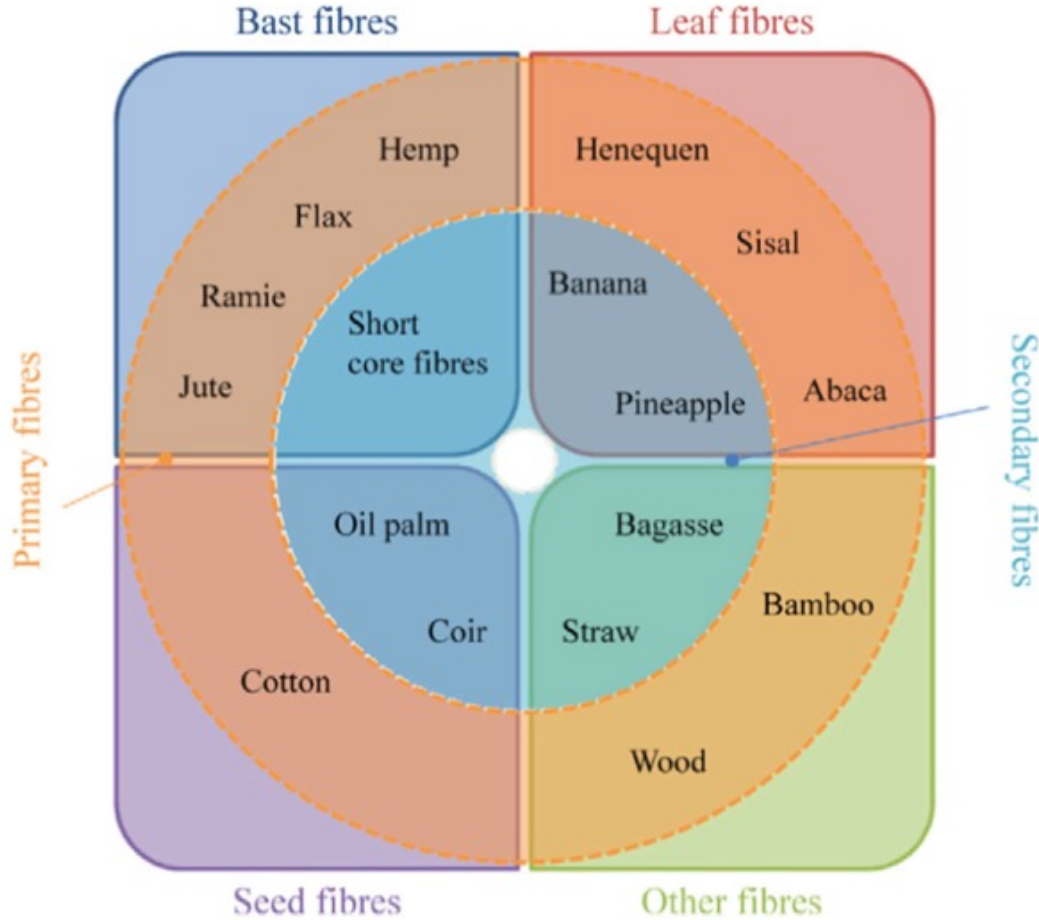


A DIVERSITY OF PLANT FIBRES



DIVERSITY OF PLANT FIBRES

Specific functions according to the fibre location within the plant





DIVERSITY OF PLANT FIBRES

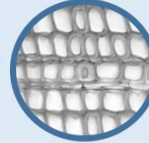
Plant fibres

Within xylem

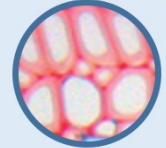
Tension wood
gelatinous cell walls
(*Castanea sativa*)



Wood tracheids
(*Pinus sylvestris*)



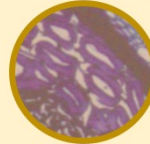
Wood Libriform
fibres (*Alchornea
lojaensis*)



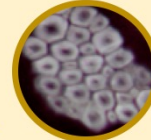
Phloem of dycotyledon plant

Procambium
primary fibres

Nettle
(*Urtica dioica*)



Flax
(*Linum usitatissimum L*)



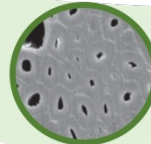
Hemp
(*Cannabis sativa*)
(primary fibres)

Outside of xylem

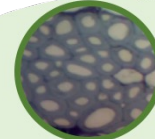
Periphery of vascular bundles

Vascular cambium
secondary fibres

Jute
(*Corchorus
olitorus*)



Kenaf (*Hibicus
cannabinus*)

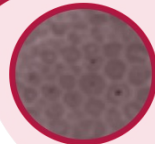


Hemp (*Cannabis sativa*)
(secondary fibres)

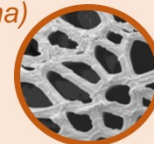


Culms

Bamboo
(*Guada
angustifolia*)



Sisal (*Agave
sisalana*)



Abaca
(*Musa textilis*)



Leaves of monocotyledon

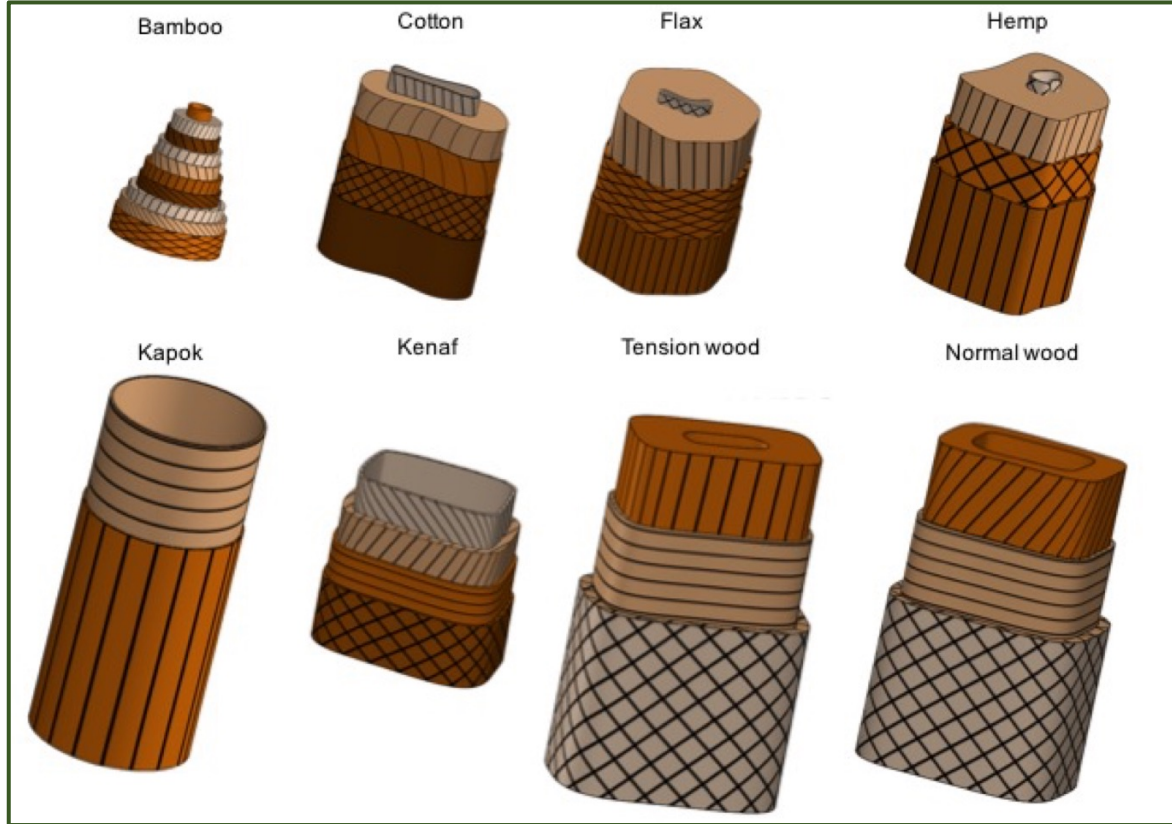
Mesocarp

Coir
(*Cocos nucifera*)





DIVERSITY OF PLANT FIBRES



Different morphologies

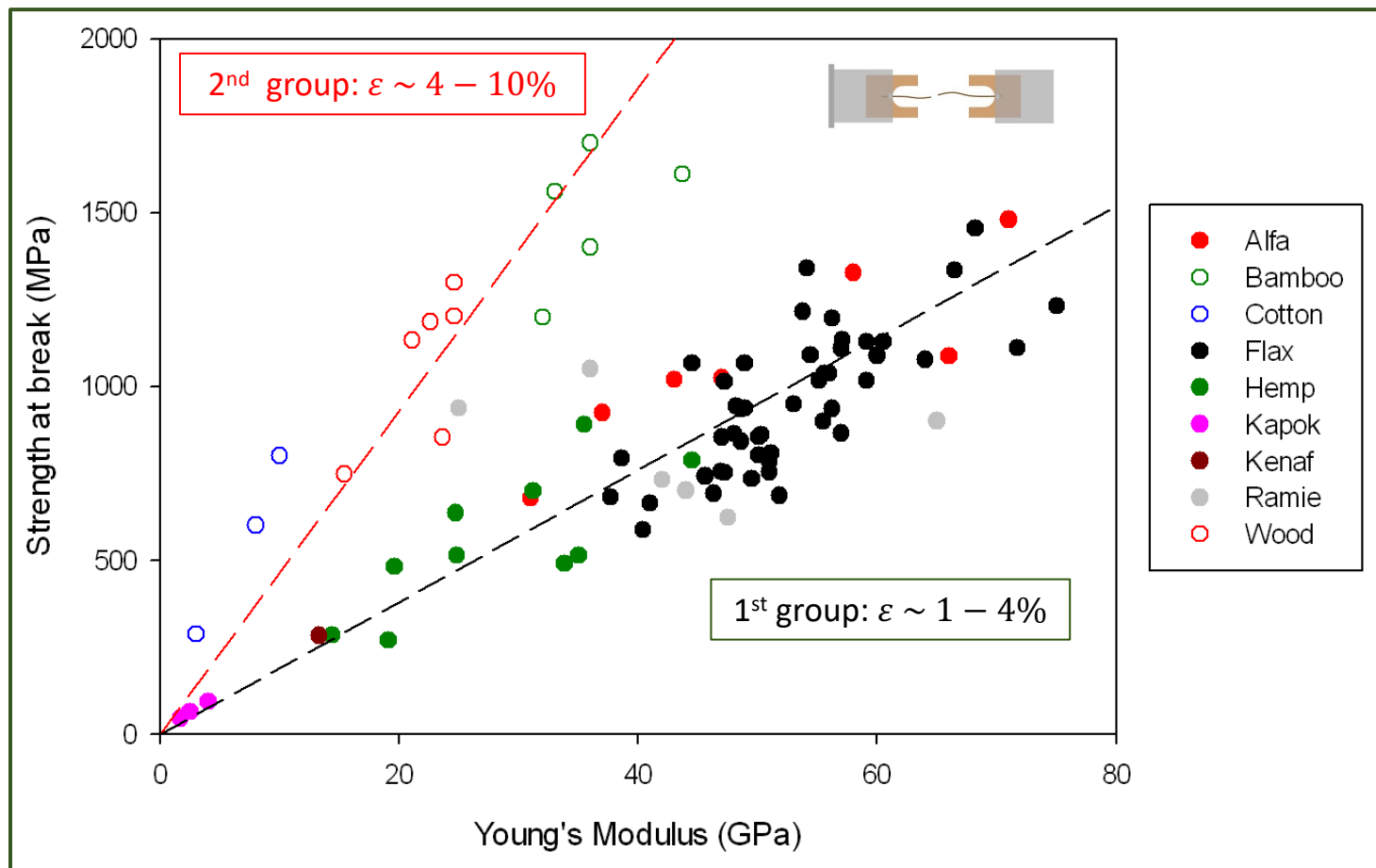
In link with function and cell wall development

Differences in length or stiffness

Filling rate, lumen size

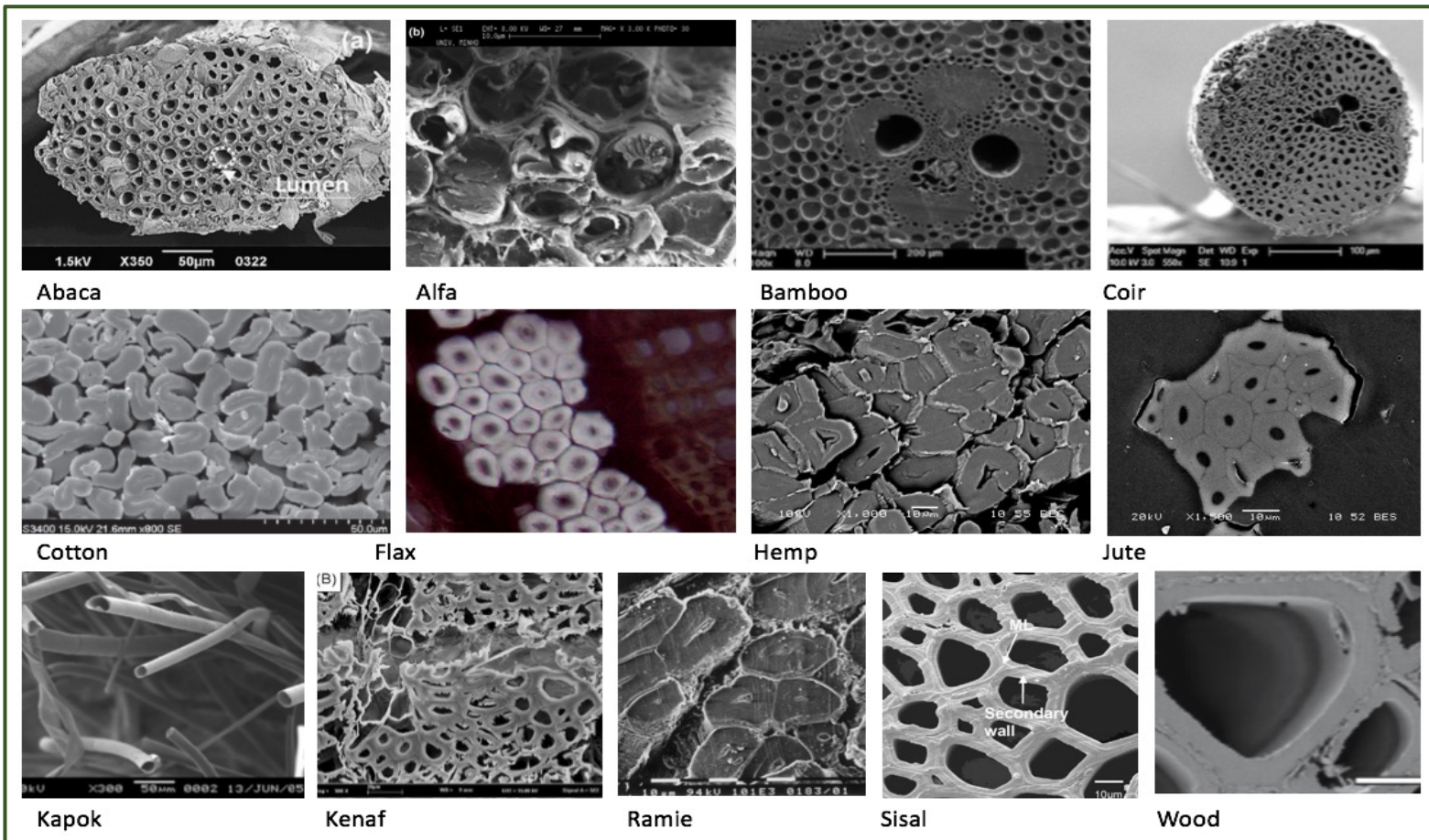


DIVERSITY OF PLANT FIBRES



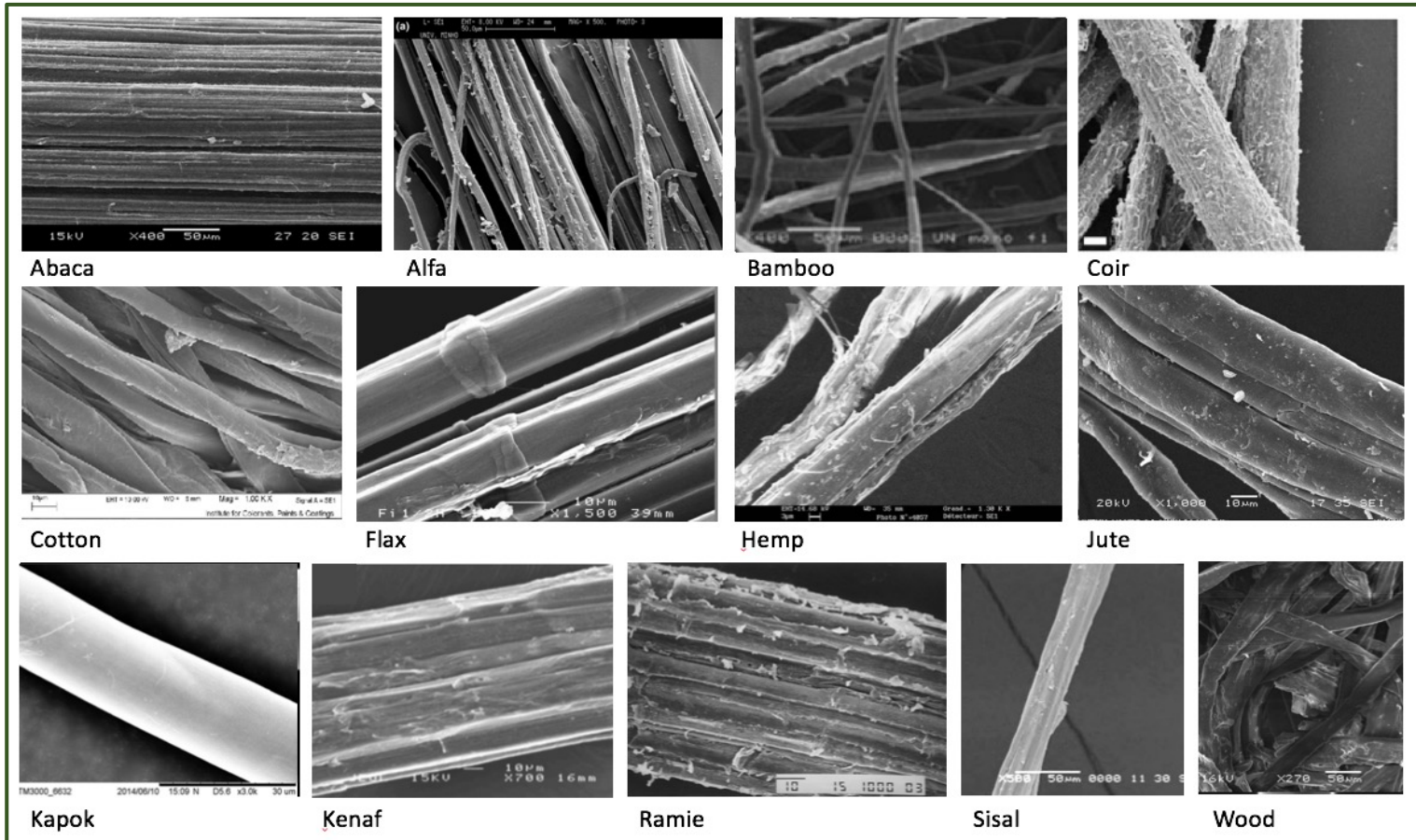


DIVERSITY OF PLANT FIBRES



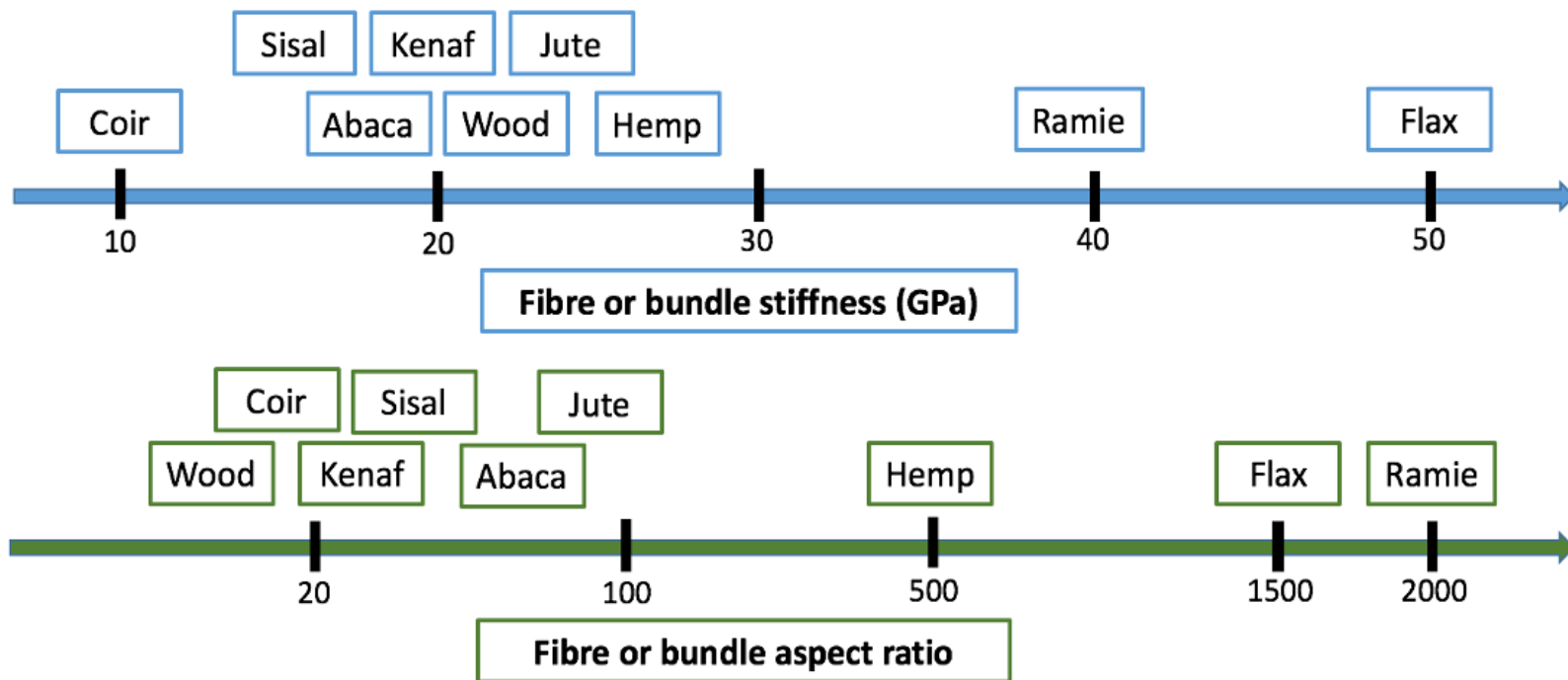


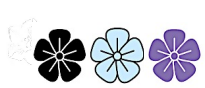
DIVERSITY OF PLANT FIBRES





DIVERSITY OF PLANT FIBRES



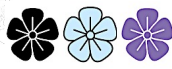


DIVERSITY OF PLANT FIBRES

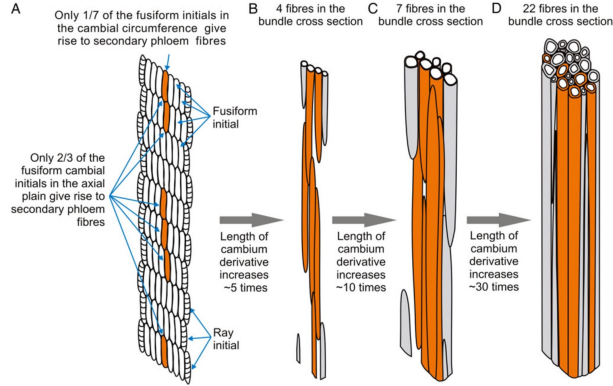
- Plant fibres are all different
- One plant fibre = specific intrinsic properties and potential different application
- One is not necessary better than another
- For their use, consider origin (LCA), competition with food, technical interest....
- Price and available volumes are also key-points



PLANT FIBRES DEVELOPMENT AND ULTRASTRUCTURE

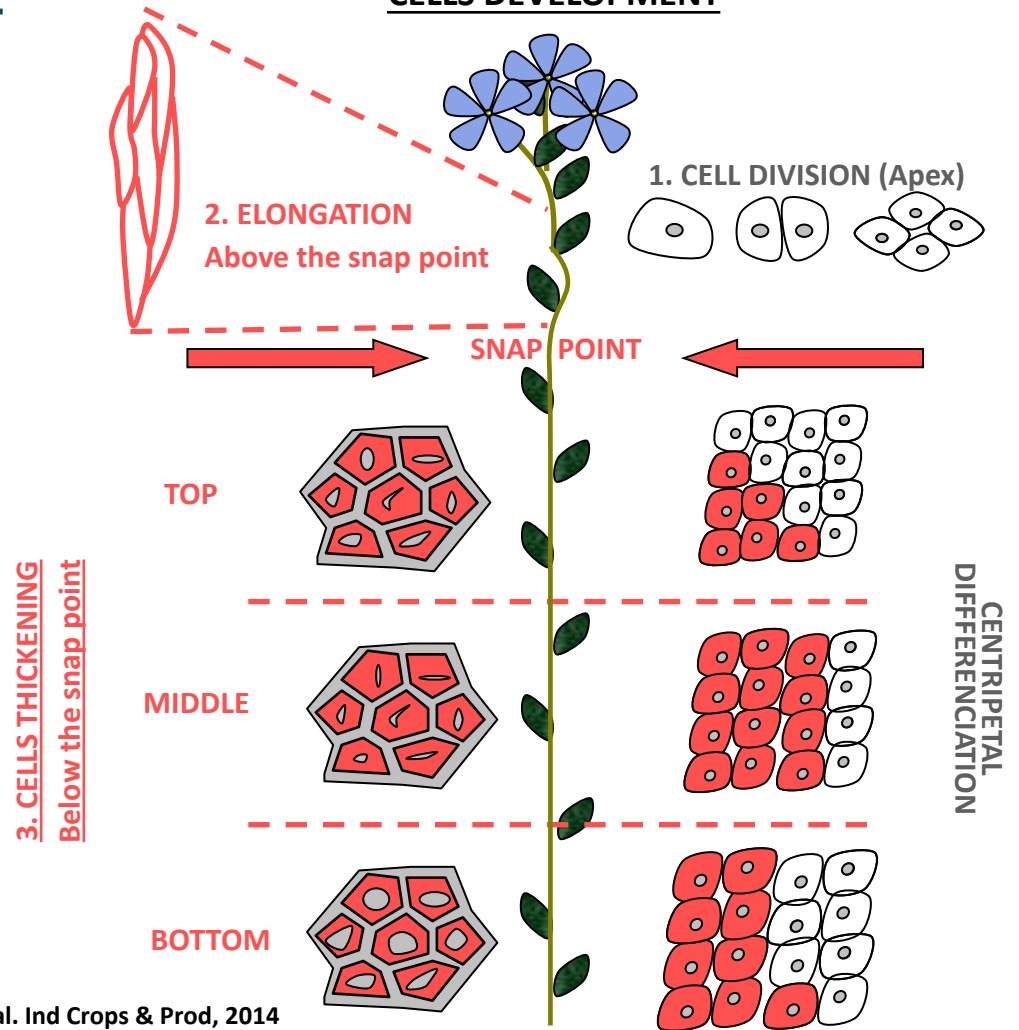


CELL WALL DEVELOPMENT

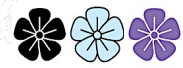


Snegireva 2015

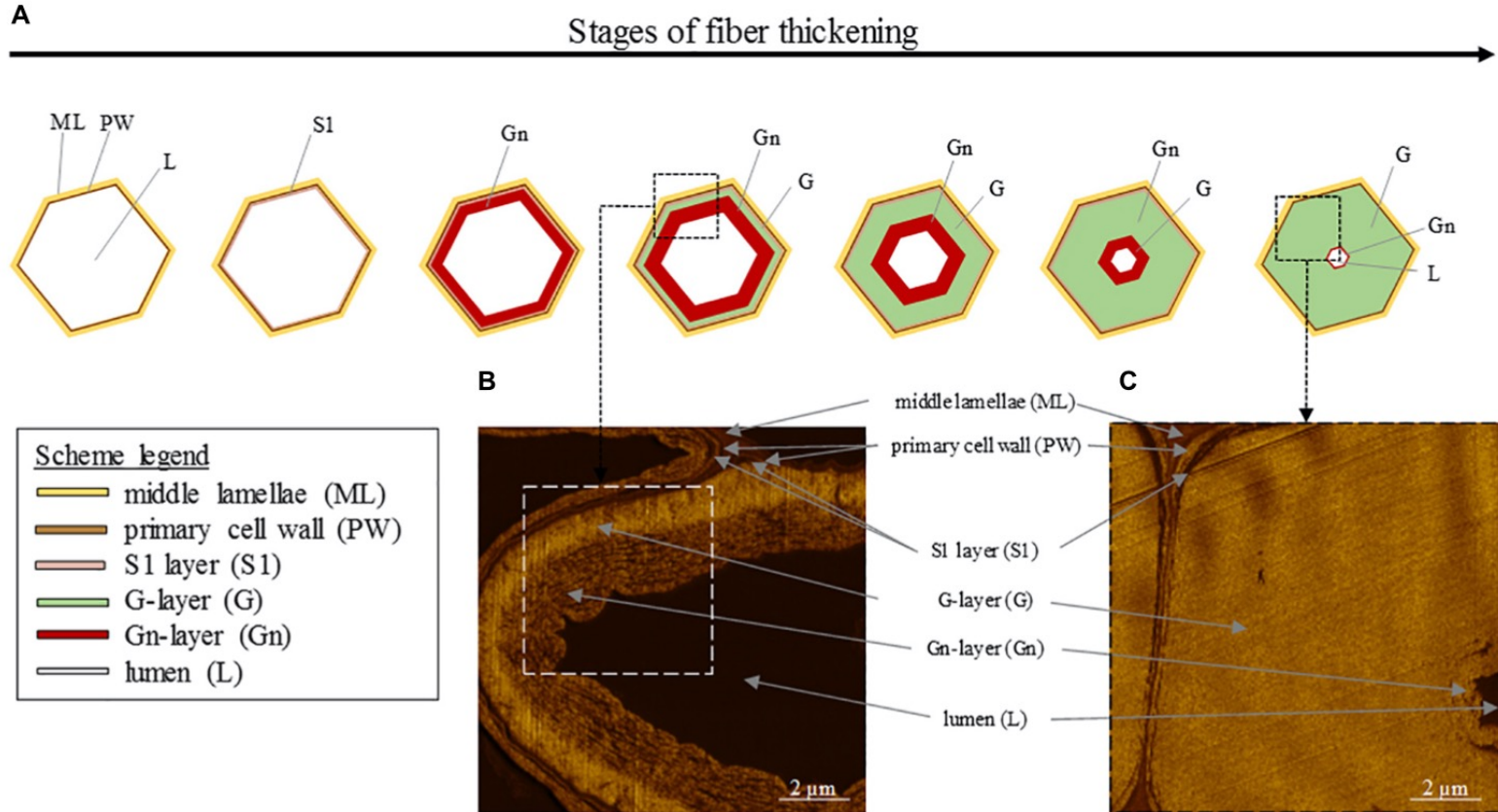
CELLS DEVELOPMENT



Bourmaud et al. Ind Crops & Prod, 2014

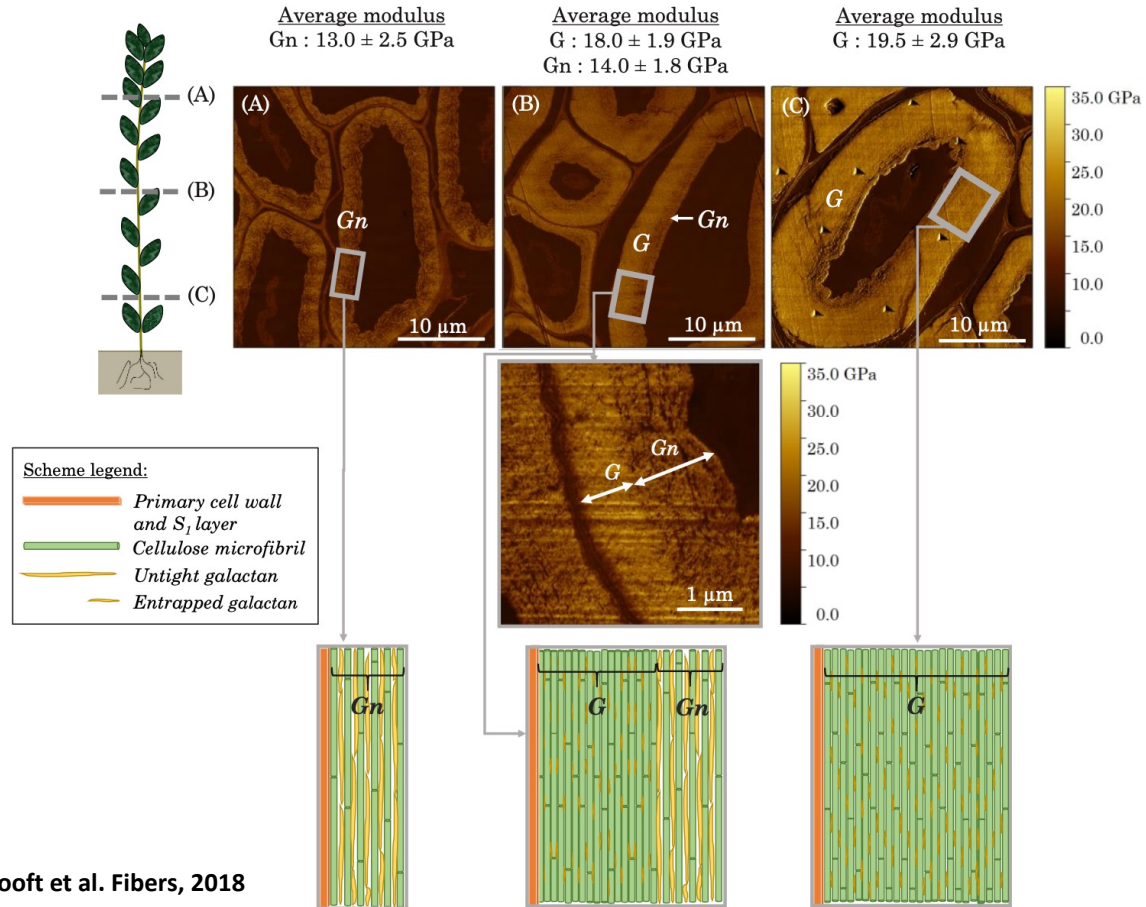


DEVELOPEMENT AND STRUCTURE OF FIBRES



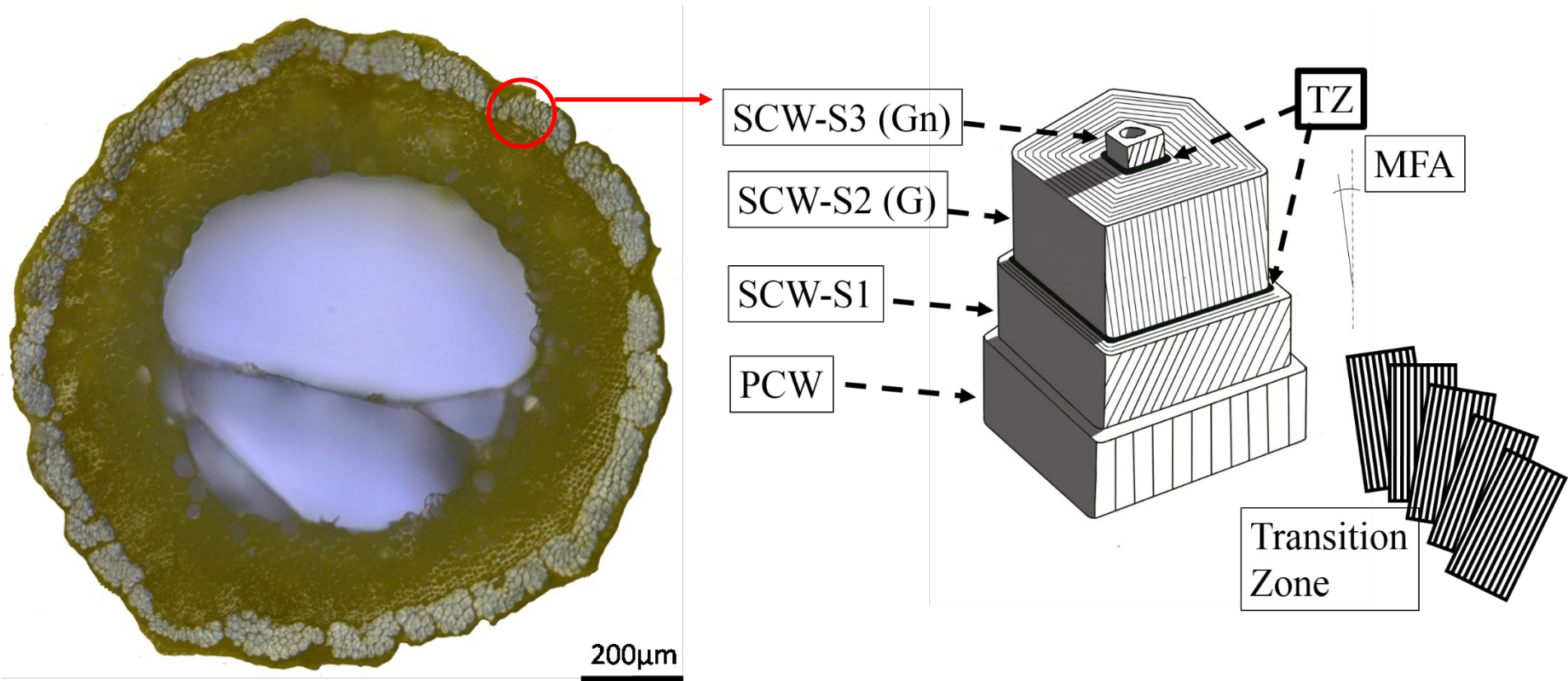


DEVELOPEMENT AND STRUCTURE OF FIBRES



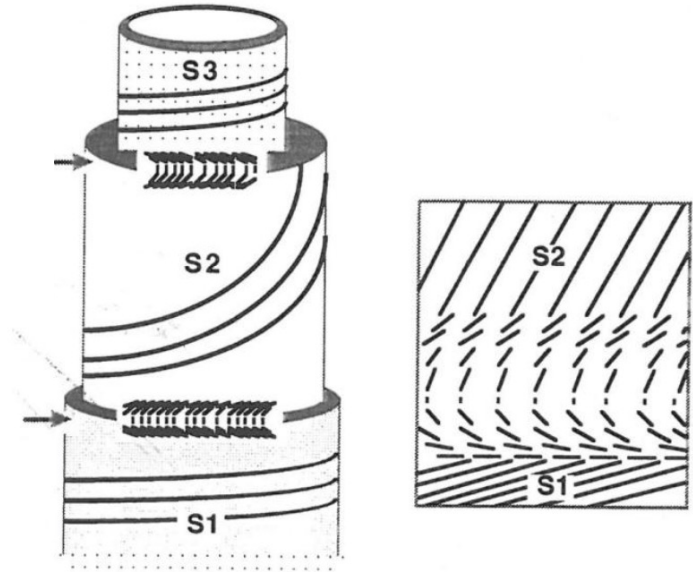
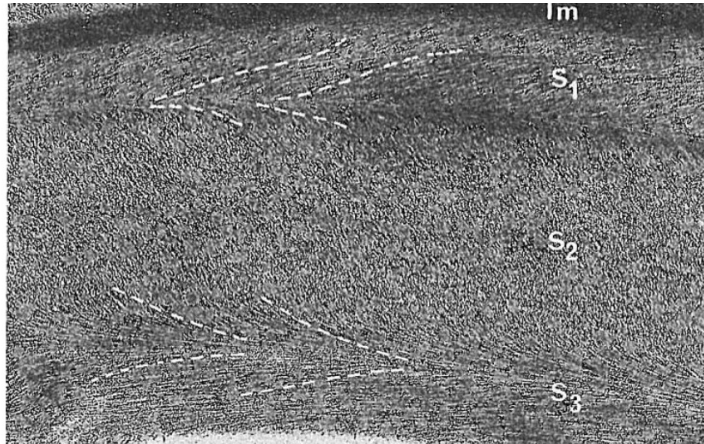
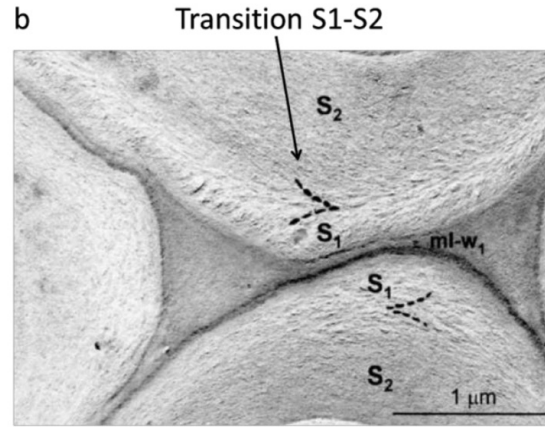
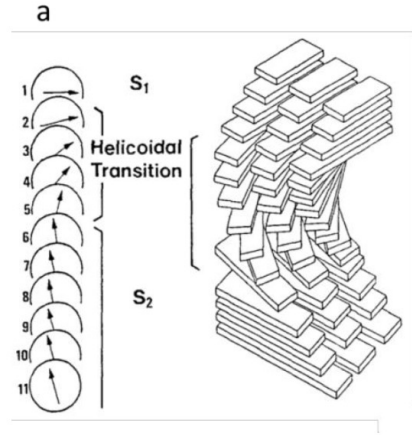


DEVELOPMENT AND STRUCTURE OF FIBRES

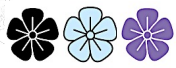




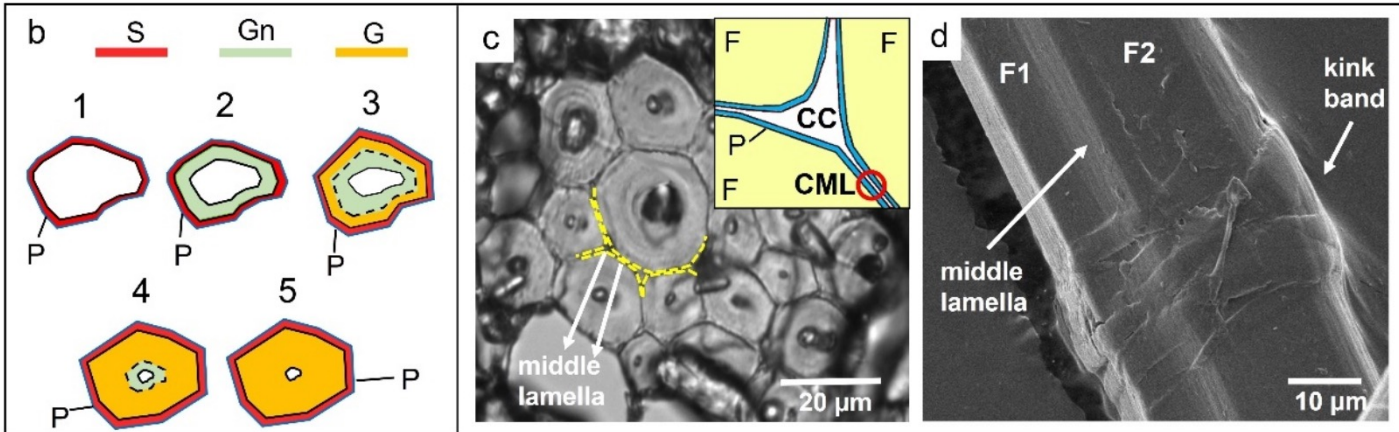
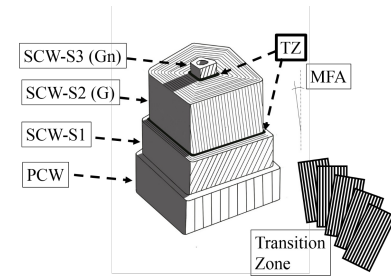
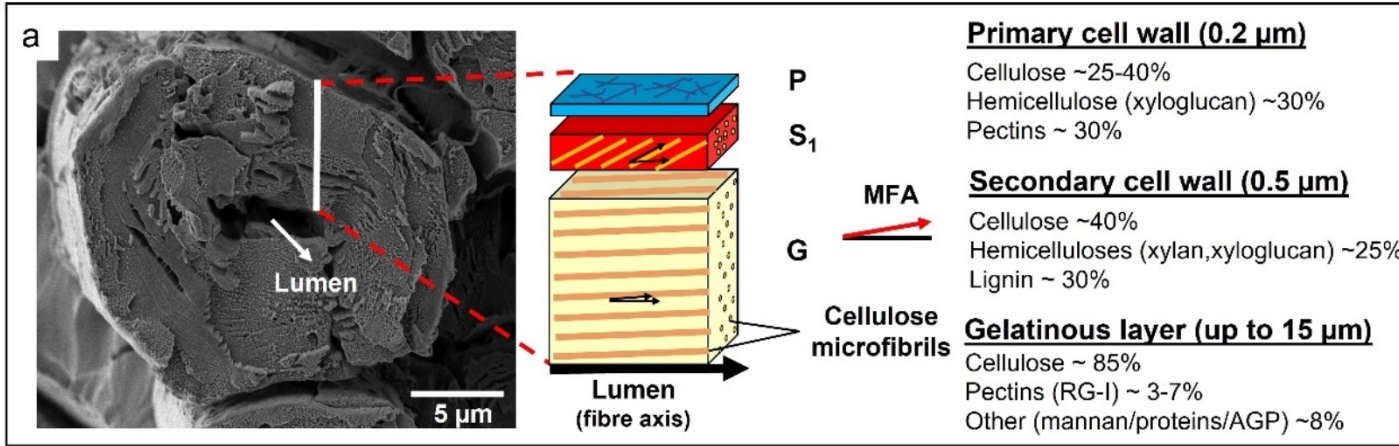
DEVELOPMENT AND STRUCTURE OF FIBRES



Roland et al. 1995



DEVELOPMENT AND STRUCTURE OF FIBRES

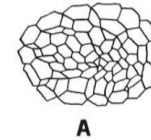
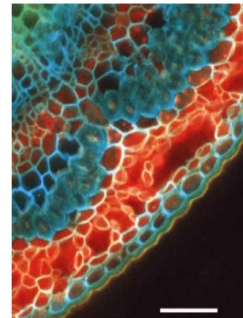
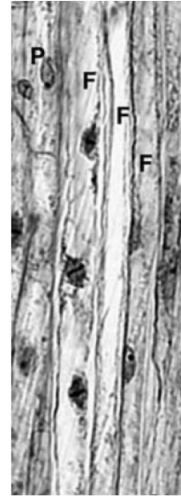


Baley et al. Bioinsp & Biom, 2018

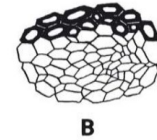


PLANT FIBRES: KEY POINTS

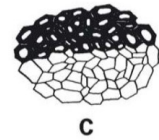
- Flax fibres are long and poly nucleated cells
- From 15 to 80 mm !
- Filled with cellulose (Approx 80%)
- Highly crystalline
- A porosity inside: 0-5%



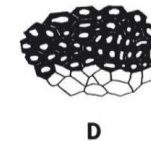
A



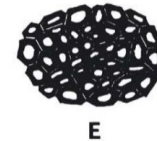
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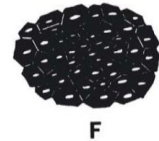
C



D



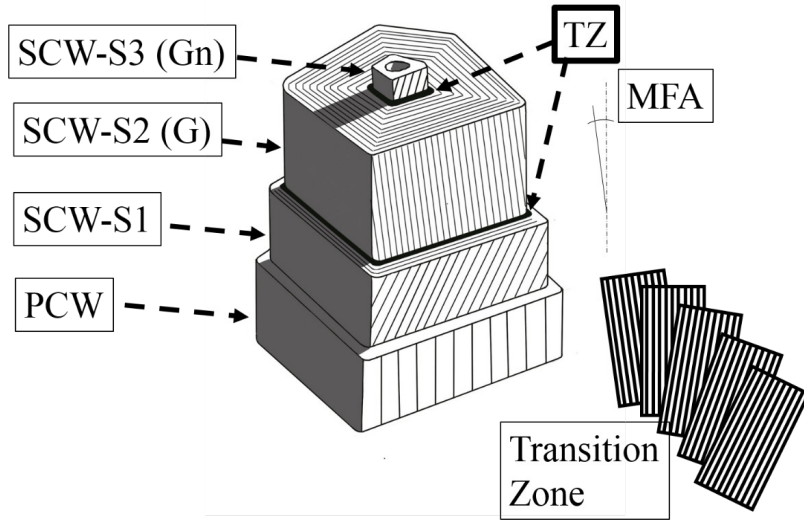
E



F



BIOCHEMICAL ARCHITECTURE OF PLANT CELL WALLS



Cell wall layer	Average thickness	Microfibrils orientation	Approximate composition
PW	0.2 μm [87]	disperse orientation, preferentially 0° [70,72]	~25-40% cellulose ~30% hemicelluloses (mainly xyloglucan and lesser amounts of arabinoxylan) ~30% pectins (mainly homogalacturonan; possibly rhamnogalacturonan (RG) I; RG II and arabinogalactan) [53,70,83,88-90]
S1	0.5 μm [73,91]	$60-80^\circ$ [92]	~30-50% cellulose ~30% hemicelluloses (xylan, xyloglucan) ~5% pectins (homogalacturonan and RG I) ~10-20% lignin [70,72,78,89]
G	up to 15 μm or 90% of the total cell wall area at maturity [71,83]	$8-10^\circ$ [91,93]	~75-90% cellulose ~15-20% hemicelluloses (glucomanan) ~5-10% pectins (RG I) [53,71,72,83,89]
Gn	0.5-1 μm through thickening [79,91]	loosely packed as an heterogeneous structure [94]	cellulose hemicelluloses (glucomanan) pectins (nascent RG I (i.e. long galactan chains)) [71,72,78,94]



BIOCHEMICAL ARCHITECTURE OF PLANT CELL WALLS

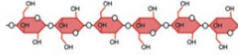
Cellulose / Hemicelluloses

Cellulose

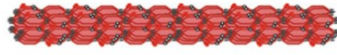
Glucose



Cellulose



Cellulose microfibril



Hemicelluloses

Glucose



Xylose



Galactose



Arabinose



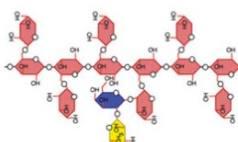
Mannose



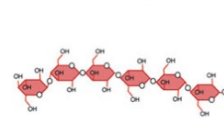
Fucose



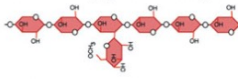
Fucoside xyloglucan



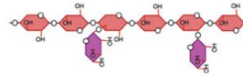
Mixed-linkage glucan



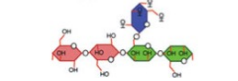
4-O-methylglucuronoxylan



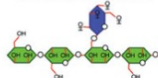
Arabinoxylan



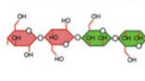
Glucogalactomannan



Galactomannan



Glucomannan



Pectins

Galacturonic acid



Rhamnose



Uronic acids

Glucuronic acid

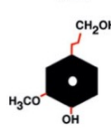


Mannuronic acid

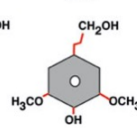


Lignins

Guaiacyl unit

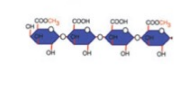


Syringyl unit

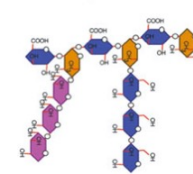


Pectins, UA & Lignins

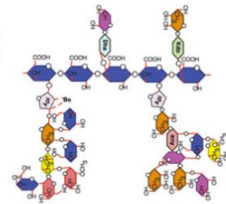
Homogalacturonan



Rhamnogalacturonan I

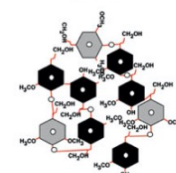


Rhamnogalacturonan II

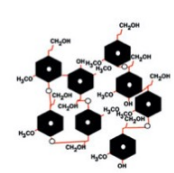


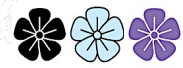
Unidentified polymers

Lignin GS

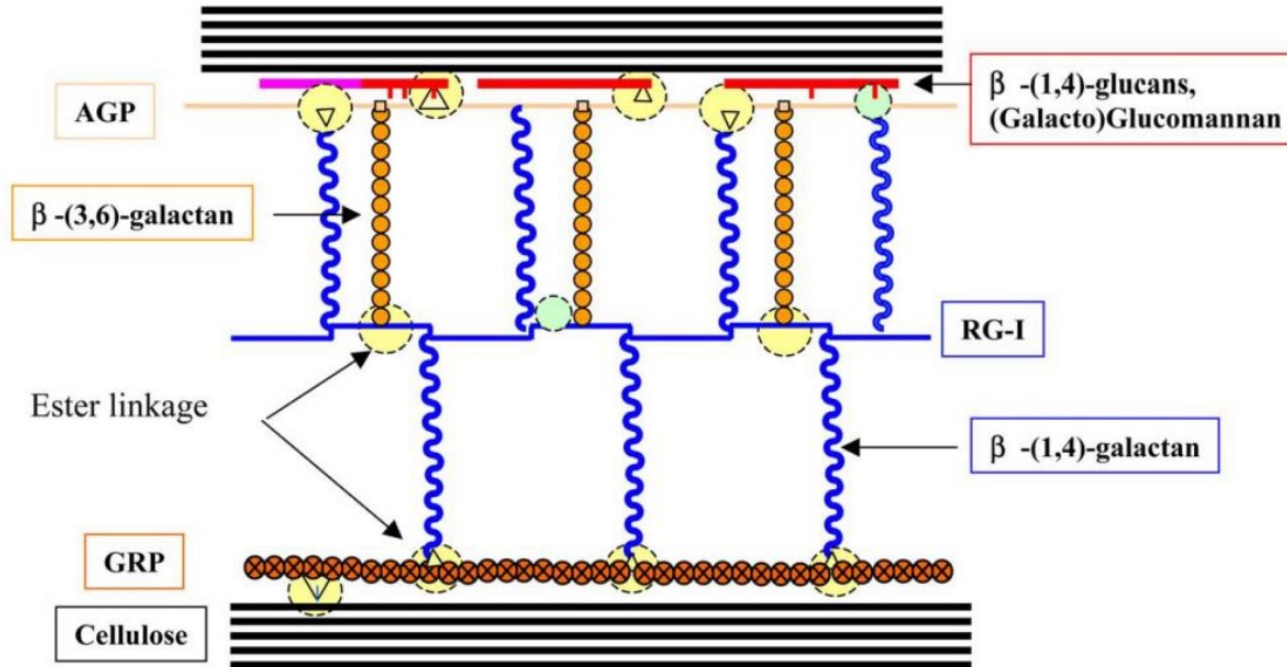


Lignin G



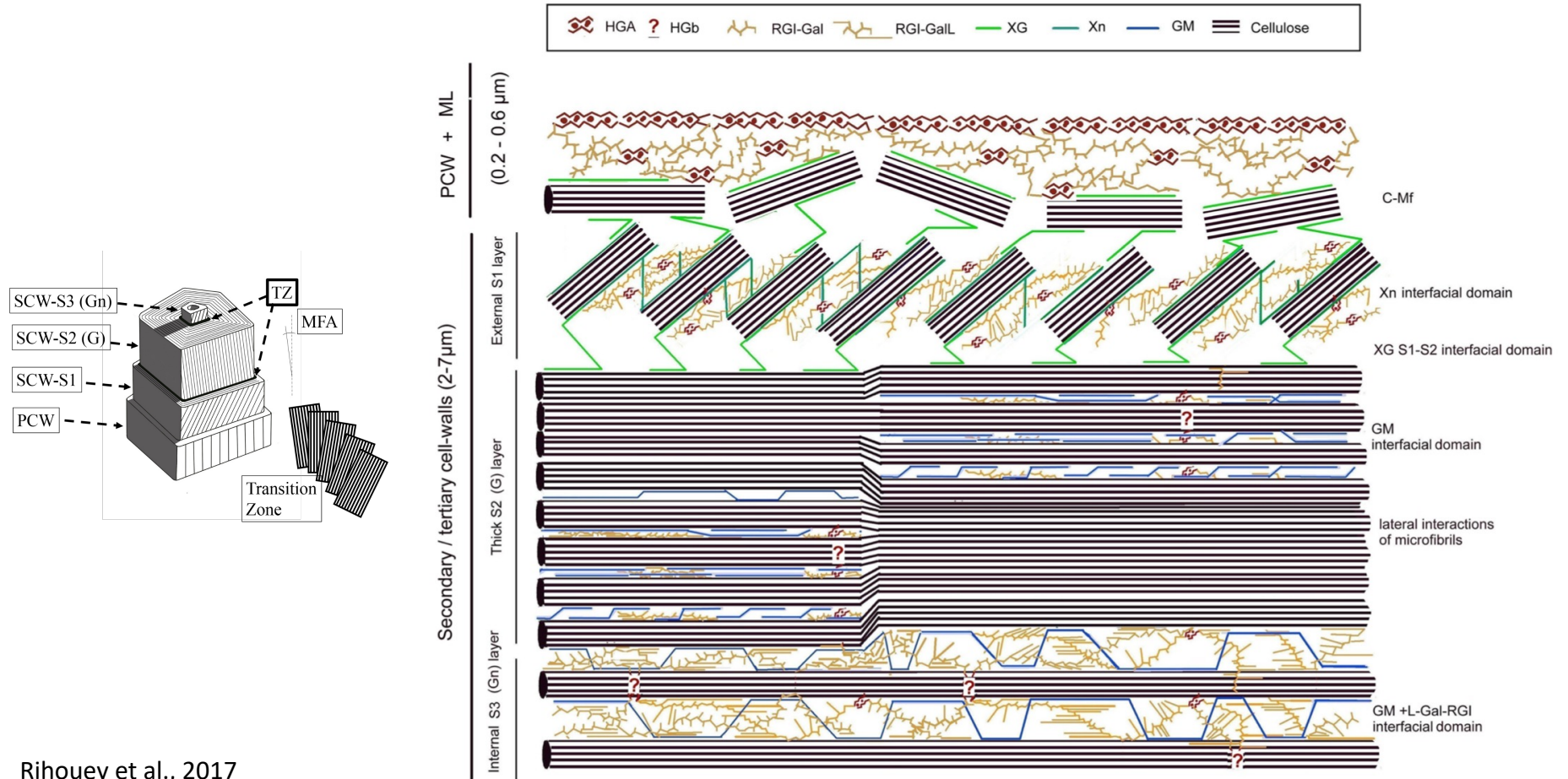


BIOCHEMICAL ARCHITECTURE OF PLANT CELL WALLS



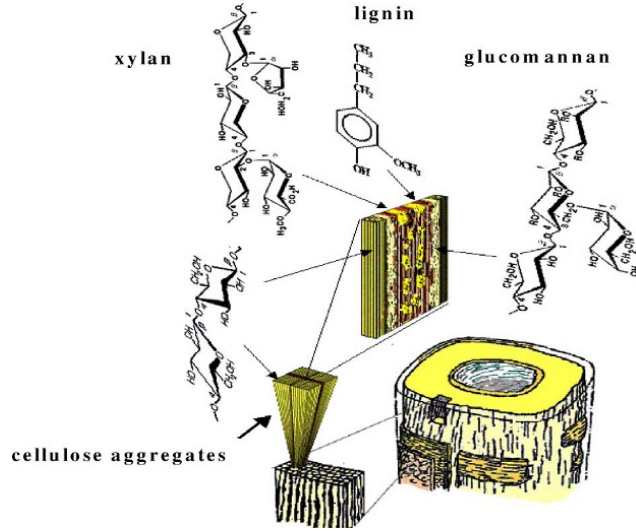


BIOCHEMICAL ARCHITECTURE OF PLANT CELL WALLS

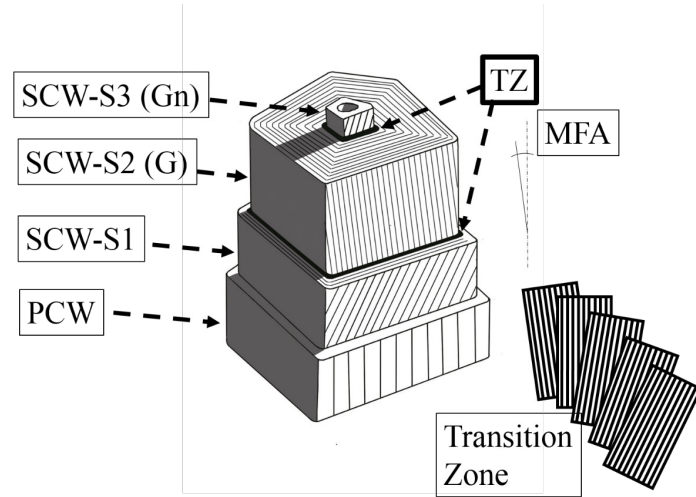




FLAX VS WOOD



Wood cell wall layer ultrastructure [Salmén, *C.R. Biologies*, 2004]



Wood S₂

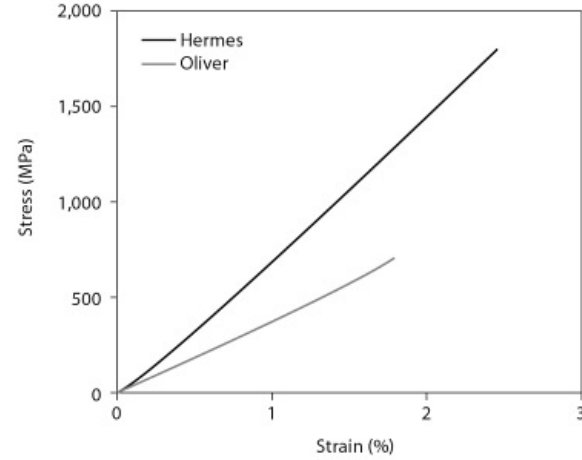
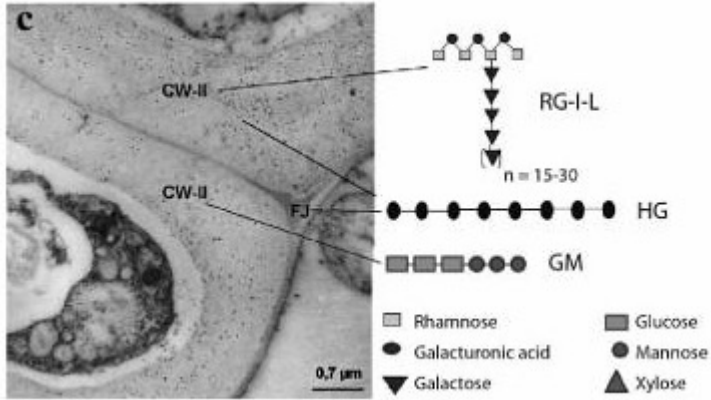
50% cellulose
 25% hemicelluloses - pectines
 25% lignin
 + extractives, ashes
 MFA = (0) 8-40°

Flax G

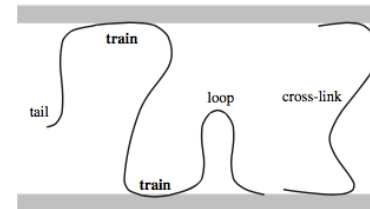
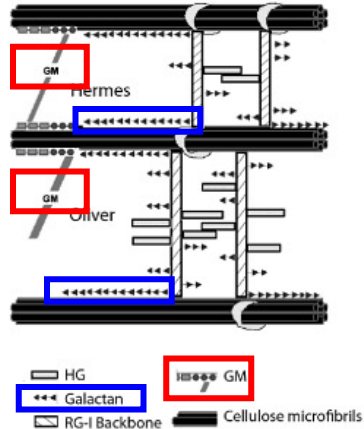
80% cellulose
 20% hemicelluloses - pectines
 ≈0% lignin
 + protein
 MFA = (0) 5-10°



IMPACT ON STIFFNESS

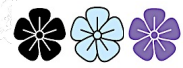


**More structuring pectins in Hermes:
higher stiffness**

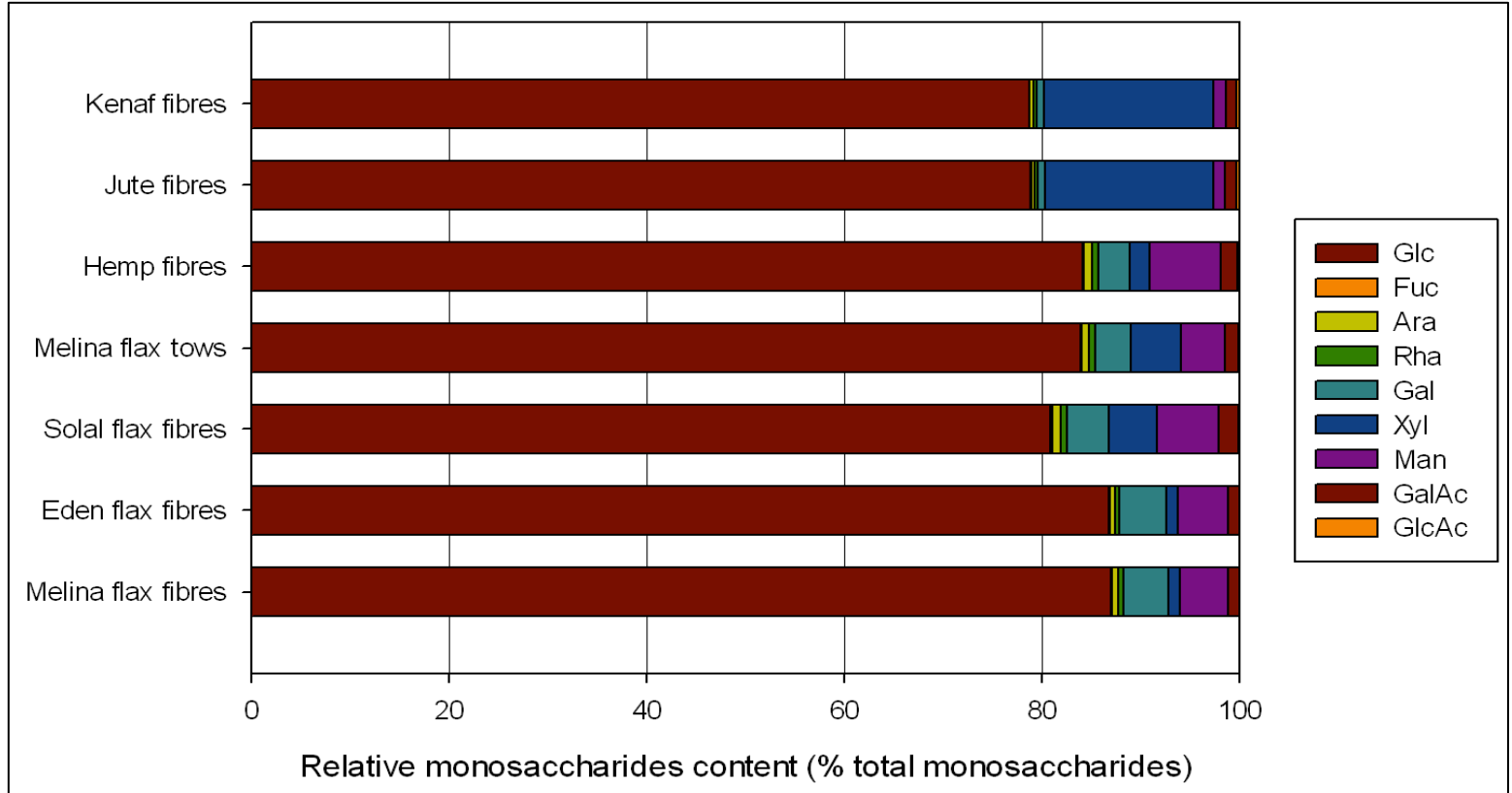


Alix et al., Pectins et Pectinases, 2008

Zykwinska, J Exp Bot, 2007



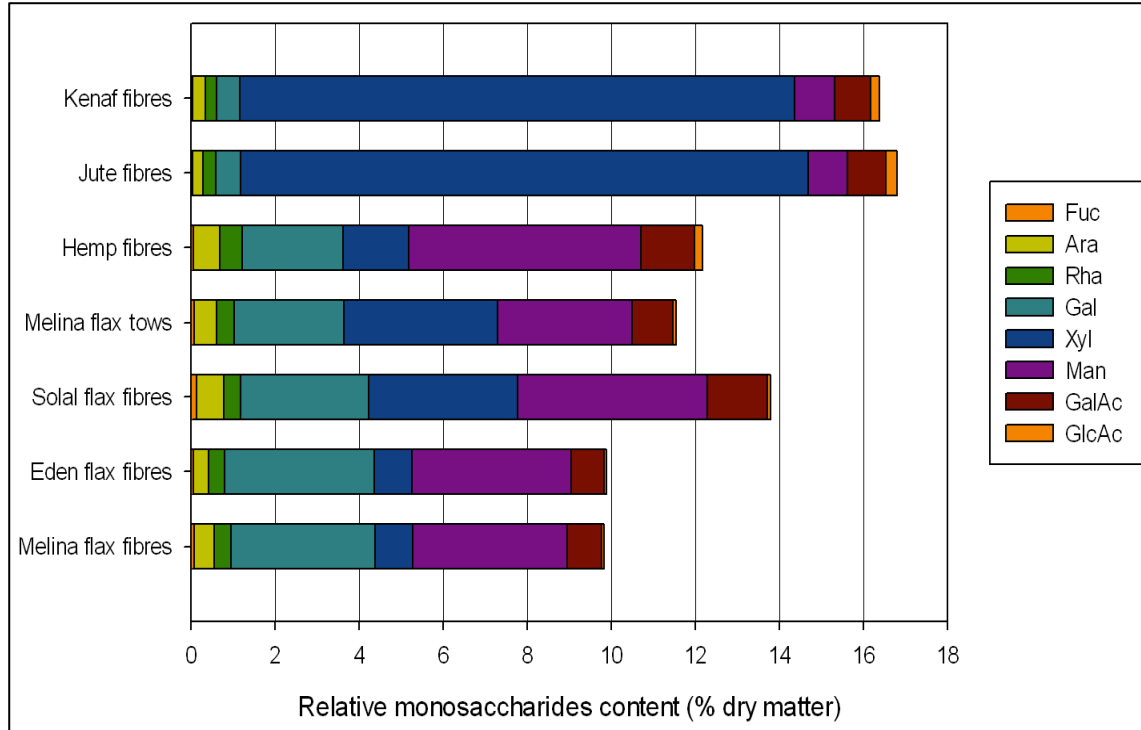
DIFFERENCES BETWEEN THE ORIGIN



More cellulose for scutched textile flax, Lower content for kenaf and jute

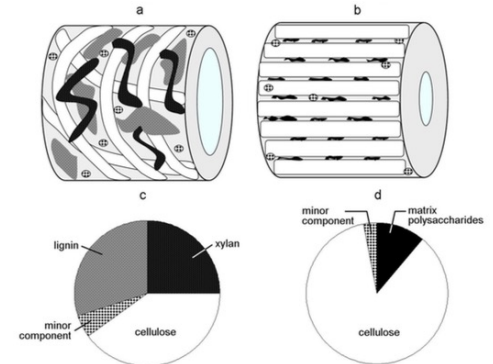


DIFFERENCES BETWEEN THE ORIGIN



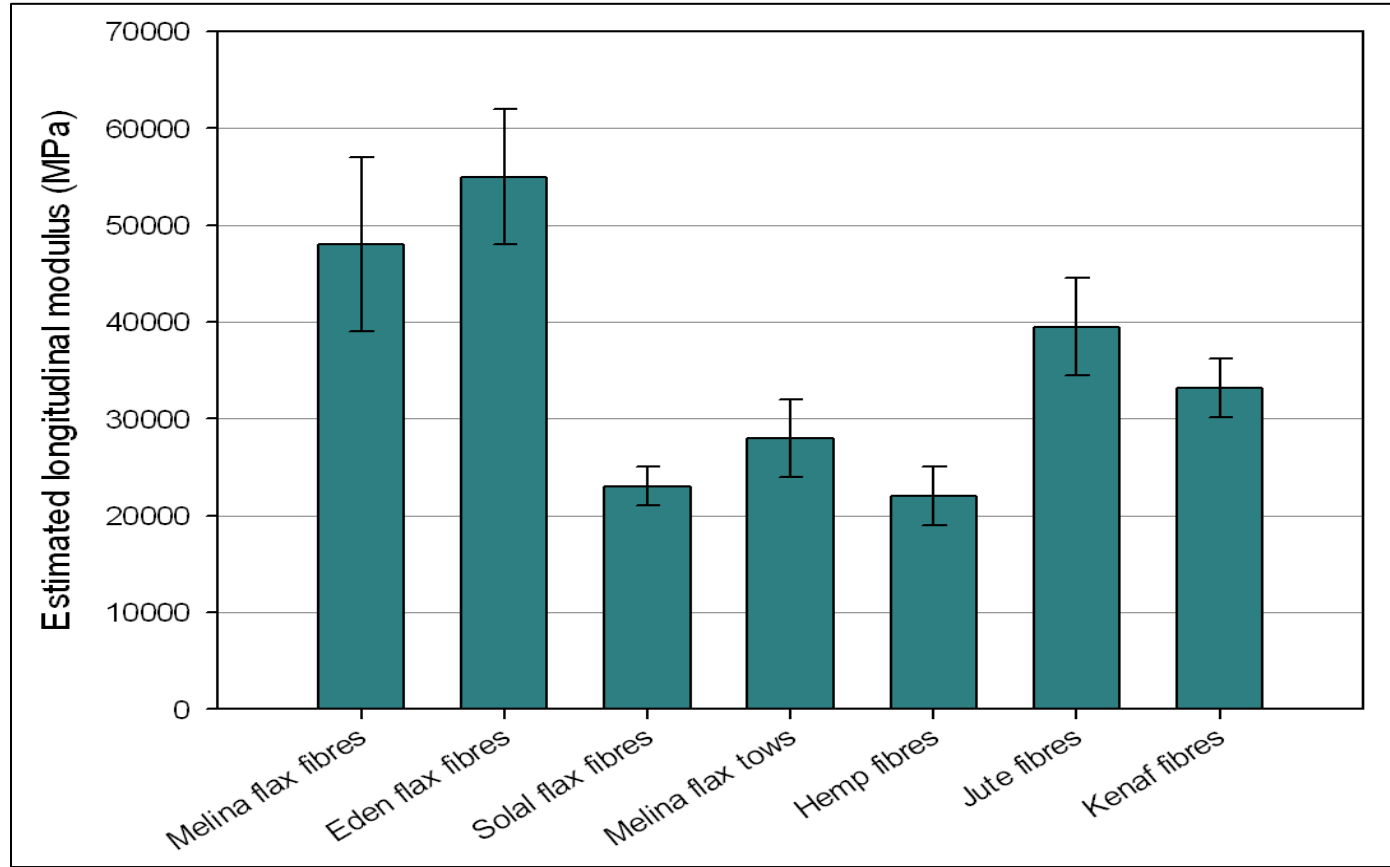
3 main families:

- Kenaf & jute
- Scutched textile flax
- Hemp, flax tows and oleaginous flax





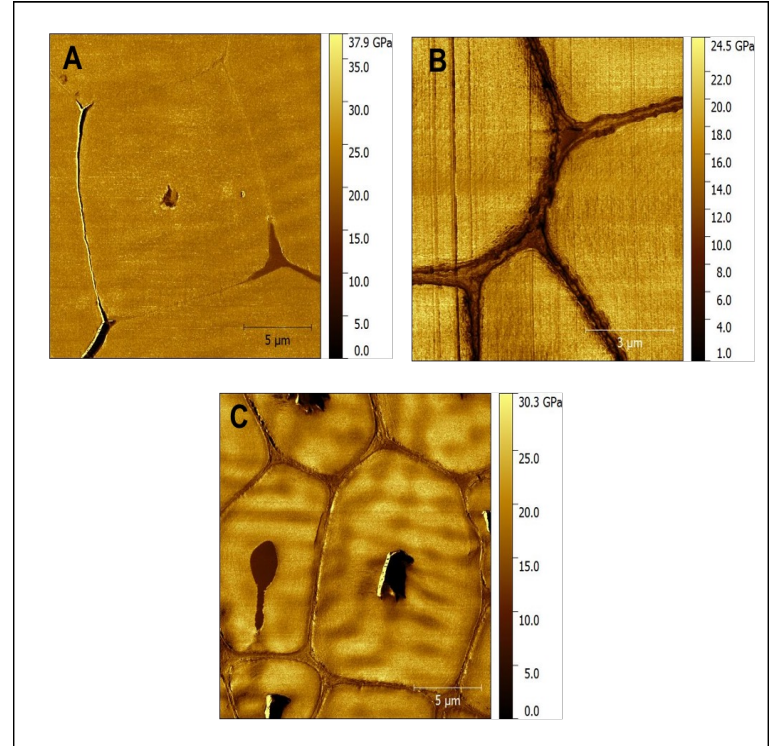
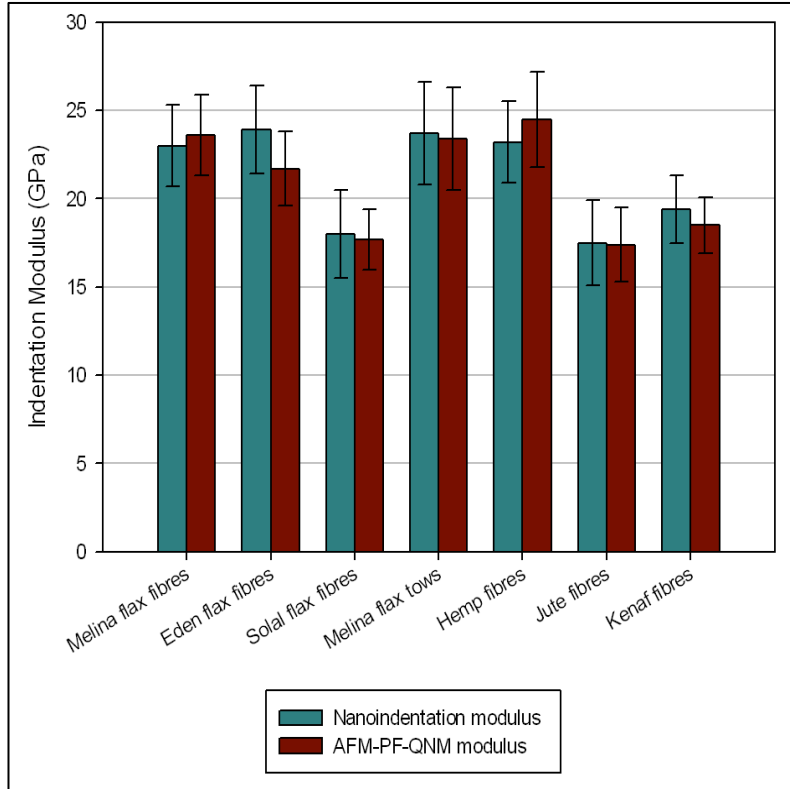
DIFFERENCES BETWEEN THE ORIGIN

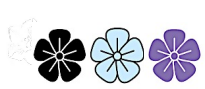


Use of ROM to estimate the fibre stiffness from epoxy-fibre UD composite

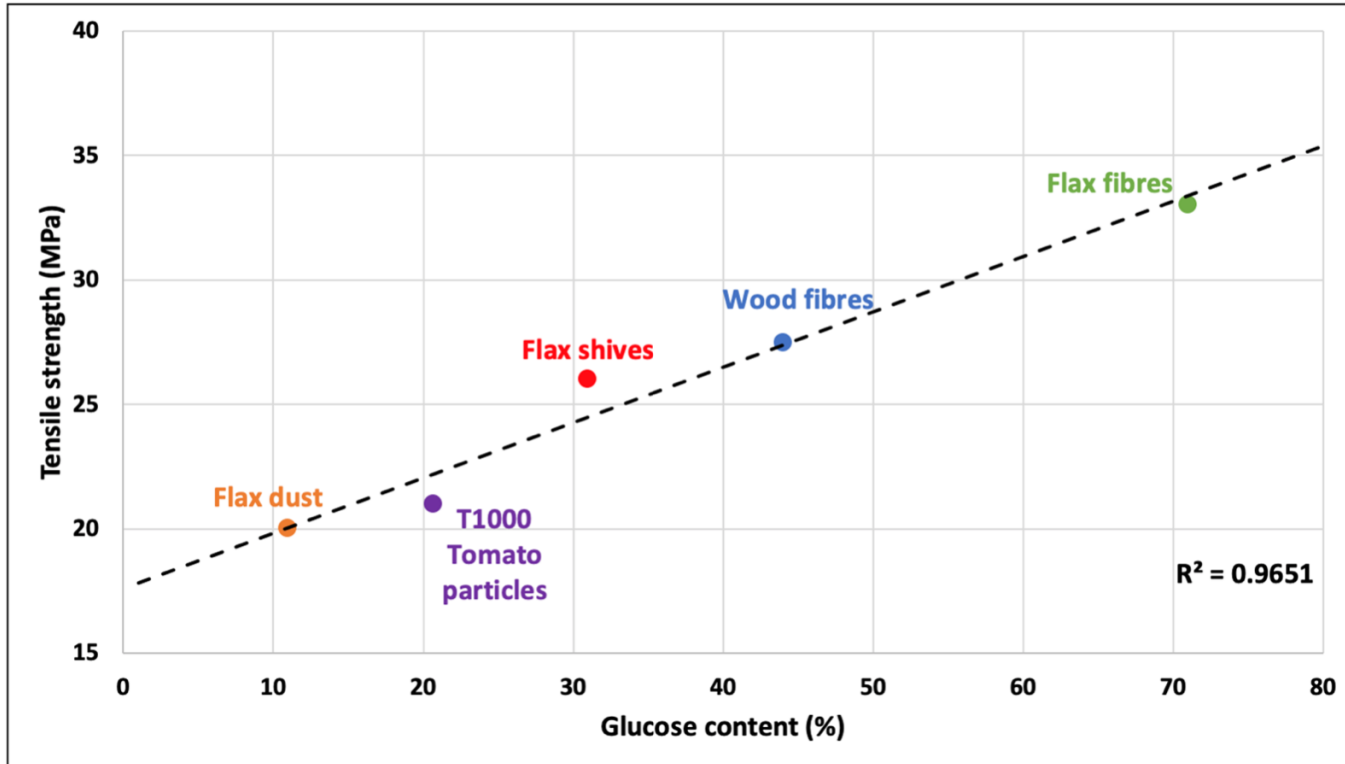


DIFFERENCES BETWEEN THE ORIGIN





DIFFERENCES BETWEEN THE ORIGIN



Injected PP-plant fibre composites – Same fibre volume fraction and same PP



MULTI SCALES MECHANICAL INVESTIGATIONS

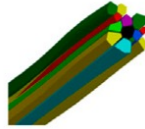


FOUR MAIN SCALES

Data for Composite Engineer: the pertinent scale for the pertinent informations



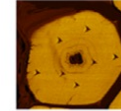
Trunc



Bundle



Fibre



Cell wall

Interest of this multi-scale approach:

- Need of understanding: Function, Relationship between biochemistry/mechanical properties
- Data for Engineer: both textile & composite



i) **Textile:** in our case: Elaboration of semi-products and preforms for composite reinforcement

ii) **Composites:** Short fibres for extrusion/injection



Mechanical properties at break

σ_T^+ , σ_T^- , σ_L^+ , σ_L^- , τ_{LT} , ϵ_L^+ , ϵ_L^- , ϵ_T^+ , ϵ_T^-

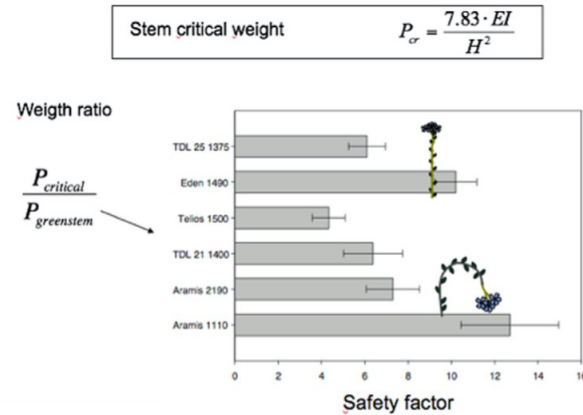
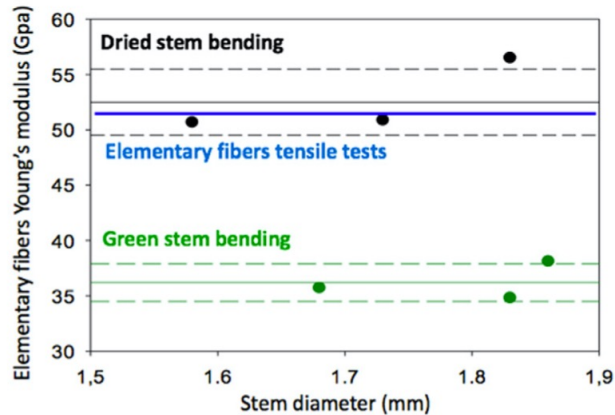
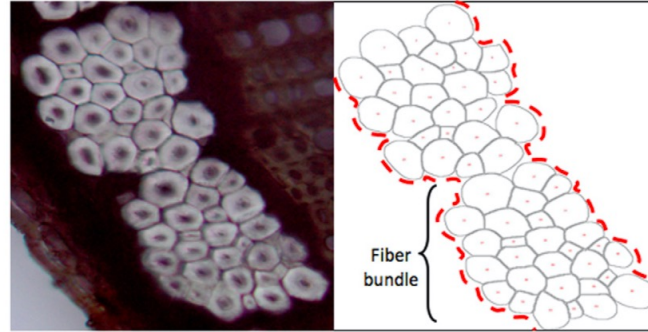
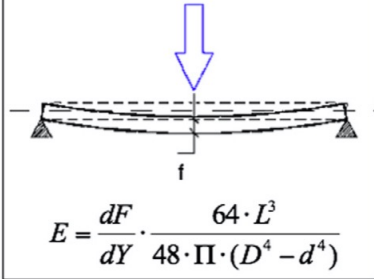
Mechanical properties of an anisotropic reinforcement

E_{fL} , E_{fT} , G_{fLT} , ν_{LT}

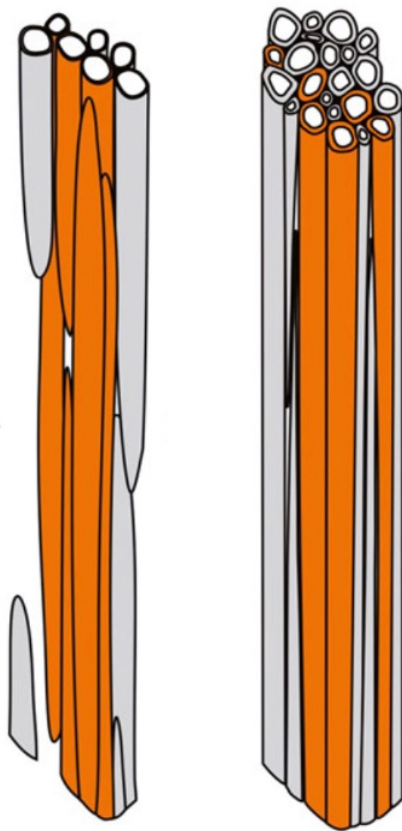


STEM SCALE

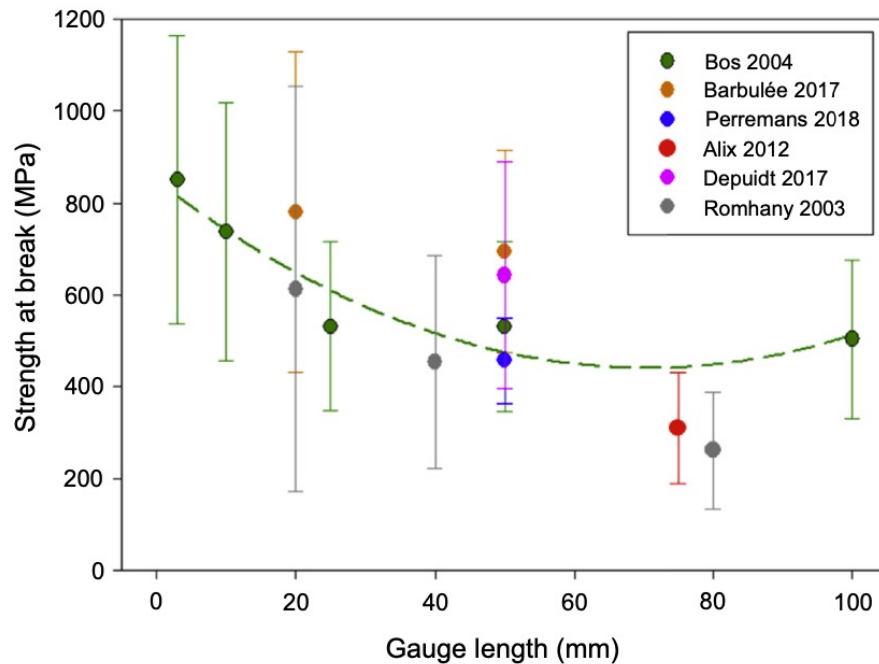
Bending tests on stems



BUNDLE SCALE

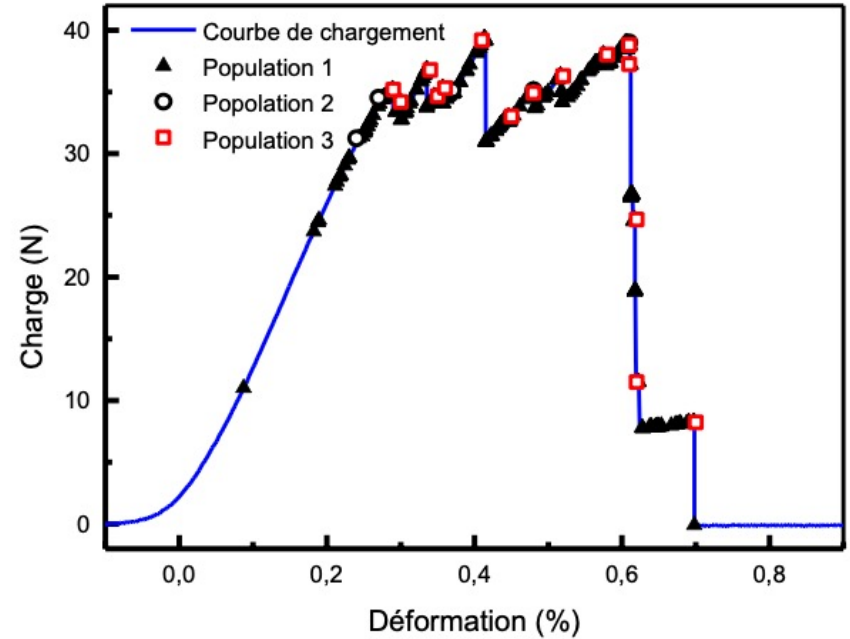
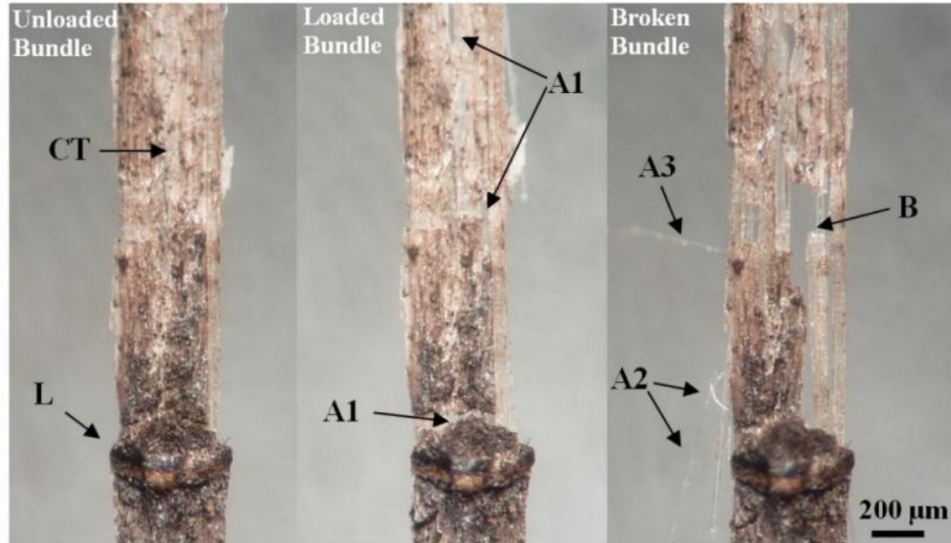


Snegireva 2015





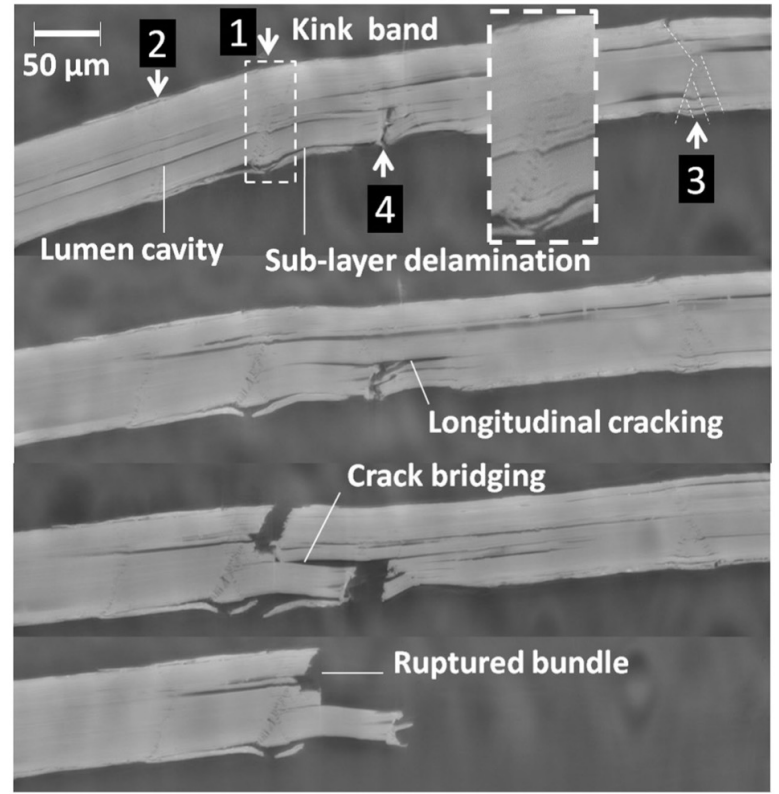
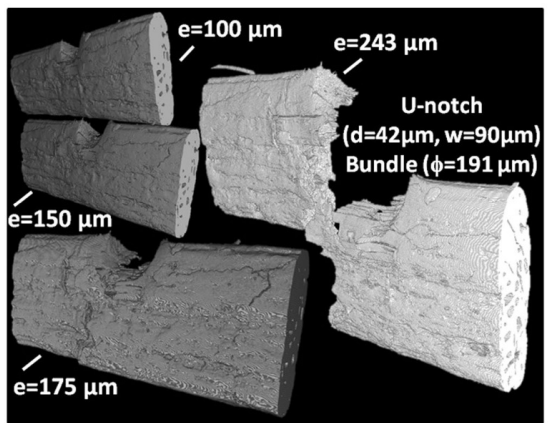
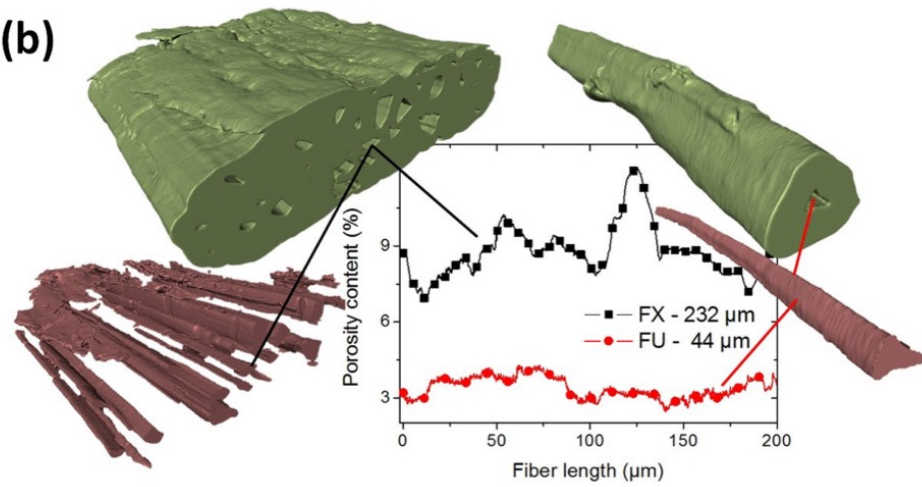
BUNDLE SCALE

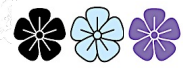




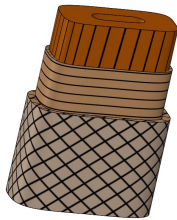
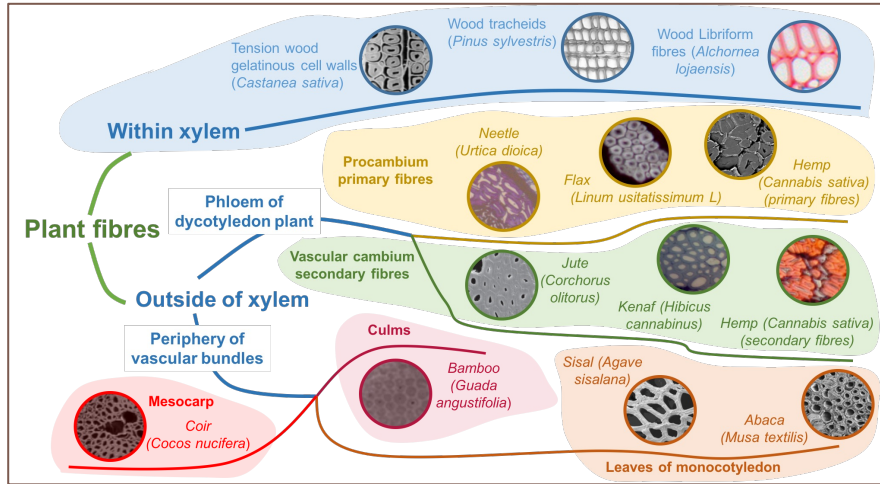
BUNDLE SCALE

(b)

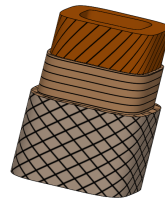




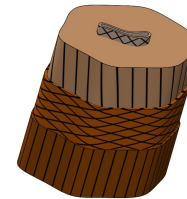
SINGLE FIBRES AND CELL WALLS



Tension wood



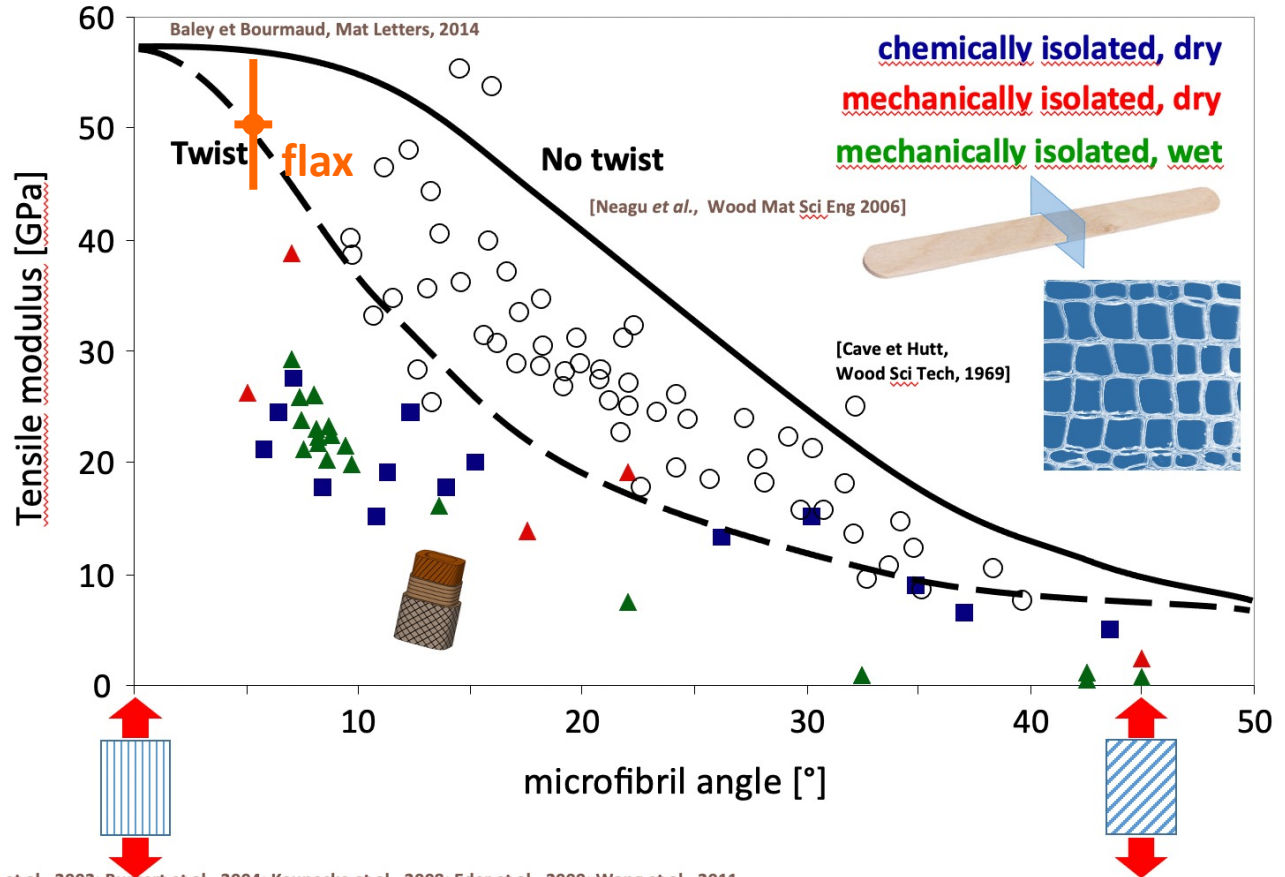
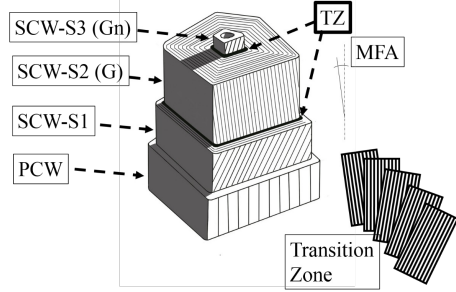
Normal wood



Flax

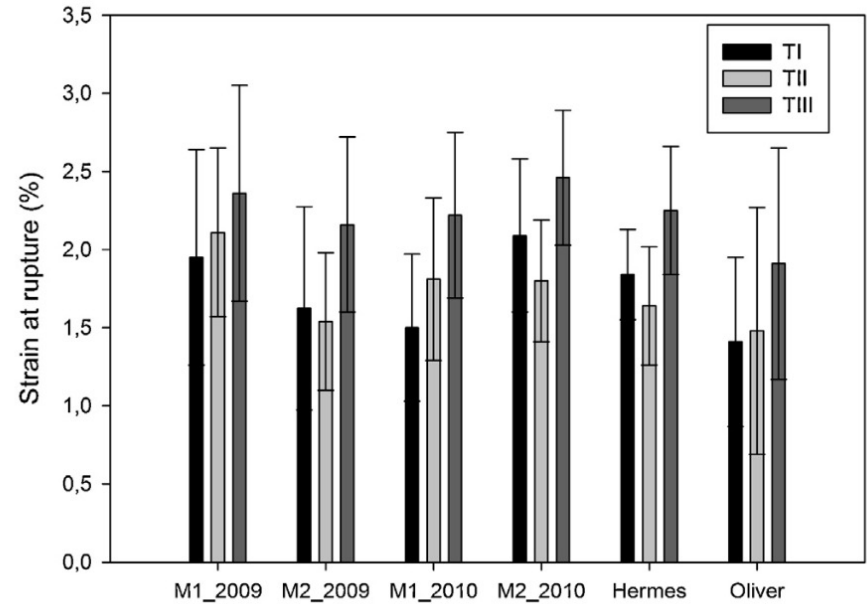
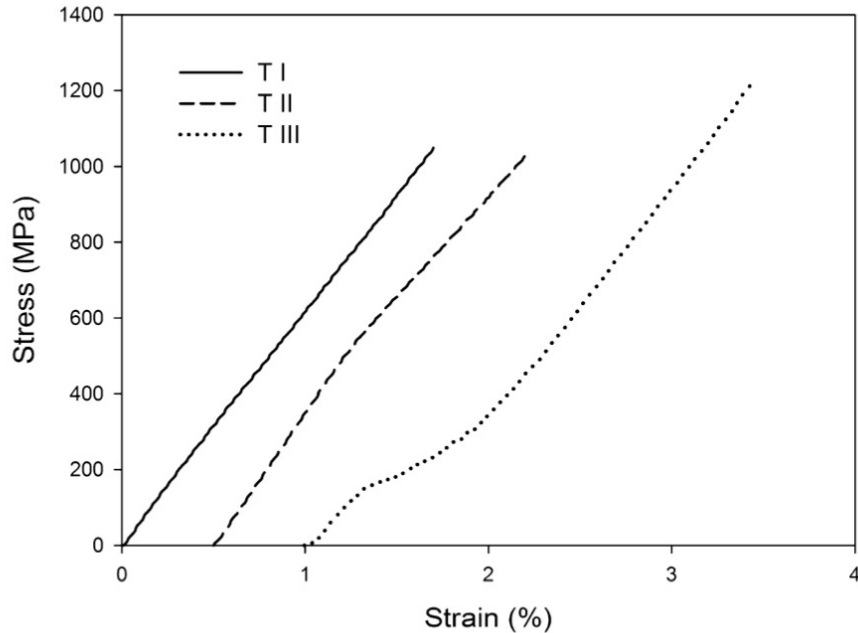
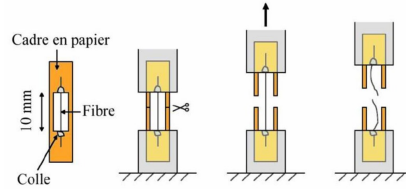


SPECIFIC MECHANICAL BEHAVIOUR OF PLANT FIBRES



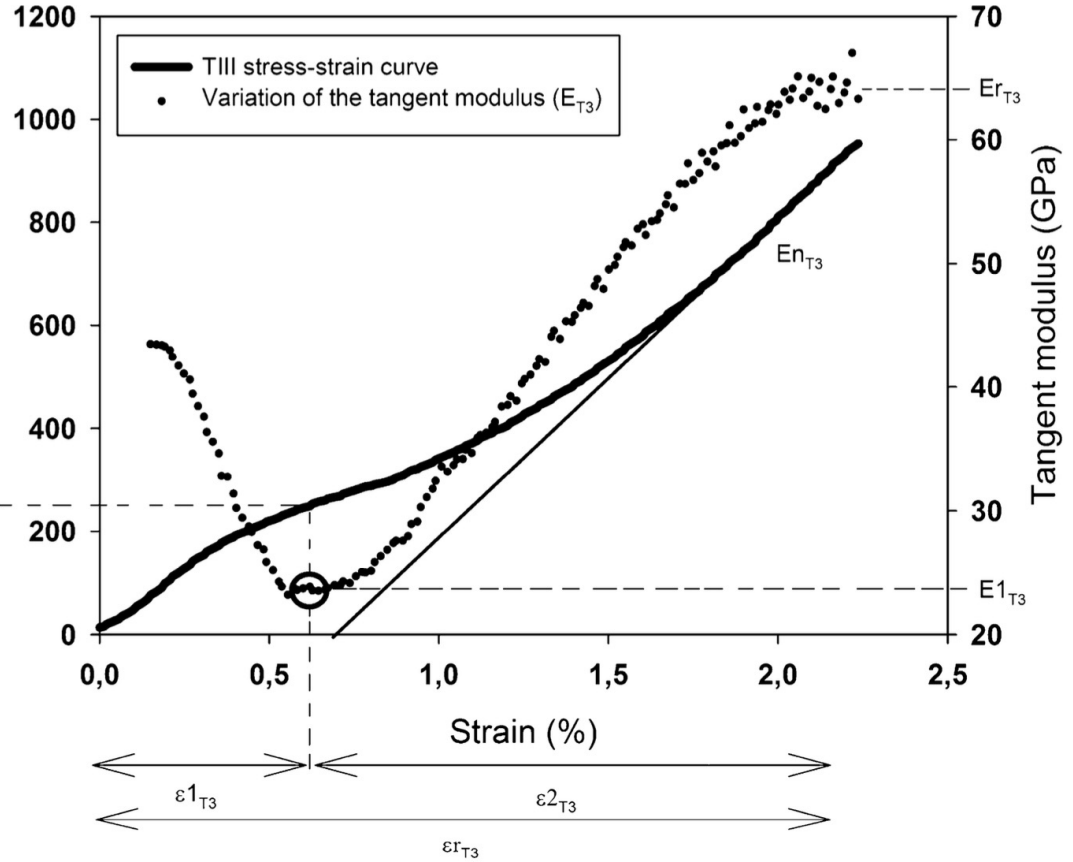
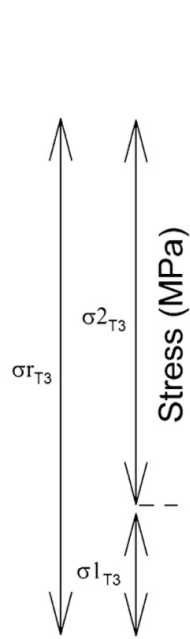
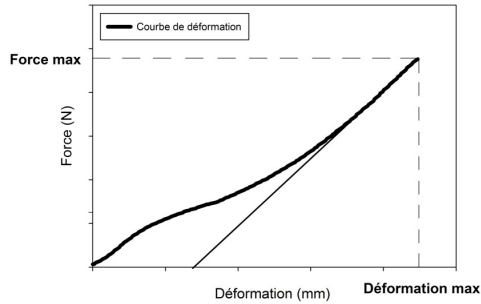
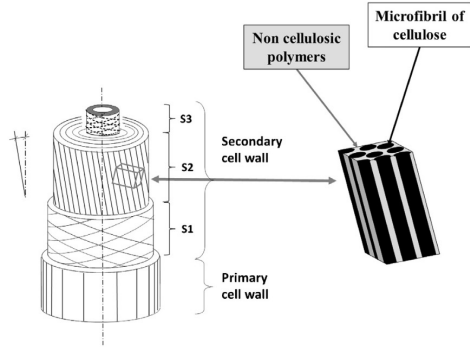


SPECIFIC MECHANICAL BEHAVIOUR OF PLANT FIBRES



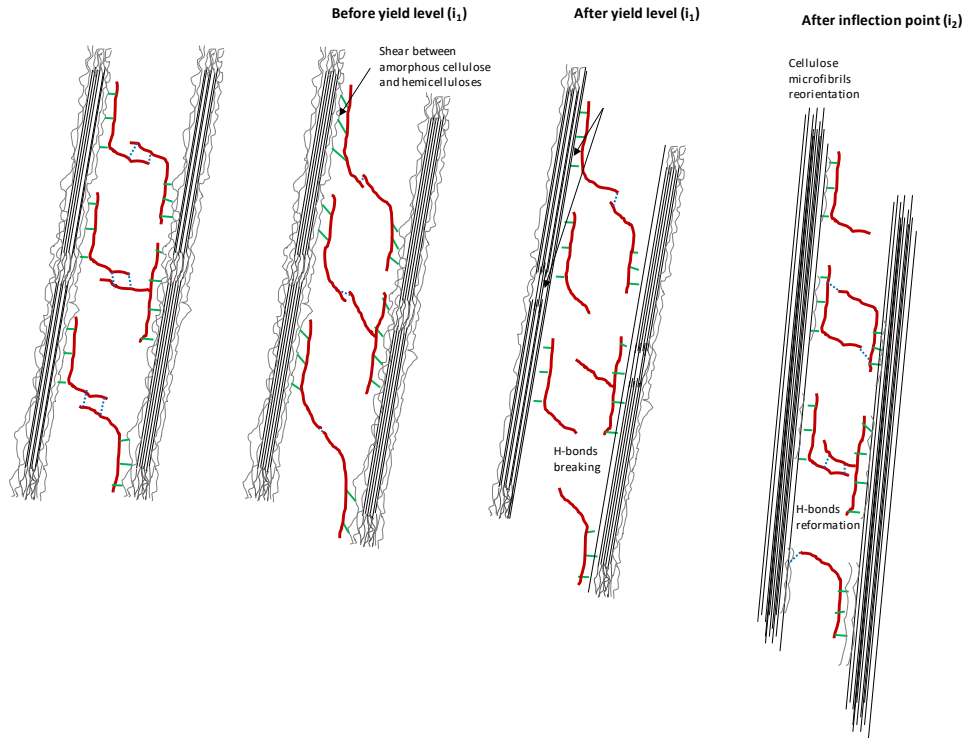


SPECIFIC MECHANICAL BEHAVIOUR OF PLANT FIBRES





SPECIFIC MECHANICAL BEHAVIOUR OF PLANT FIBRES



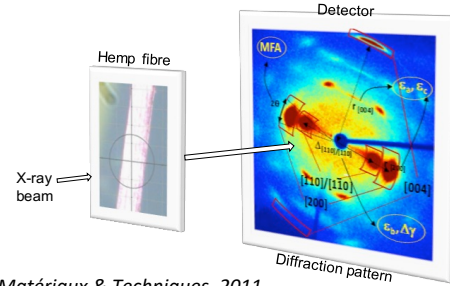
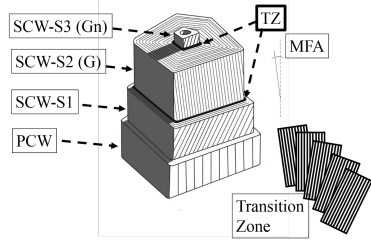
Possible scenario describing the various mechanisms contributing to the multiple nonlinearities of the stress-strain curve of hemp fibre.

Segment (point) of the stress-strain curve	Observations	Possible mechanisms
I	Quasi-linear behaviour with slightly irreversible strain	<ul style="list-style-type: none"> Elastic deformation of the cellulose microfibrils and amorphous polymers Slight rotation of the microfibrils towards a more parallel orientation
i_1	Yield level	<ul style="list-style-type: none"> Matrix flow threshold: bonds break in the amorphous matrix
II	Apparent decrease in fibre stiffness	<ul style="list-style-type: none"> Viscous flow of the amorphous components under shear strain and lock-in at a new position Stress-induced crystallisation of the paracrystalline cellulose Spiral spring-like extension of the cellulose microfibrils in the amorphous matrix
i_2	Inflection point	<ul style="list-style-type: none"> Maximum flow point of the matrix Crystallisation saturation point
III	Quasi-linear or parabolic	<ul style="list-style-type: none"> Deployment of cellulose microfibrils in dislocation areas Decrease of the mean MFA Interfacial rupture between crystalline cellulose and the amorphous matrix

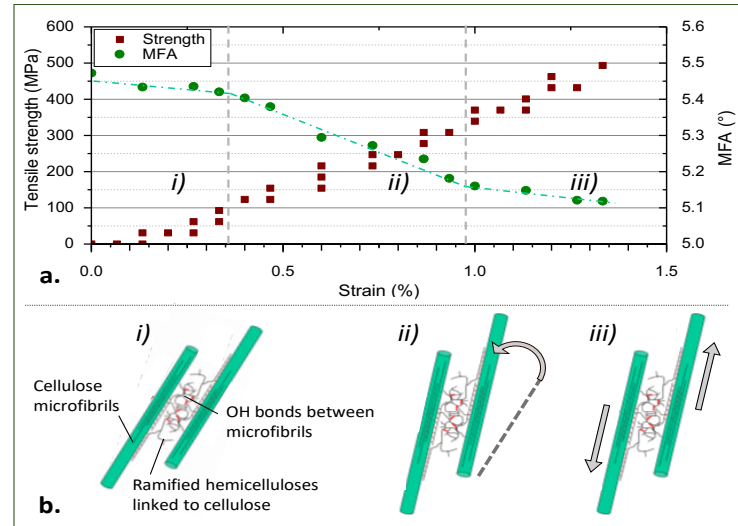
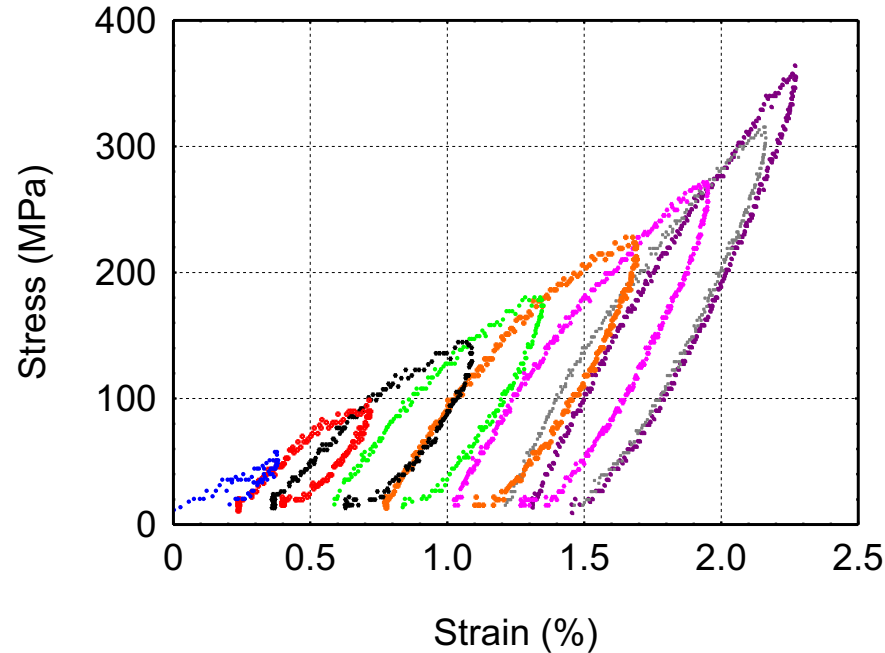


SPECIFIC MECHANICAL BEHAVIOUR OF PLANT FIBRES

Stiffening effect

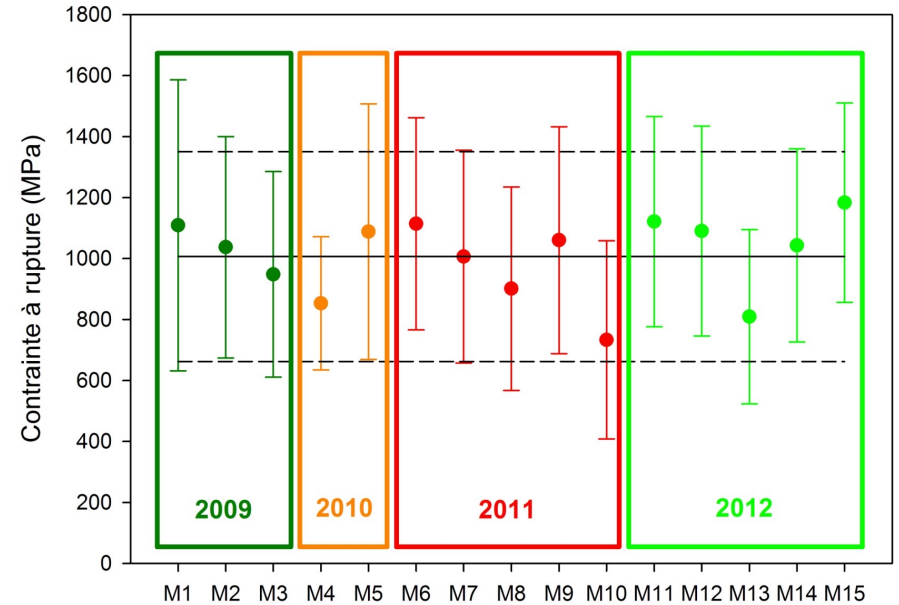
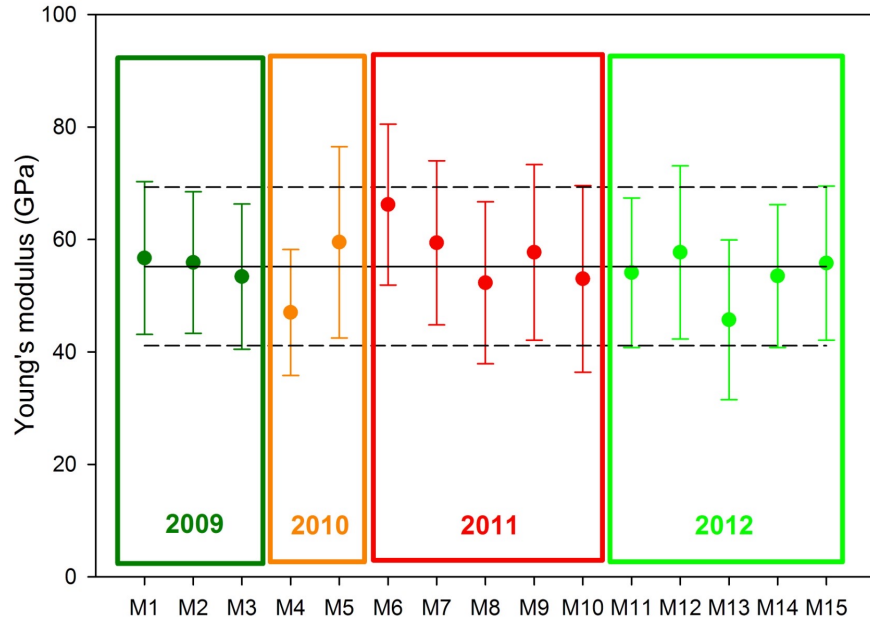


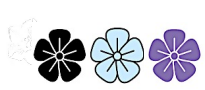
Placet et al. *Matériaux & Techniques*, 2011
 Bourmaud et al. *Ind Crop Prod*, 2013



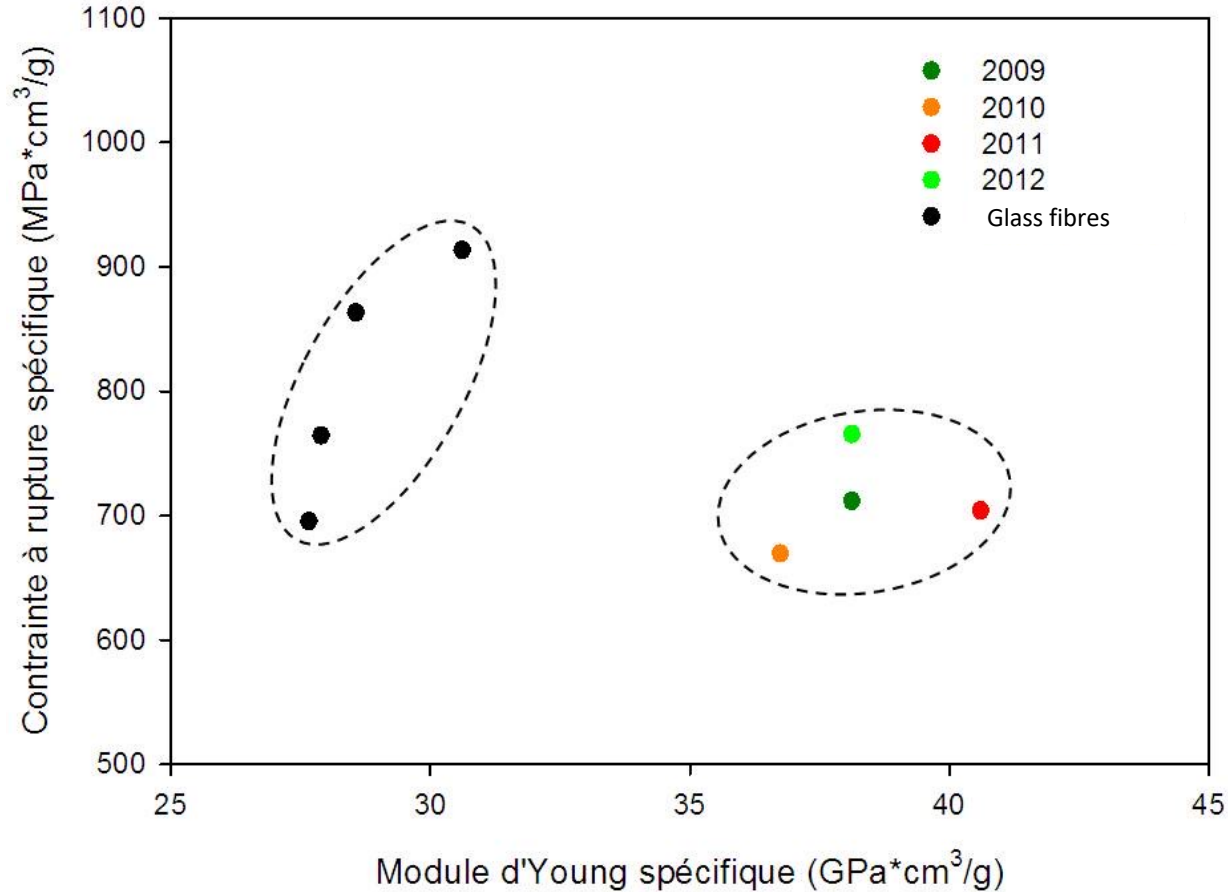


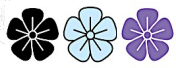
SPECIFIC MECHANICAL BEHAVIOUR OF PLANT FIBRES



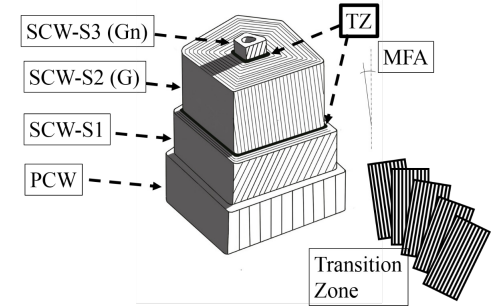
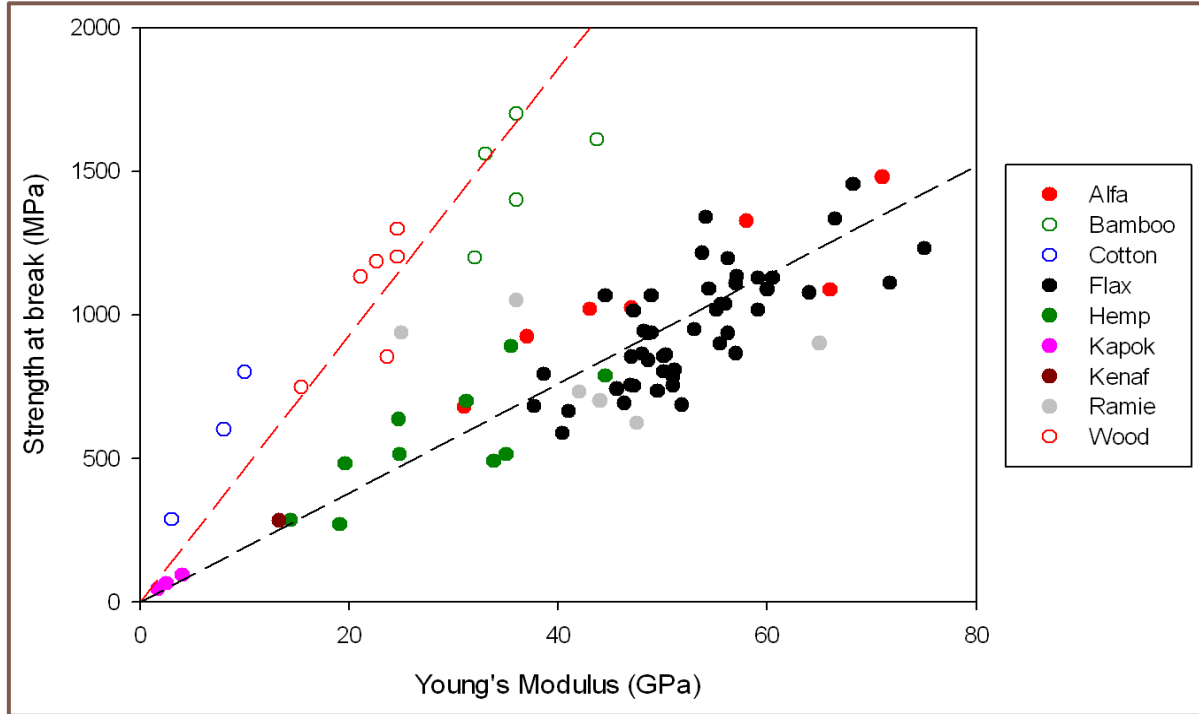


SPECIFIC MECHANICAL BEHAVIOUR OF PLANT FIBRES

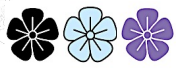




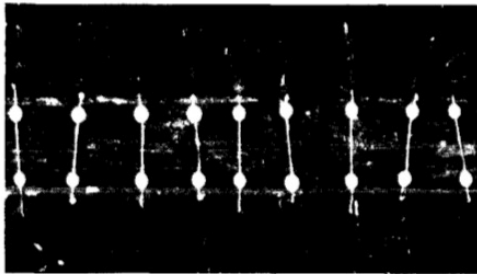
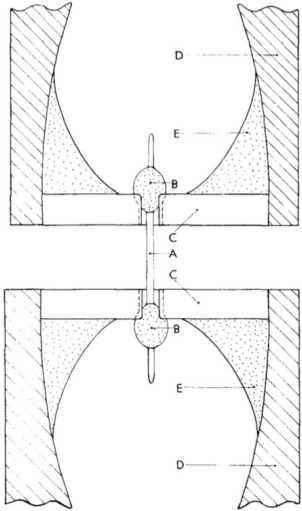
SINGLE FIBRES AND CELL WALLS



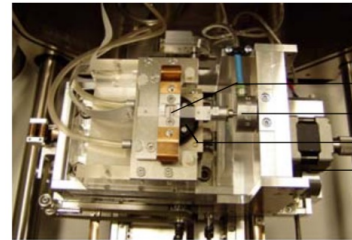
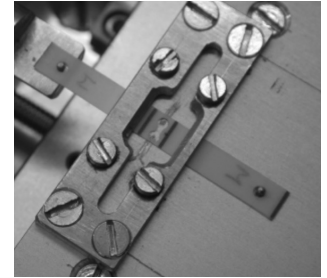
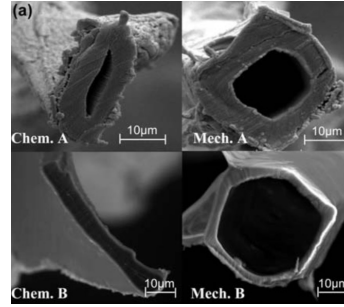
**But strong limits,
especially in link with
the length of fibres for
a range of species**



TENSILE TESTS ON SHORT FIBRES

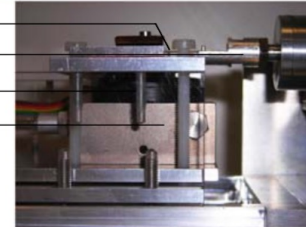


Kersevage, Wood Fiber Sci, 1973



- sample
- 500 mN load cell
- Peltier cooling stage
- step motor

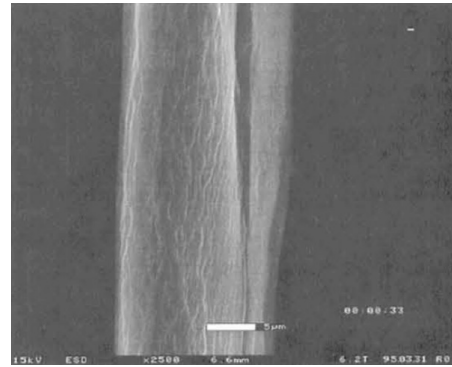
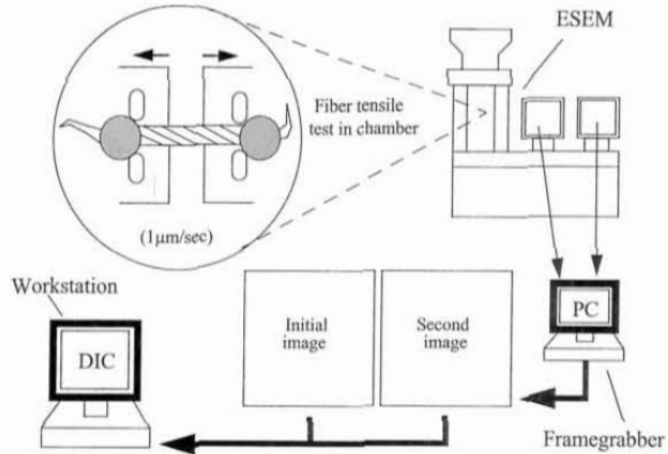
- foliar frame with sample
- load cell (with pin)
- Peltier cooling stage
- water cooling stage



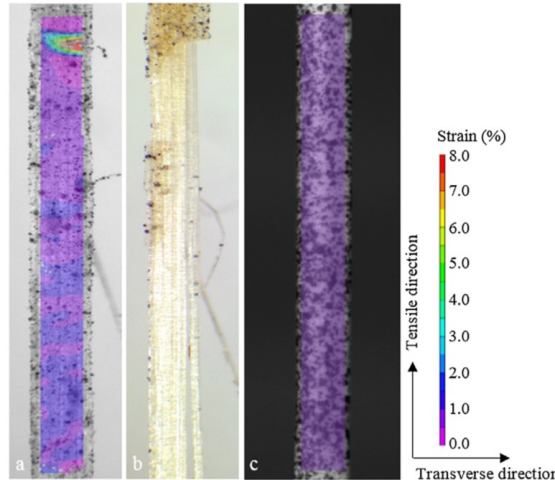
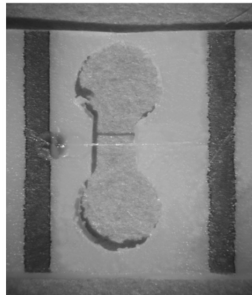
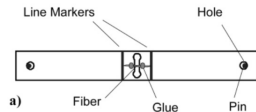
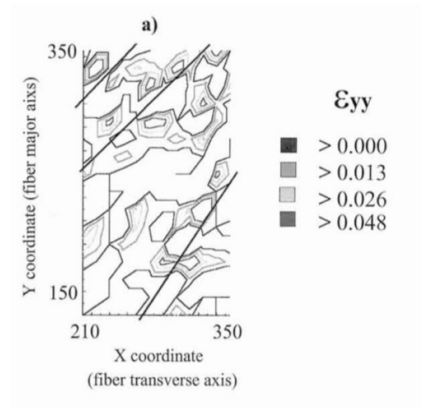
Eder, Wood Sci Technol, 2008



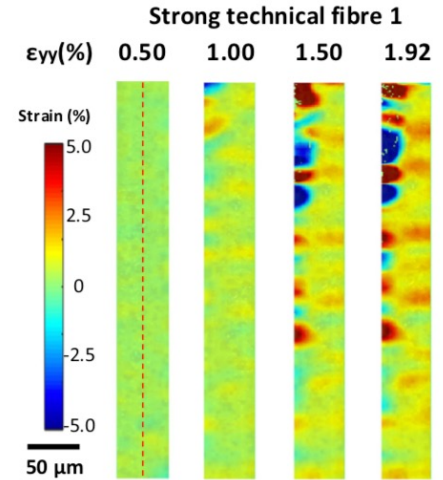
STRAIN MONITORING



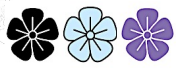
Mott, Wood Sci Technol, 1996



Burgert, Holzforschung, 2003

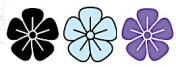


Fuentes, Comp Part A, 2017

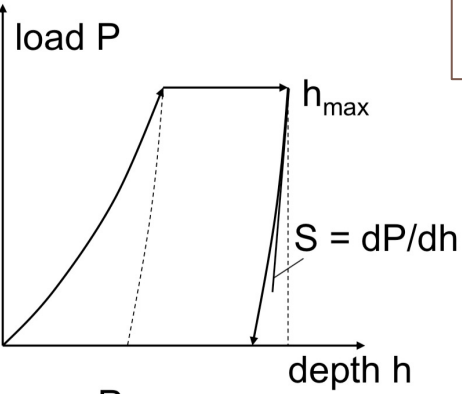


LIMITS OF TENSILE TESTING ON SINGLE FIBRES

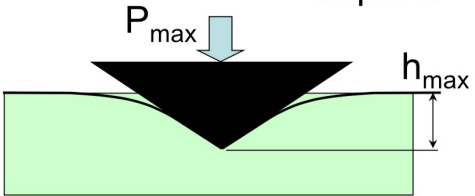
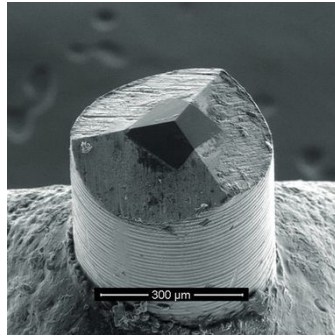
- **Difficult to implement for a range of short plant fibres: wood, sisal, jute....**
- **Apparent mechanical properties: interesting for composite reinforcement but not at a lower scale.**
- **There is a need for mechanical investigations at the cell wall scale.**
- **Link between ultrastructure and mechanical properties: impact of MFA, multi layers arrangement.....**



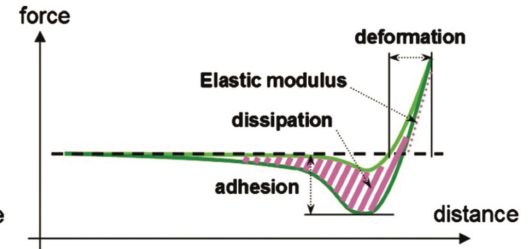
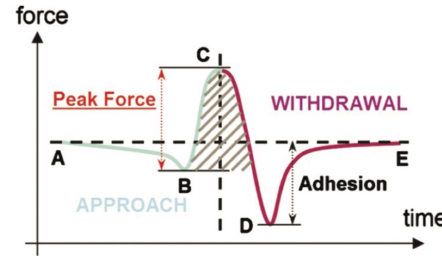
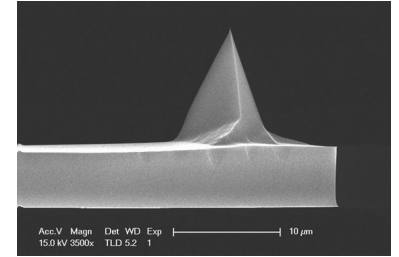
NANOINDENTATION & AFM PEAK FORCE MEASUREMENTS



80-120 nm

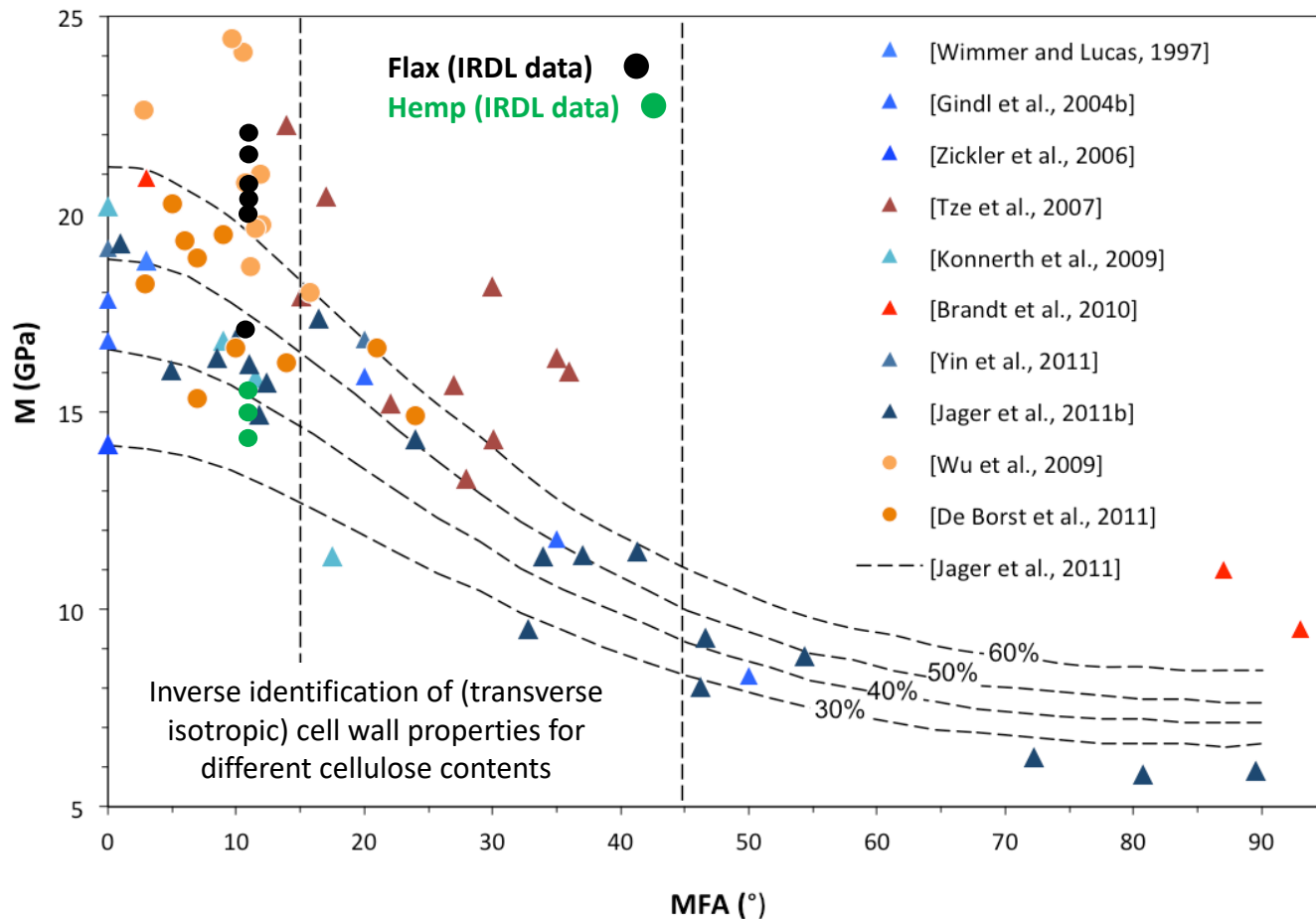


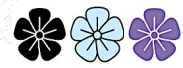
2-3 nm





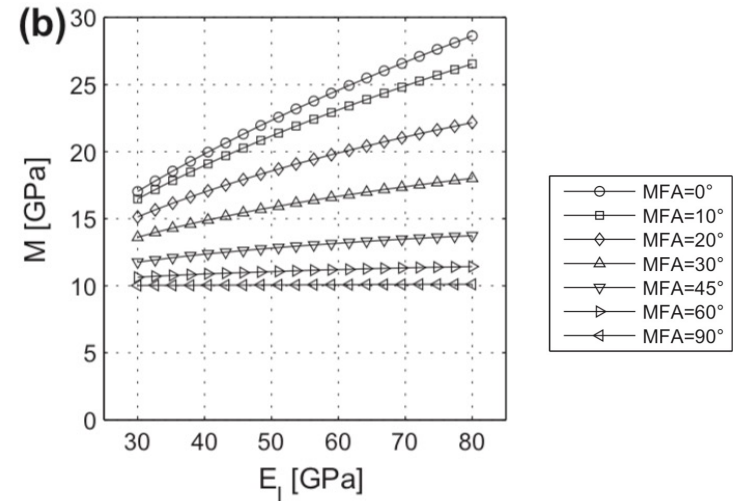
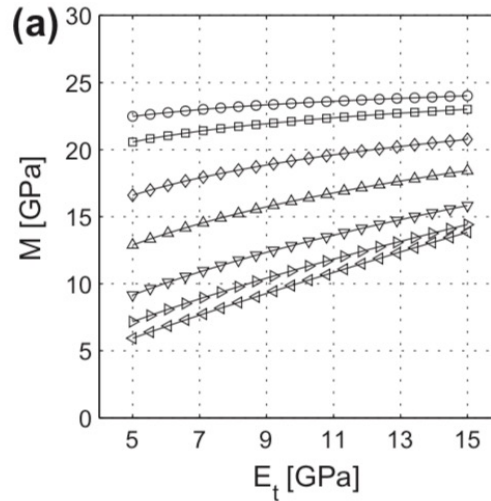
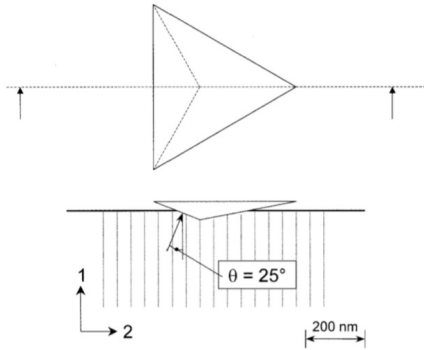
NANOINDENTATION MEASUREMENTS





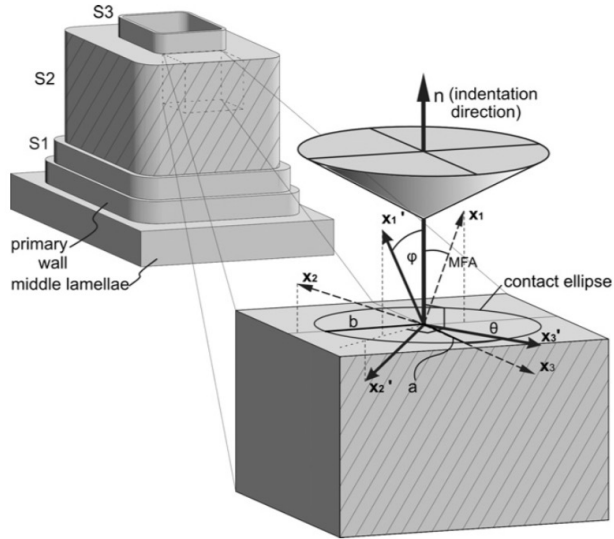
NANOINDENTATION Vs TENSILE TEST

Usual NI models do not consider anisotropy of plant cell walls:
Underestimation of M compared to E_{Lf}





NANOINDENTATION Vs TENSILE TEST



Specific models for anisotropic materials

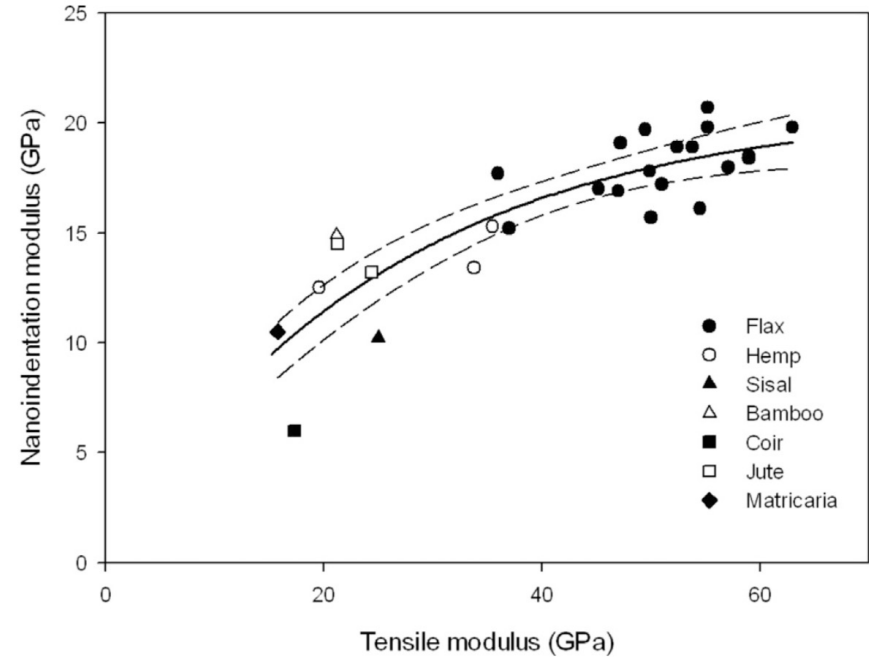
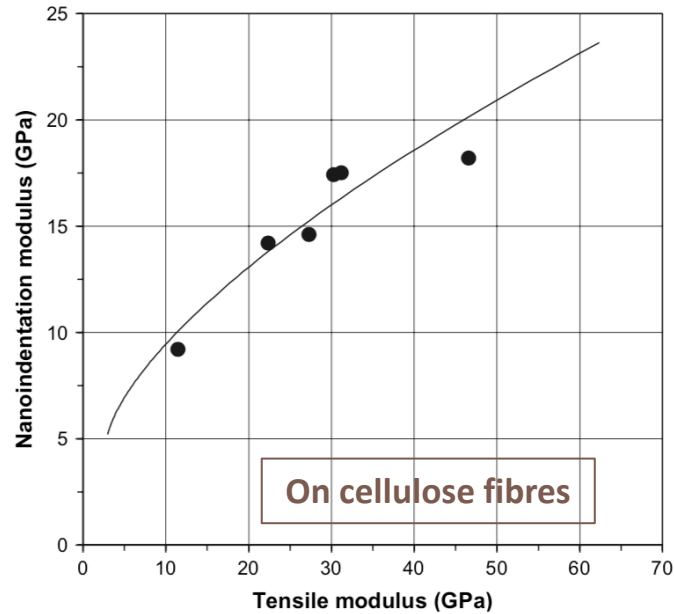
(Vlassak 2003, Swadener 2001)

No direct relation between M and E_{Lf}

The indentation modulus is a function of E_{Lf} , E_{Tf} , and, especially, shear modulus of the cell wall material



NANOINDENTATION VS TENSILE TEST



Some correlation attempts in literature between M and EI

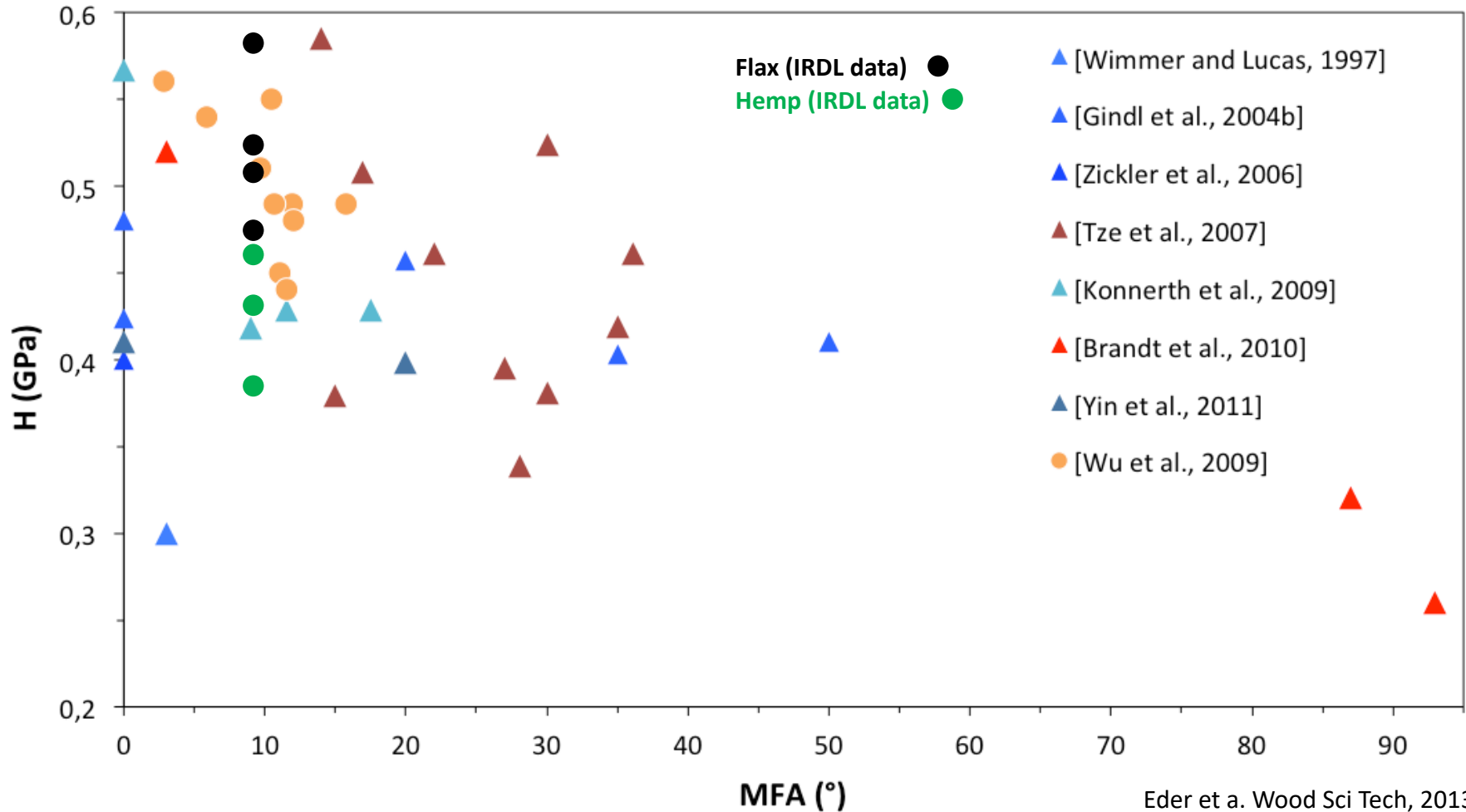
But very questionable due to the moderate impact of E_{Lf} on M, especially for high MFA and possible discrepancy

Gindl, Polymer 2008

Tanguy, Mat Letters, 2014

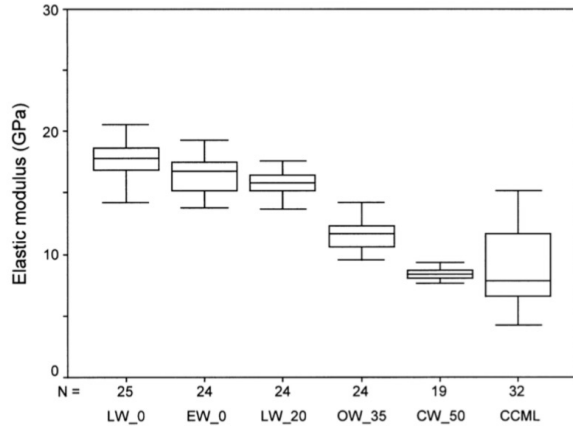


HARDNESS MEASUREMENT

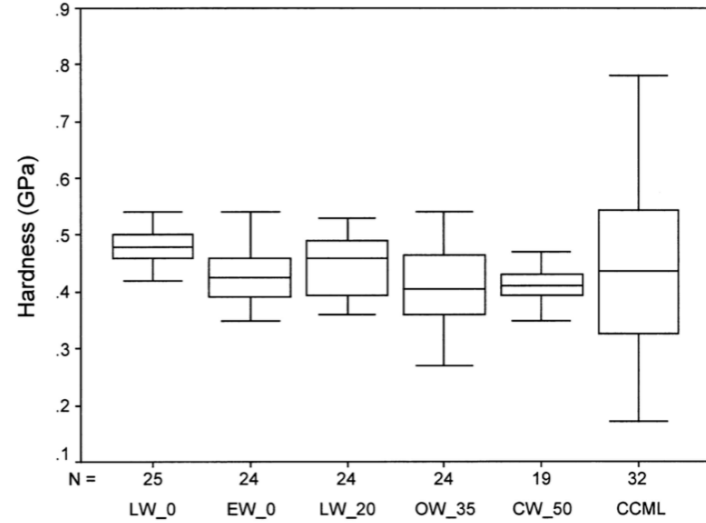




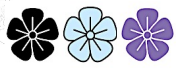
IMPACT OF MFA



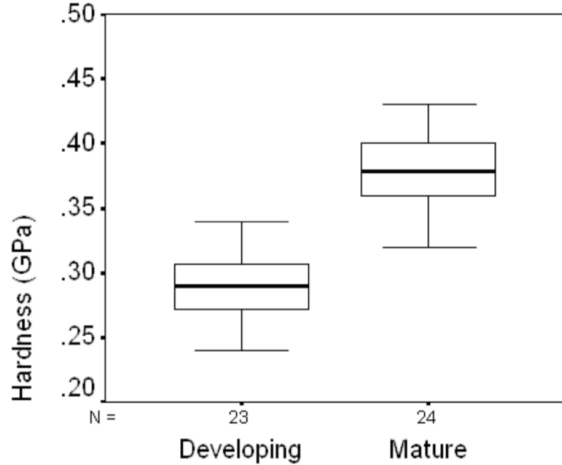
Gindl, Applied Physics, 2004



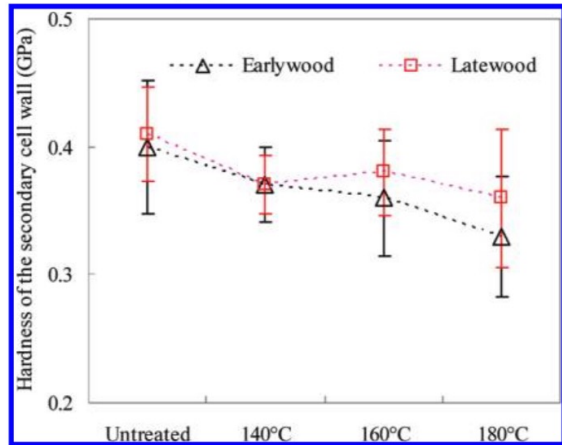
Significant on M, but low on H



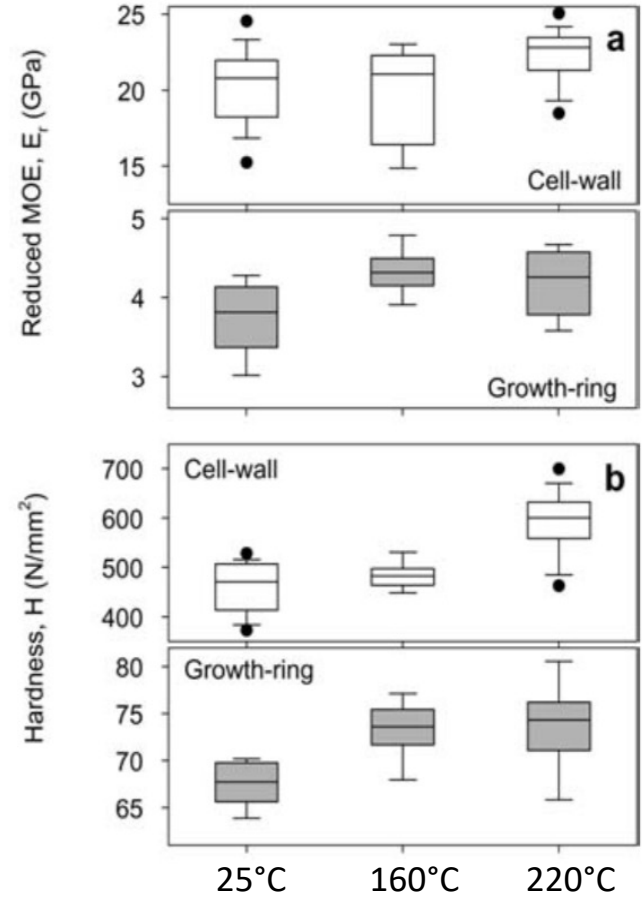
IMPACT OF NON CELLULOSIC POLYMERS



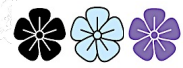
Gindl, Can J Bot, 2002



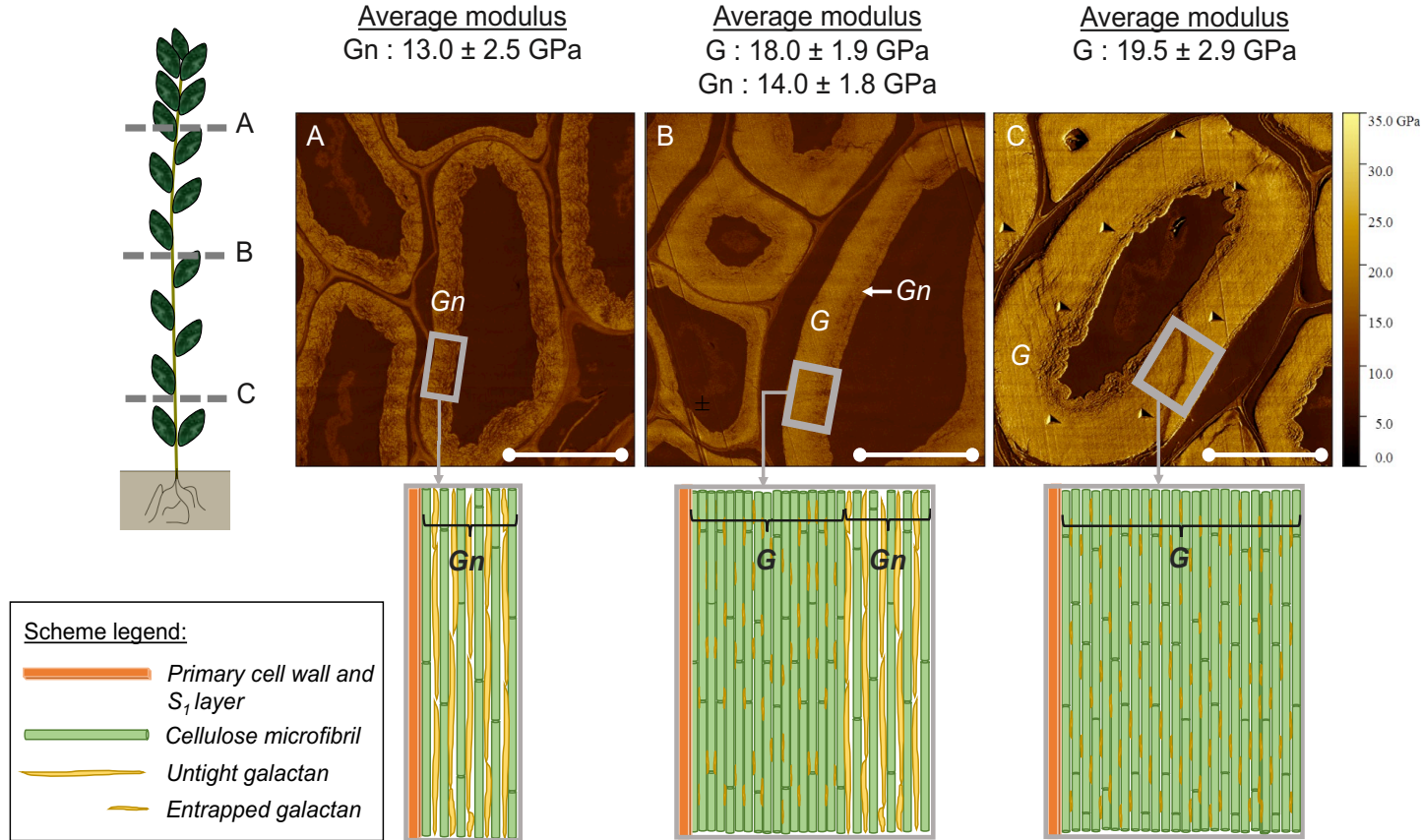
Yin, Biomacromol, 2011

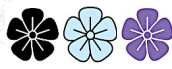


Stanzl-Tschegg, Holzforschung, 2009



APPLICATION: GROWING FLAX FIBRES (60 DAYS)

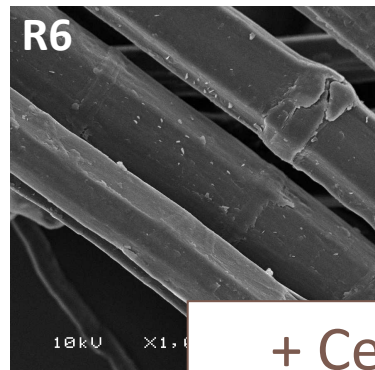
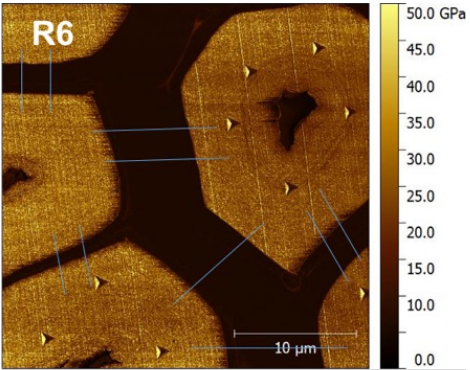
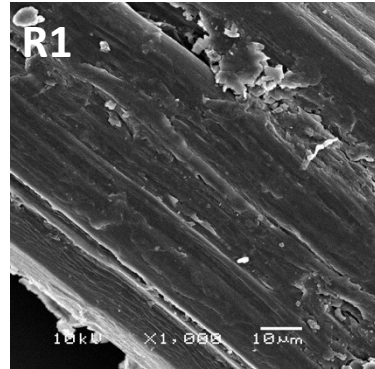
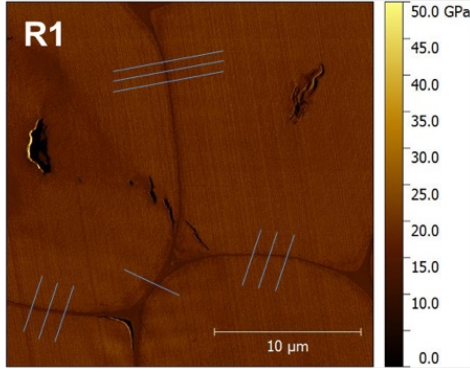




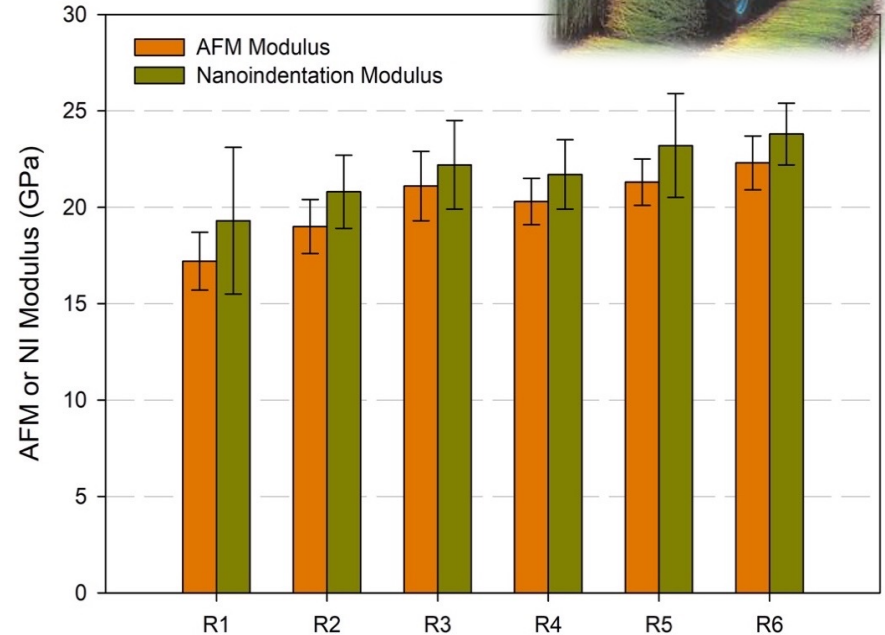
APPLICATION: IMPACT OF RETTING



Contact Modulus (Gpa)



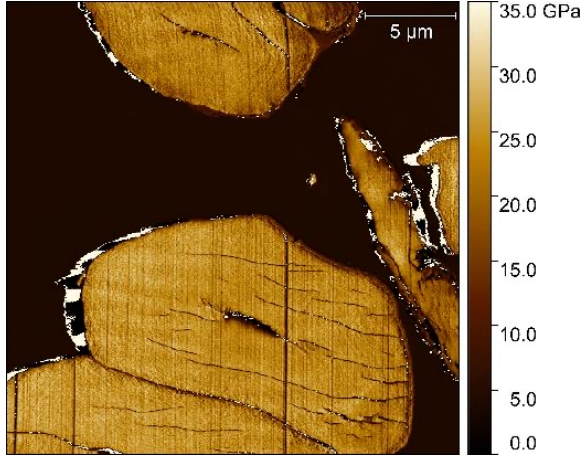
SEM Images



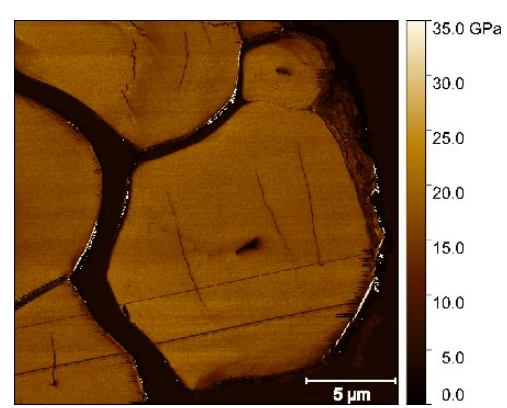
+ Cellulose analysis by XRD and NMR
(↑ cristallinité, macrofibrils compaction)



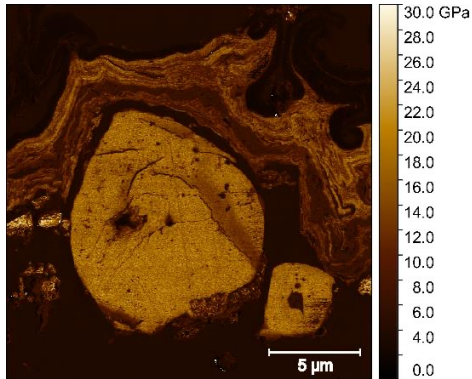
APPLICATION: CULTURAL HERITAGE



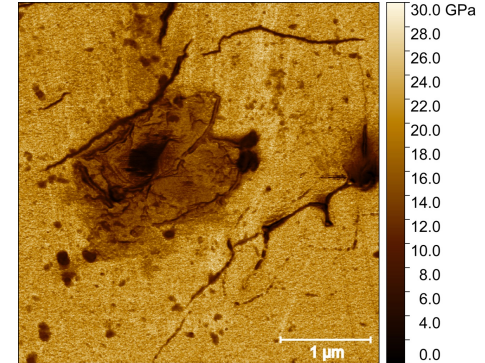
Giulio Benso, San Cristoforo, 1590



Nicola Monti, Madonna col bambino, 1765<<



Tommaso Sciacca,
Crocifissione, 1765

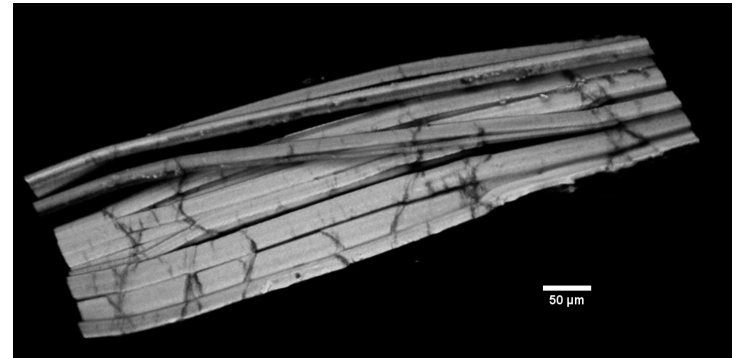
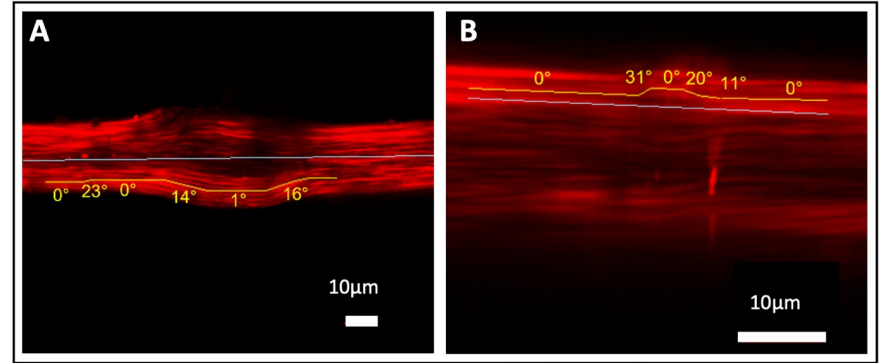
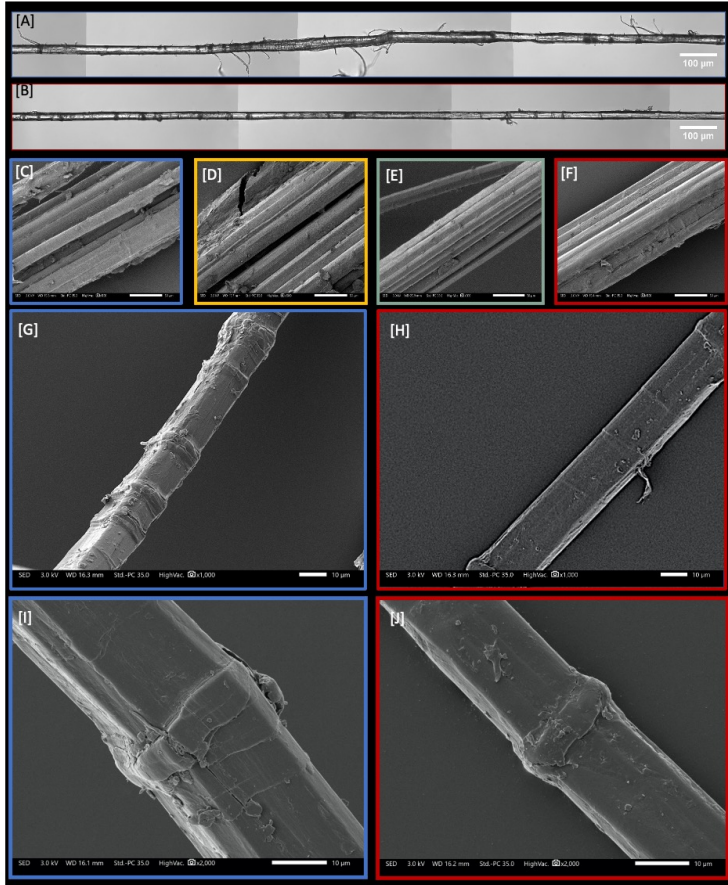




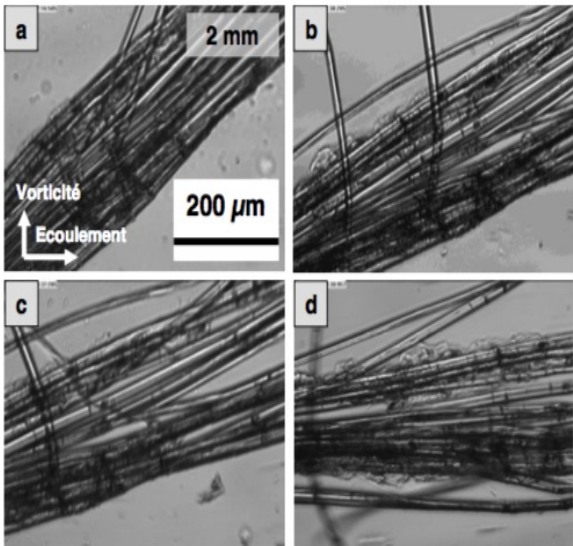
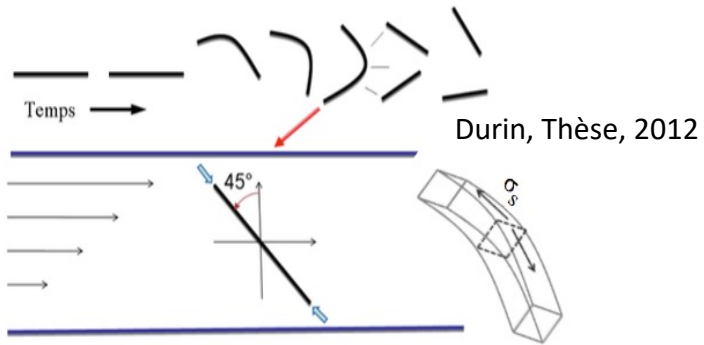
KINK-BAND: A SPECIFIC AND SENSITIVE REGION



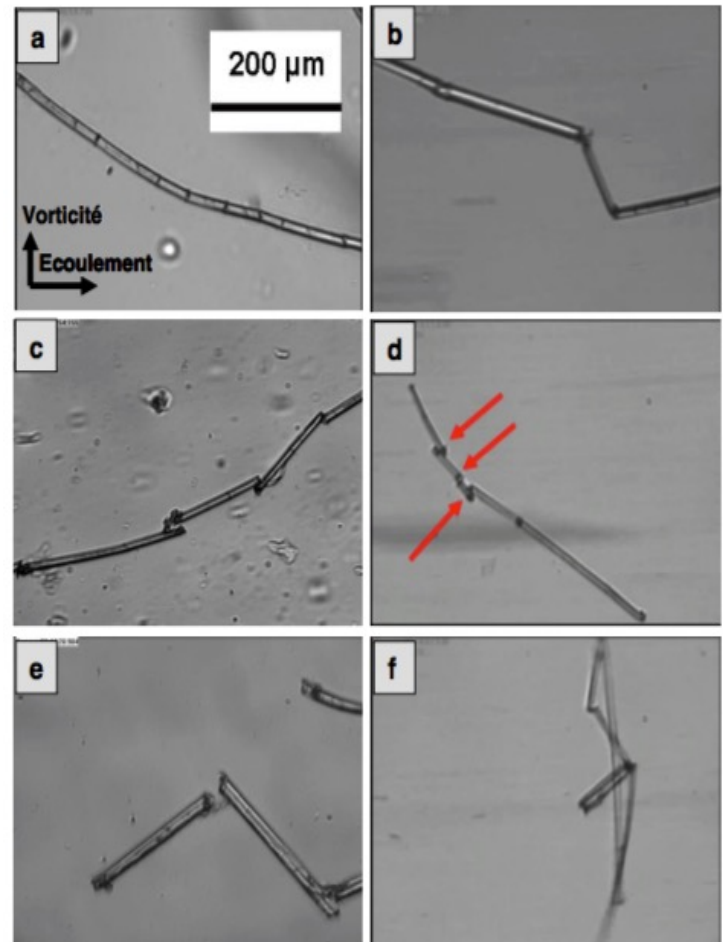
KINK BANDS



KINK BANDS



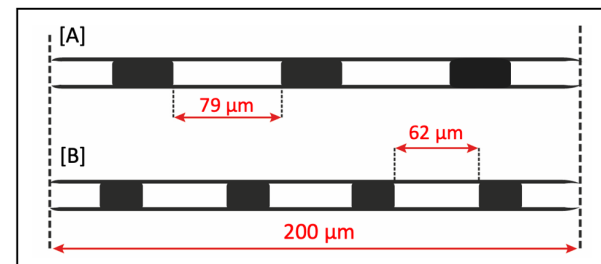
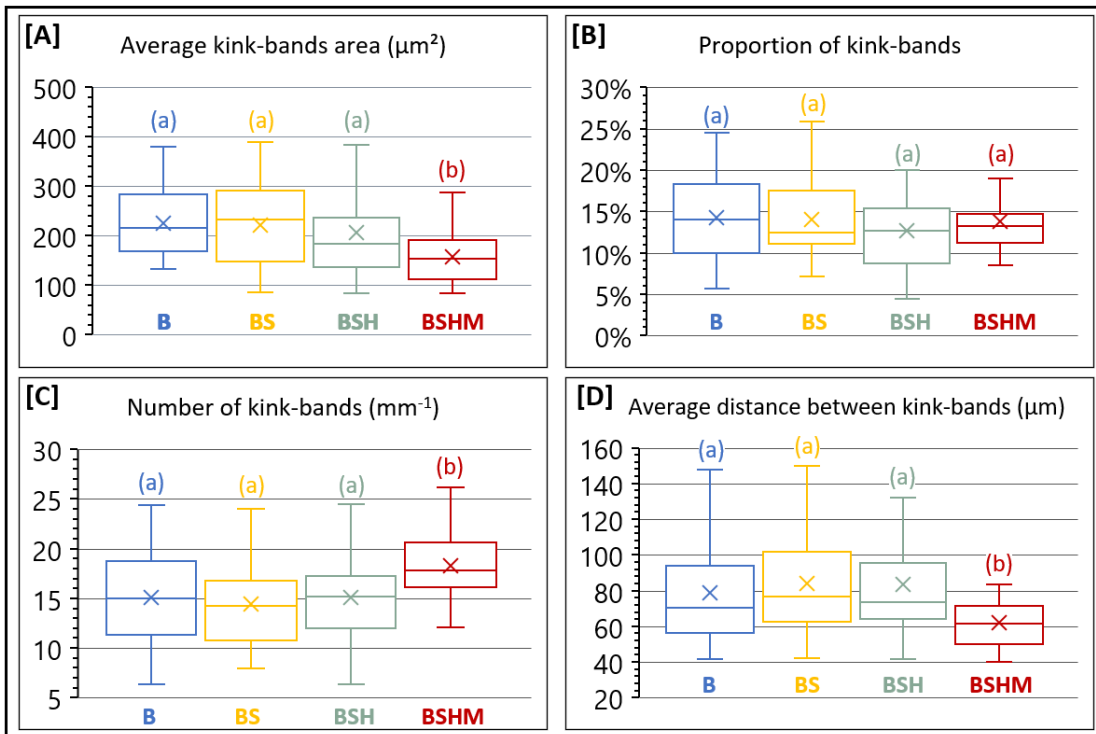
Le Duc, Thèse, 2014



Le Duc et al., Comp Part A, 2011

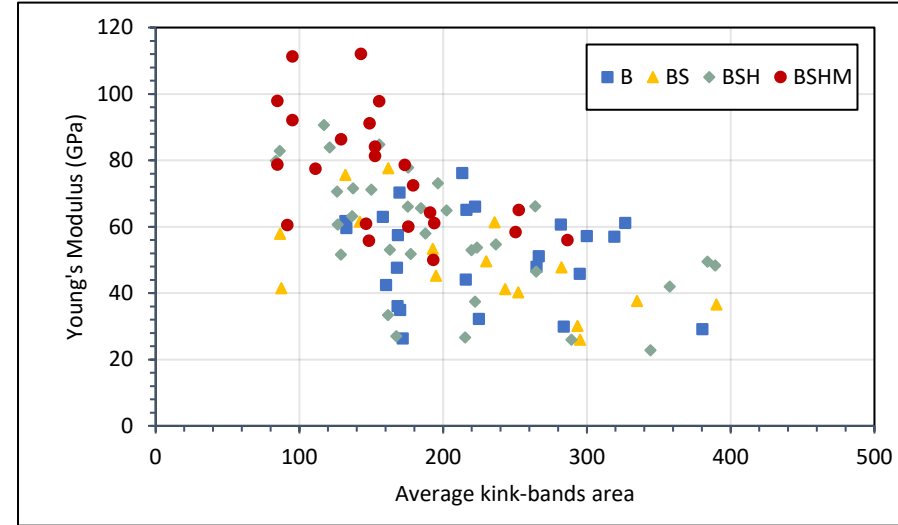
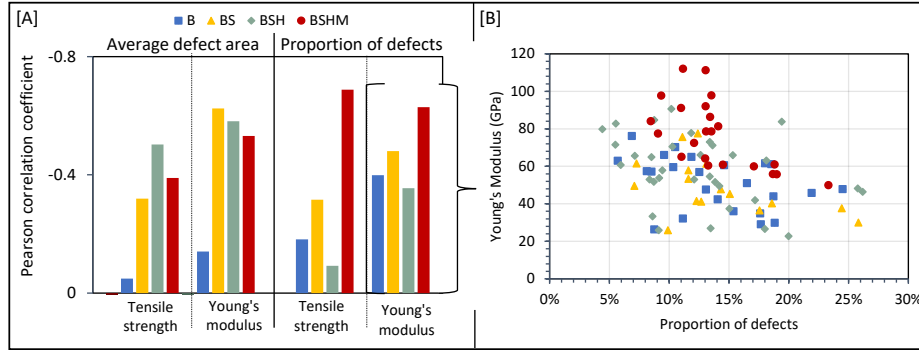


KINK BANDS





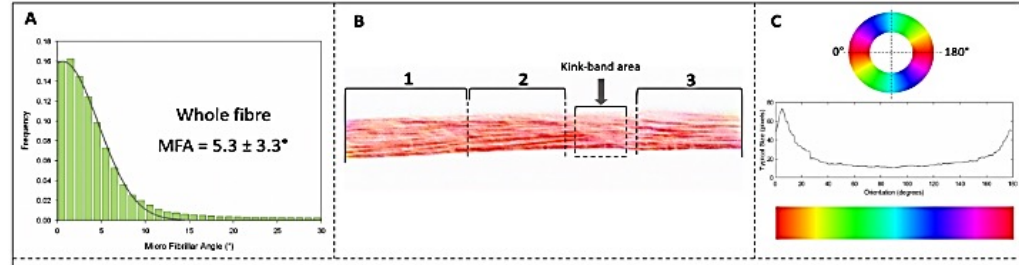
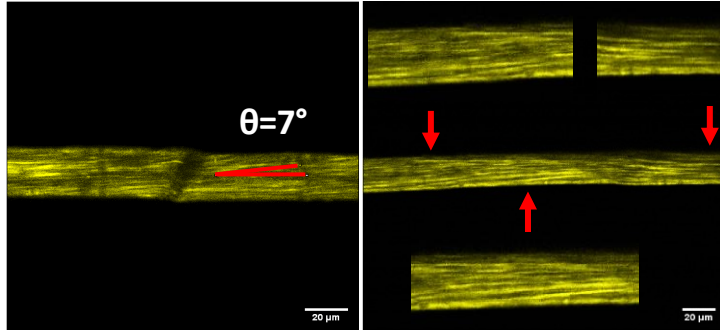
KINK BANDS – LINK WITH MECHANICAL PROPERTIES





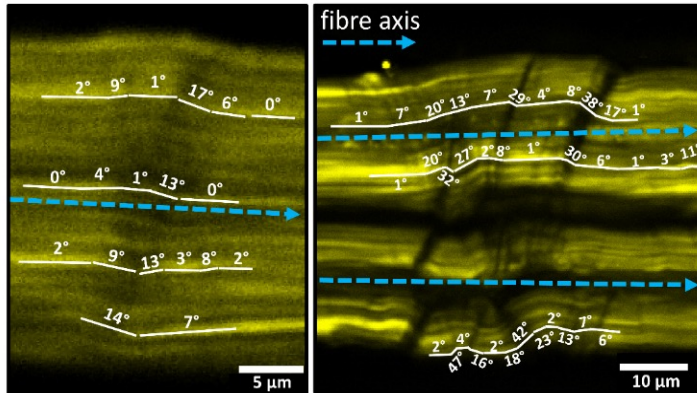
KINK BANDS – PROPERTIES AND STRUCTURE

Melelli et al., *Ind Crops Prod*, 2020

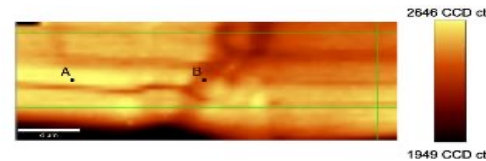


MFA of flax Bolchoi variety: $5.3 \pm 3.3^\circ$

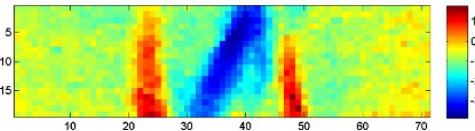
Melelli et al., *Ind Crops Prod*, 2021



Polarised Raman microspectroscopy on hemp

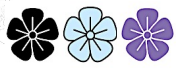


MFA



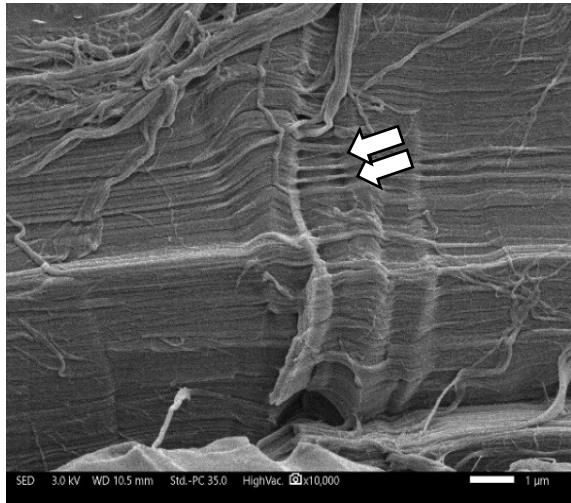
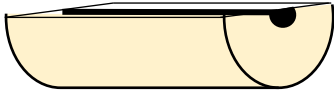
$+10^\circ$ -30°

Thygesen et Gierlinger,
J. Struct. Biol., 2013



KINK BANDS – PROPERTIES AND STRUCTURE

Fibre cut in a half



Detachment of macrofibrils because some macrofibrils less deviated than others -> **creation of cavities and pores**

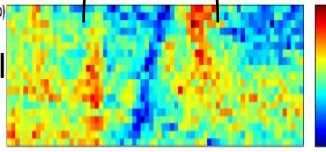
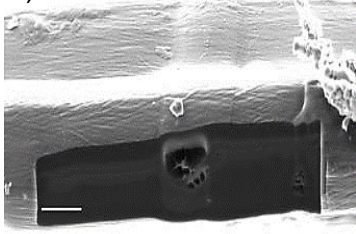
Thygesen et Gierlinger, *J. Struct. Biol.*, 2013

Crystallinity
calculated with Agarwal method
 I_{380}/I_{1096}

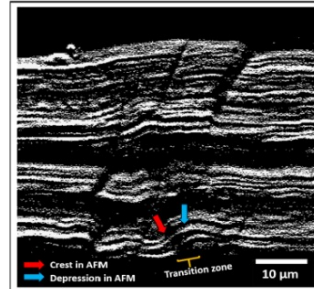
Zhang et al., *WCE*, 2015

FIB-SEM

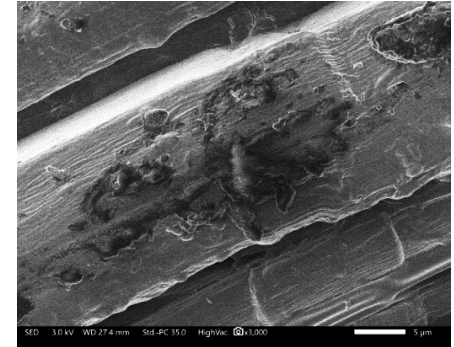
kink-band

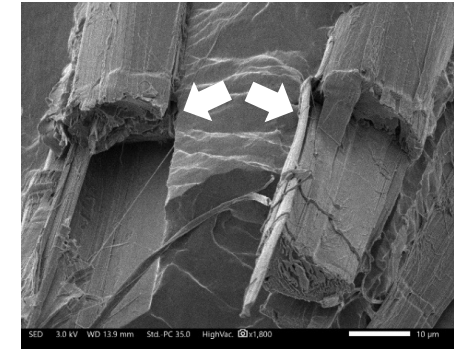
SHG



Biological colonization



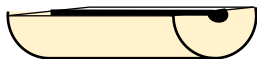
Highly prone to fracture





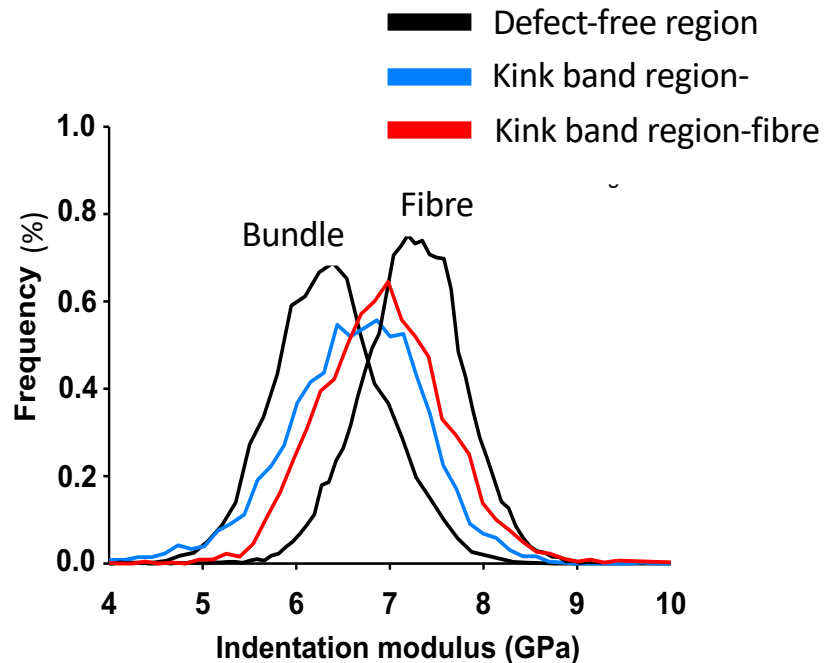
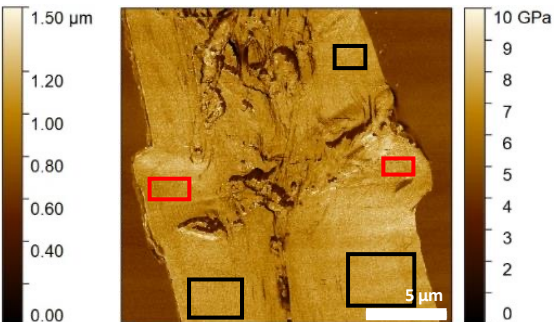
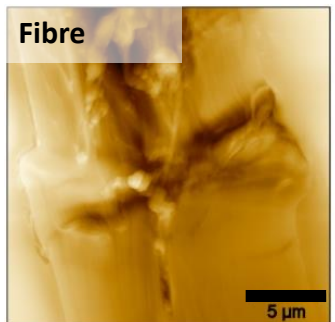
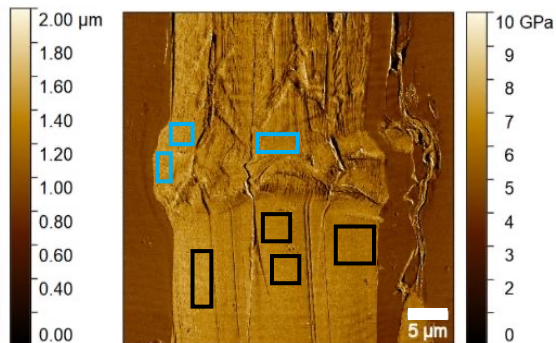
KINK BANDS – PROPERTIES AND STRUCTURE

Fibres cut in a half



TOPOGRAPHY

INDENTATION MODULUS



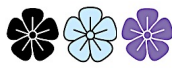
Stiffness:

Kink-band areas 6.9 ± 1.2 GPa

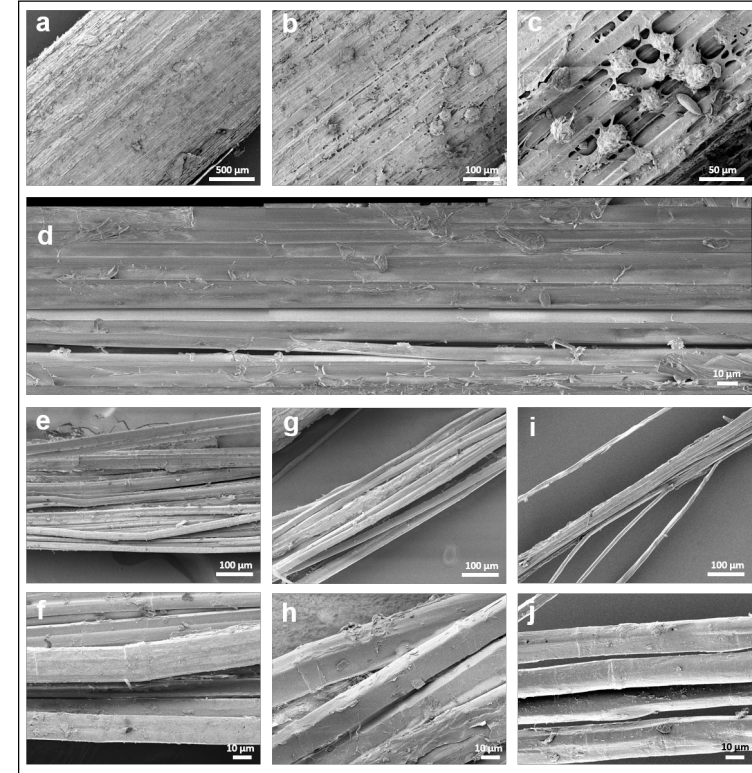
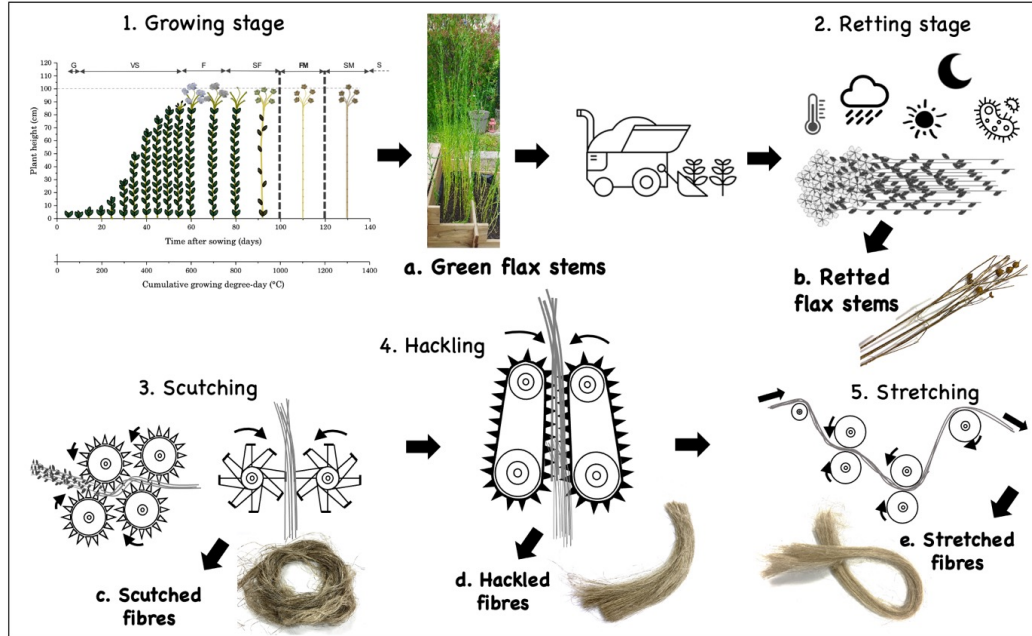
Defect-free regions 6.5 ± 1.5 GPa



SYNCHROTRON INVESTIGATIONS

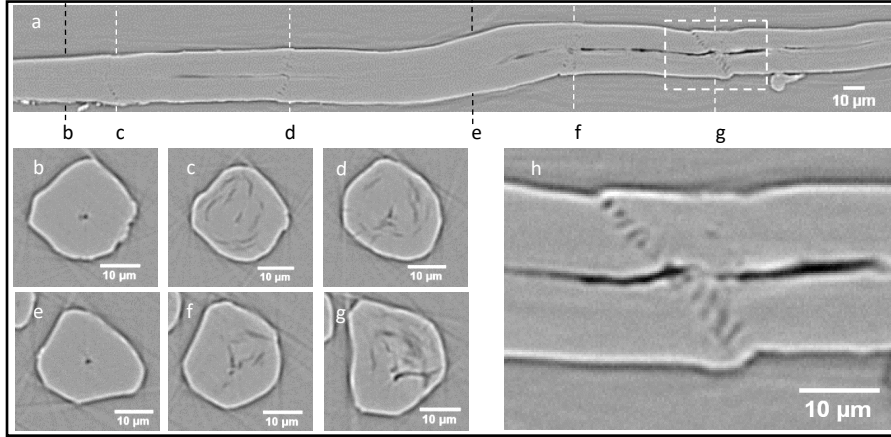


ELUCIDATING THE DEVELOPMENT OF KINK BANDS

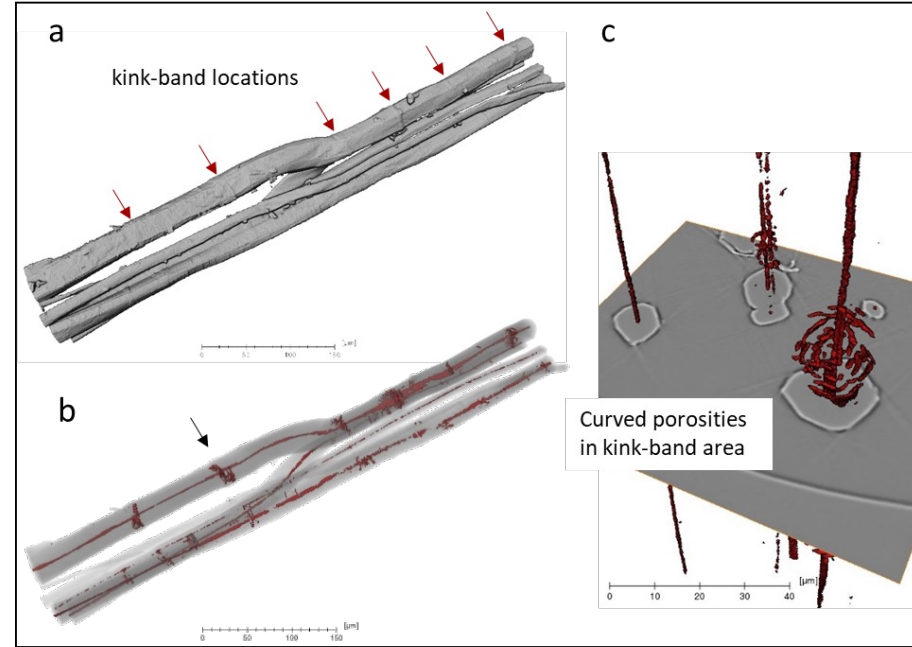


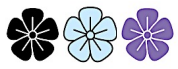


ELUCIDATING THE DEVELOPMENT OF KINK BANDS



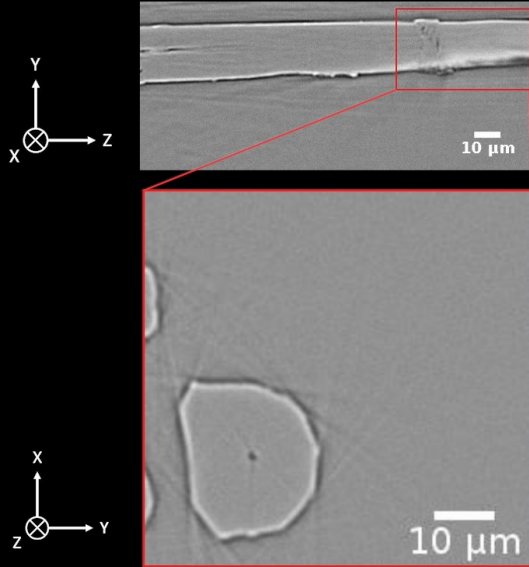
Observations on extracted fibres



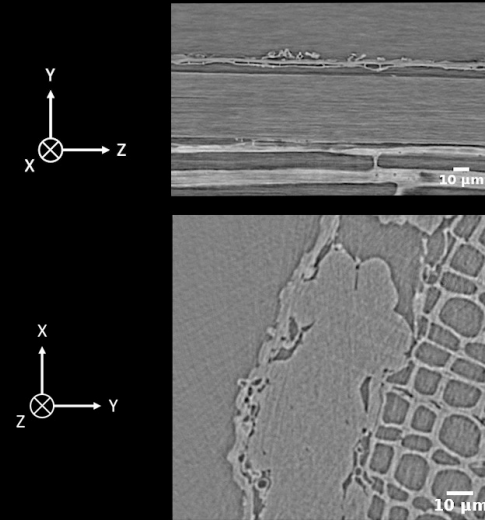


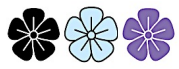
ELUCIDATING THE DEVELOPMENT OF KINK BANDS

Video 1: Stretched flax fibres



Video 4: Retted stem





COMPLEX STRUCTURE OF KINK BANDS

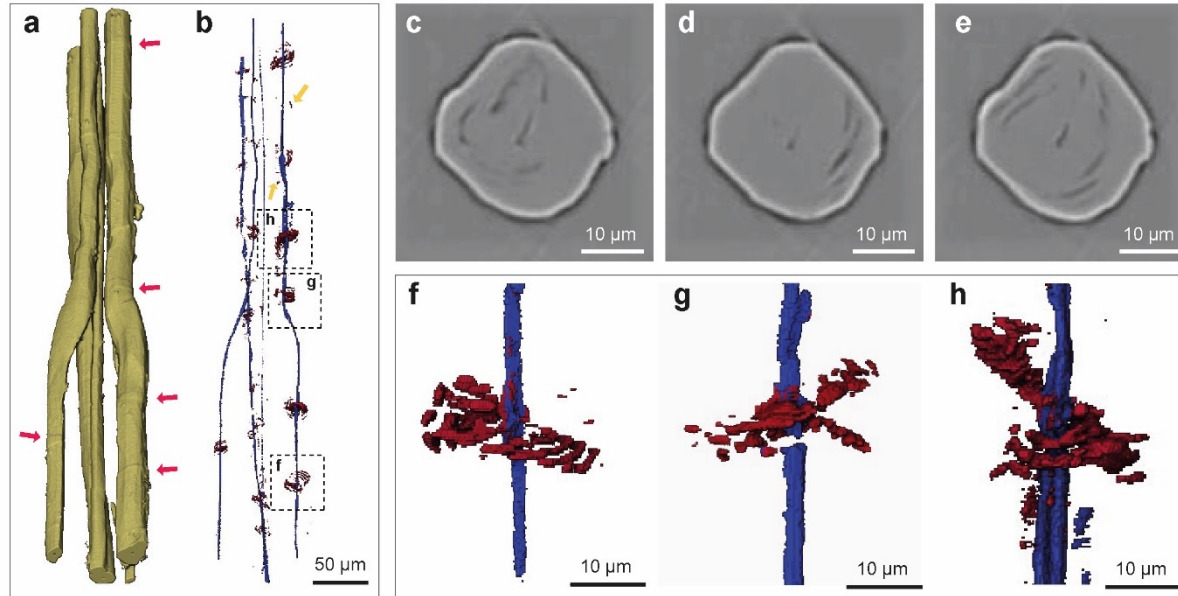
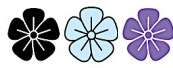


Figure 1. 3D reconstruction of the fibre, before (a) and after (b) segmentation, tomographic slice images of fibres along transverse planes where voids can be seen as dark shapes (c, d, e), 3D reconstruction of porosities in red around the lumen in blue



COMPLEX STRUCTURE OF KINK BANDS

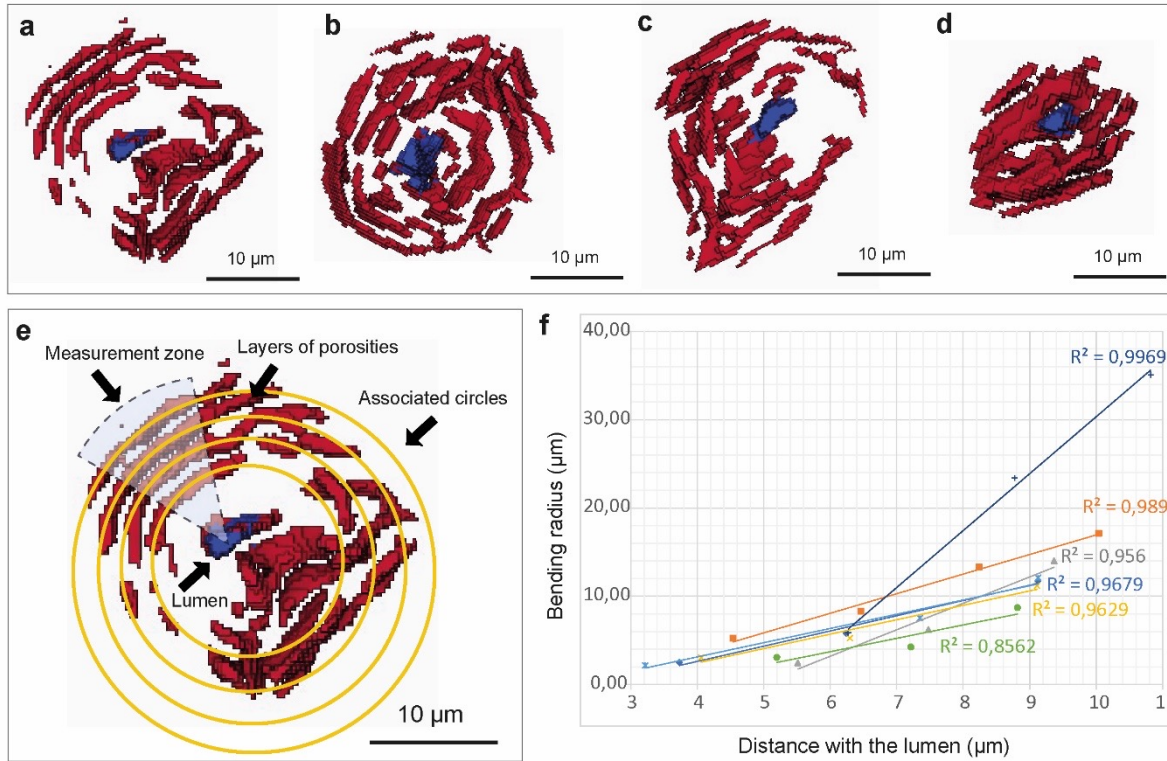


Figure 2. Defects viewed from the Z axis, in blue the lumen can be seen, and the pores layers are in red showing a distinct onion layer organisation (a-d), detail of the measurement method for the curvature radius (e), linear regression of the radius of curvature of the pores layers evolving with the distance with the lumen axis (f)

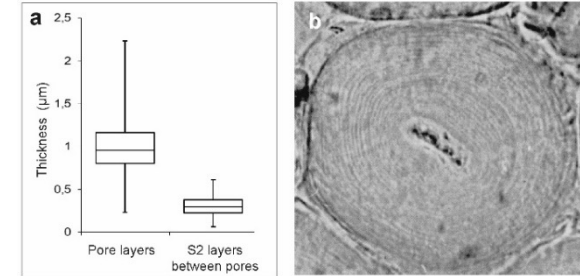
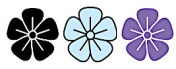
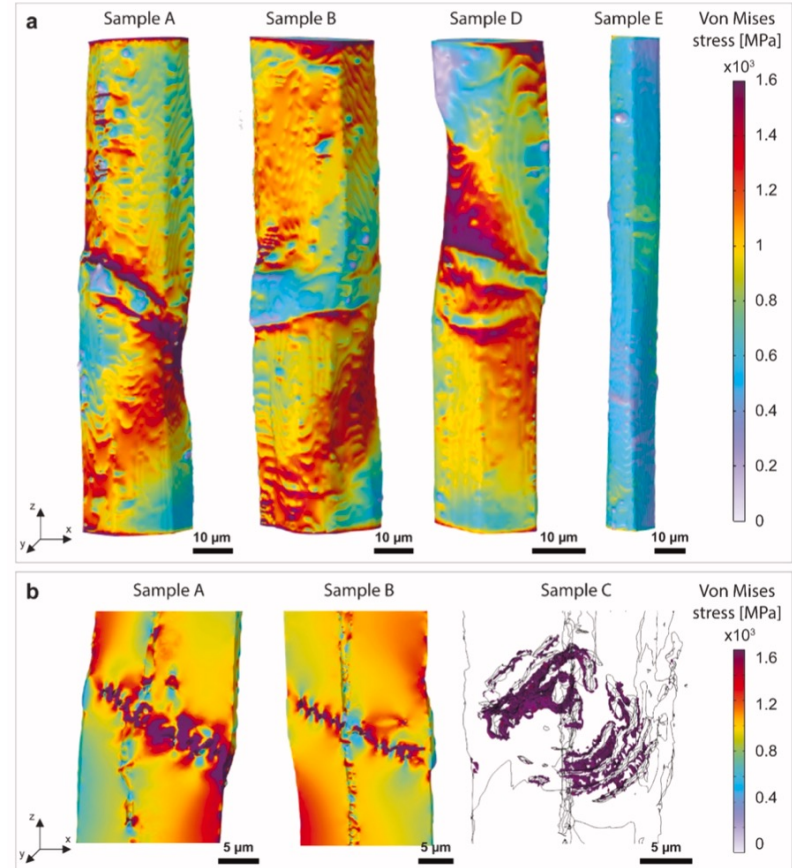
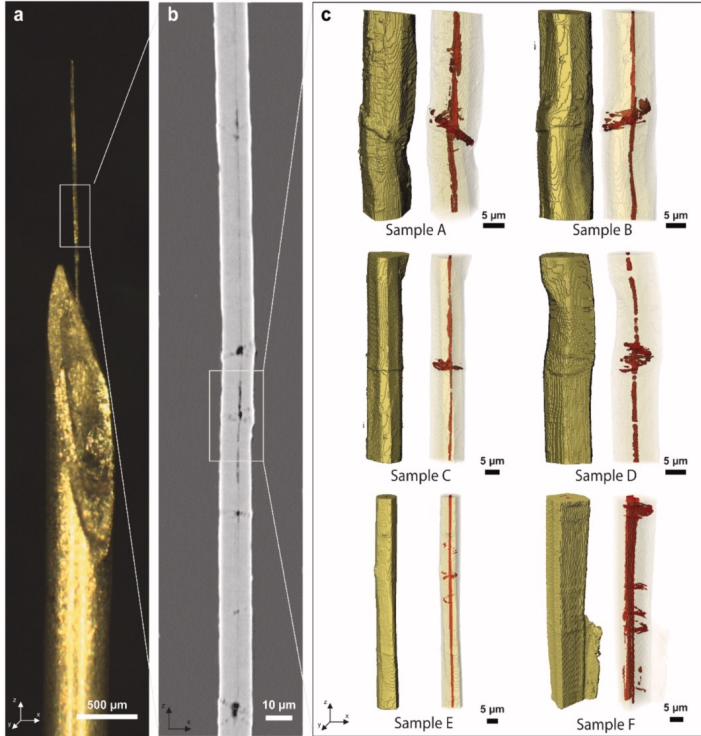


Figure 3. Box-plot presenting the pores thickness and the distance between two pores (a), cross-section of a flax fibre evidencing the successive cellulose layers (b) (Hock, 1942)

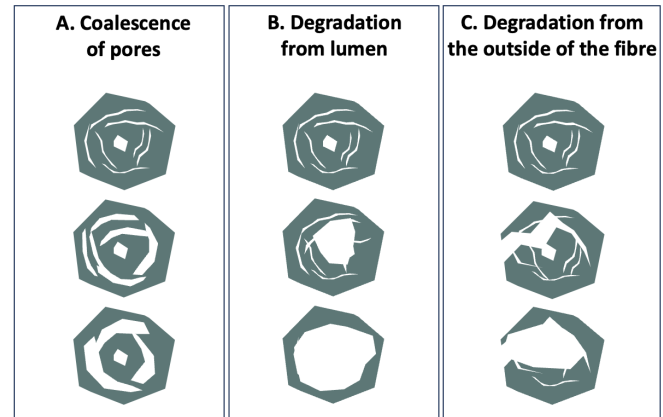
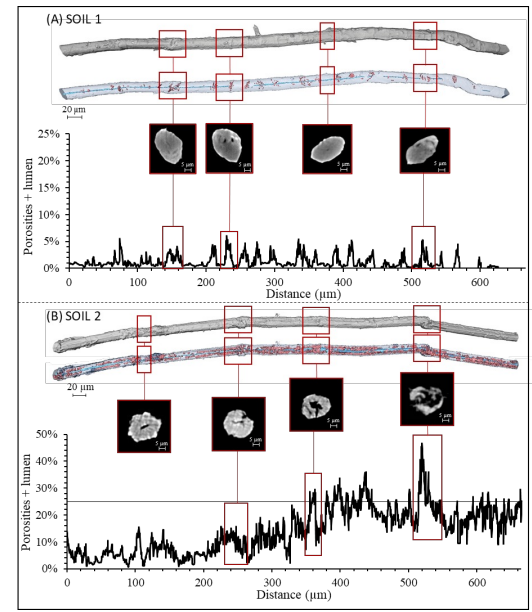
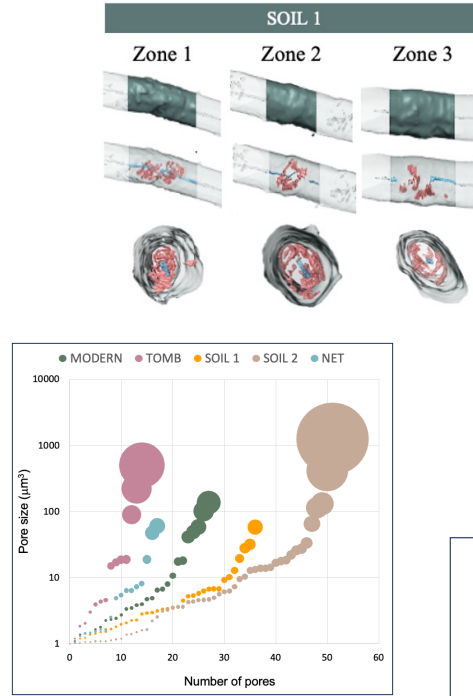
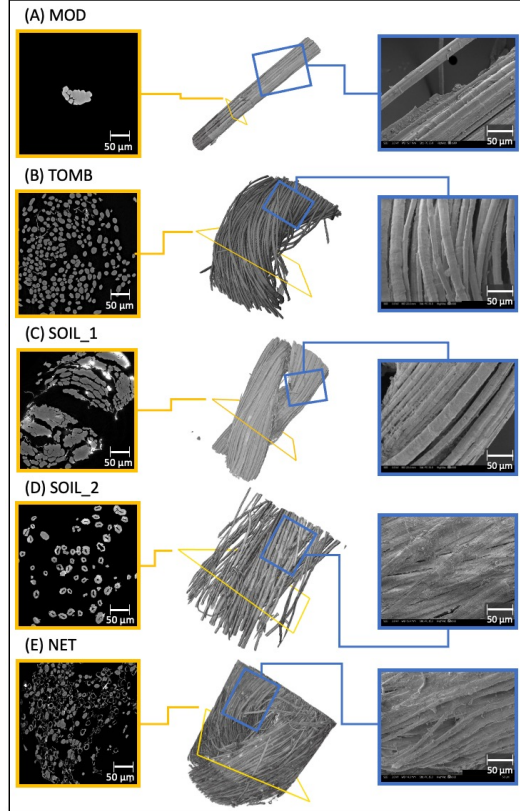


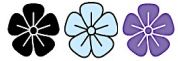
COMPLEX STRUCTURE OF KINK BANDS





KINK-BANDS AND DURABILITY

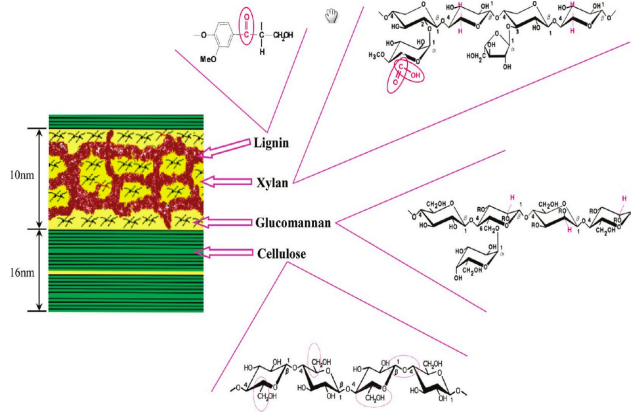




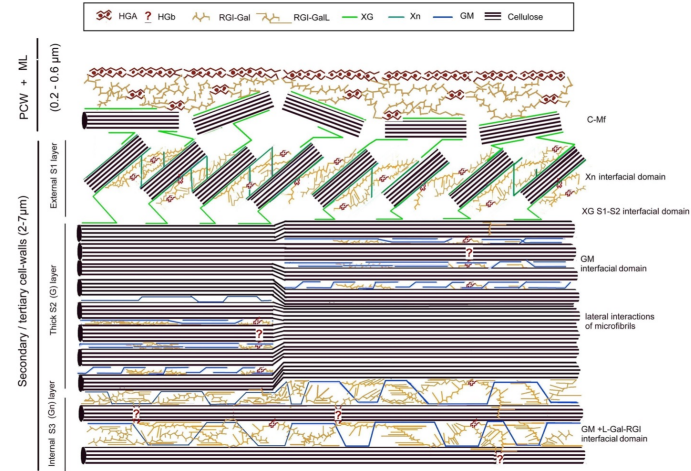
**IMPACT OF PROCESS ON PLANT FIBRES AND
COMPOSITES PROPERTIES
CASE OF INJECTION AND EXTRUSION**



PLANT FIBRES ARE SPECIFIC AND SENSITIVE



Yin et al. Biomacromolecules, 2011



Rihouey et al, Cellulose, 2017

Wood S₂

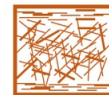
50% cellulose
25% hemicellulose
25% lignin
+ extractives, ashes
MFA = (0) 8-40°

Flax G

80% cellulose
20% hemicellulose
≈0% lignin
+ protein
MFA = (0) 5-10°



PROCESSING: AN AGGRESSIVE STAGE



Injection

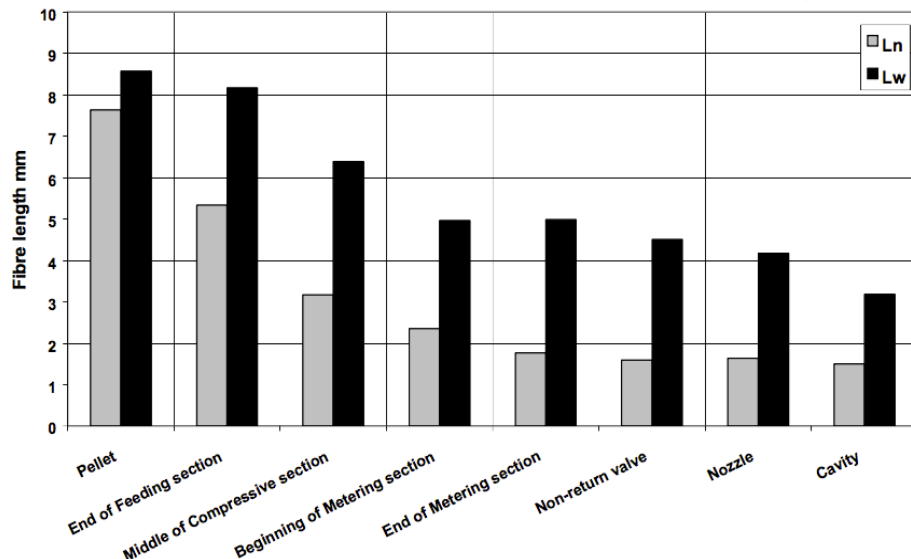
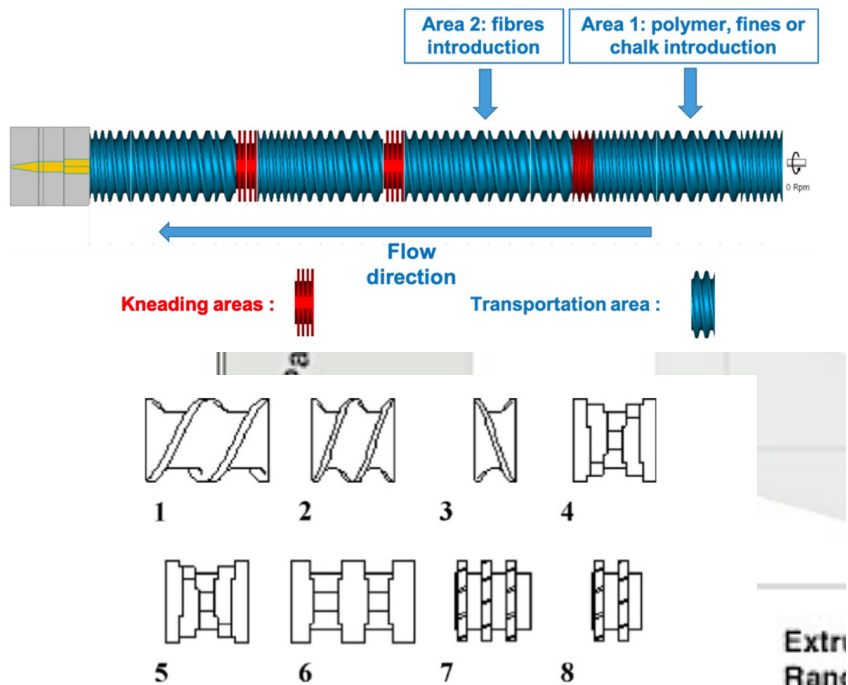


Fig. 2. Screw elements for twin-screw extruder ZE25 from Berstorff; conveying elements (1 and 2), back-conveying element (3), kneading elements (4-6), mixing elements (7 and 8).

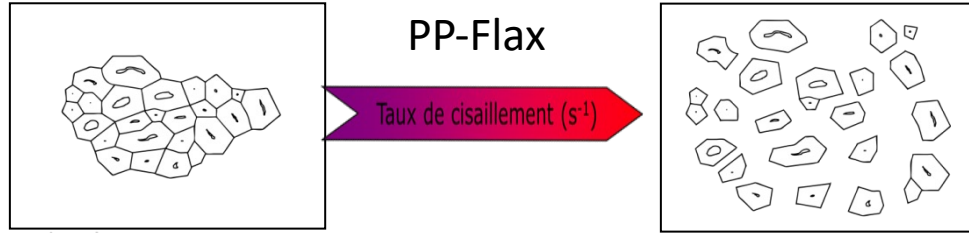
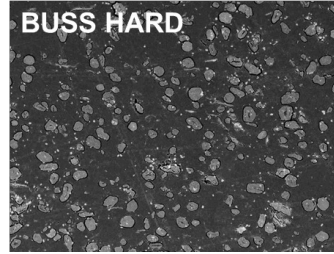
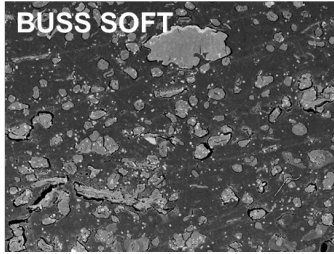
COMPOUNDING

SHEAR RATE, 1/s

INJECTION



IMPACT OF PROCESSING TOOL



R = 0,79

R = 1,34

Compounding: 2 ways

Buss Comalaxor (soft)



Buss Comalaxor (hard)



	Buss Soft	Buss Hard
Dispersion factor	0,79	1,34

Increase of dispersion,
reduction of bundles
=> Improvement of
mechanical properties

	Buss Soft	Buss Hard
Stress (MPa)	23,8	28,1

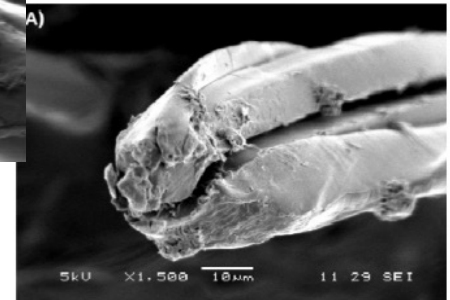
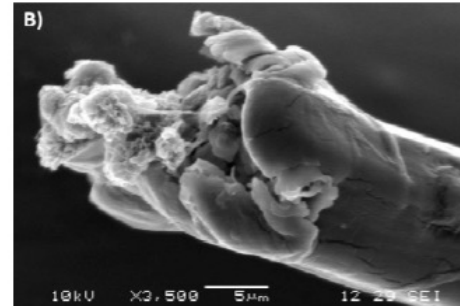
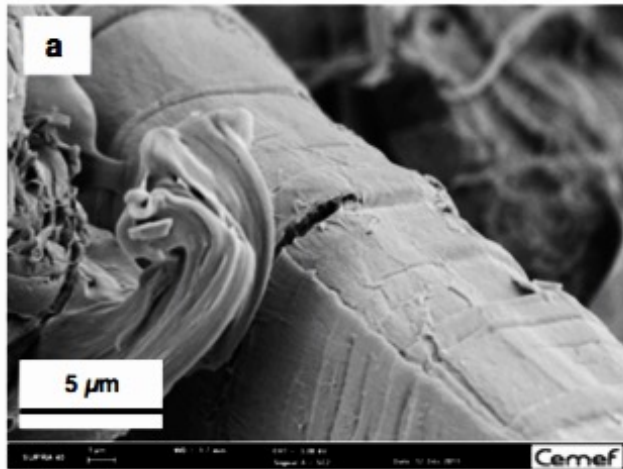
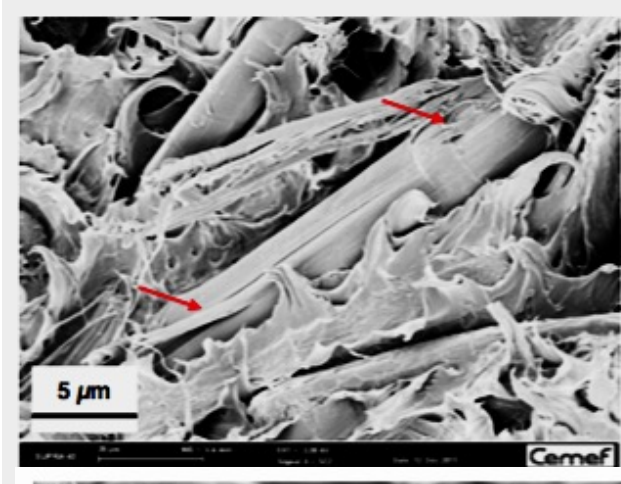


PROCESSING: AN AGRESSIVE STAGE



Injection

Baley,
Macromol
Symp,
2005



Coroller, Phd Thesis,
IRD, 2013

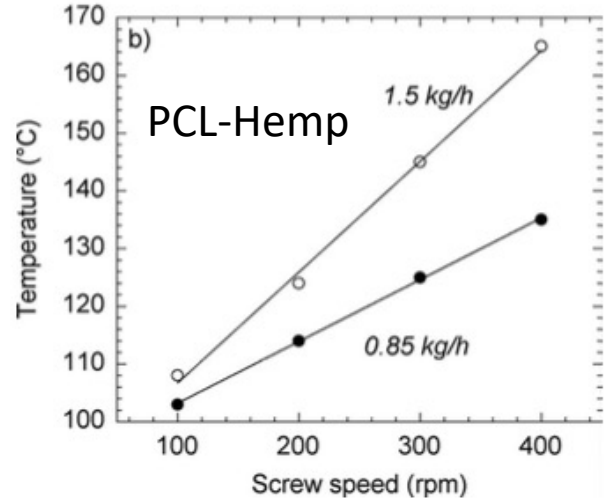
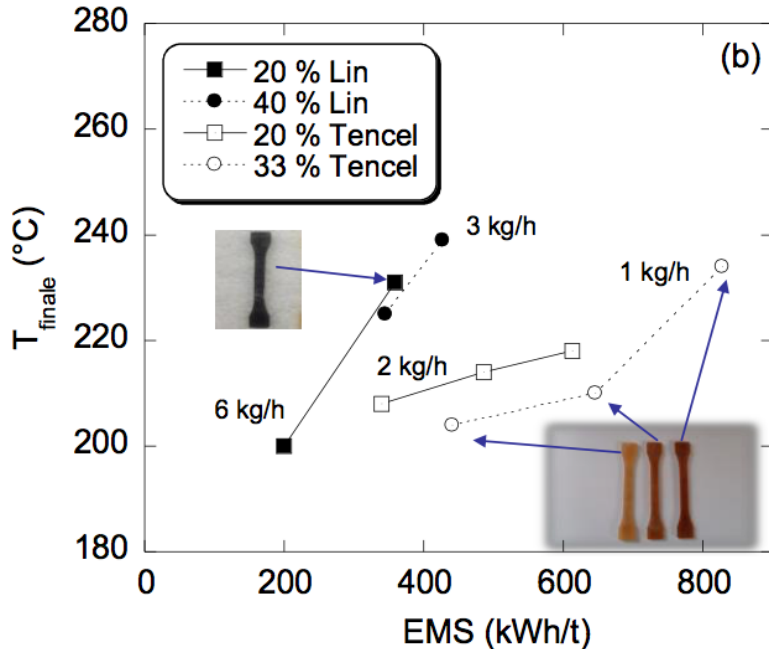


PROCESSING: AN AGRESSIVE STAGE



Temperature is related to the polymer properties but only defining a temperature is too simplistic

Self-heating



Beaugrand & Berzin, JAPS, 2012

Importance of the fibre content on the *in-tool* temperature

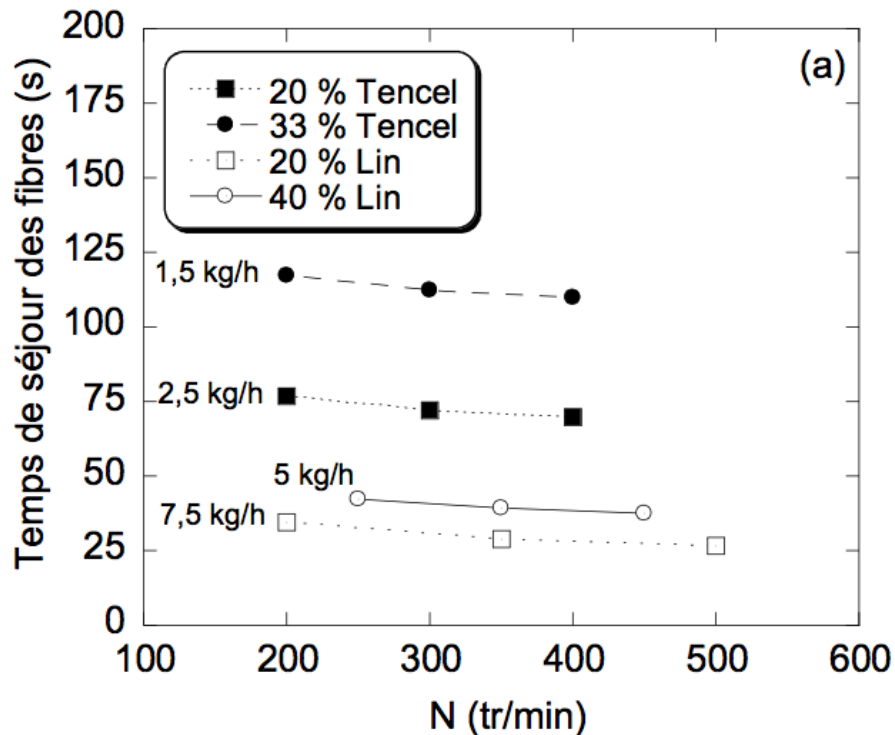


PROCESSING: AN AGRESSIVE STAGE

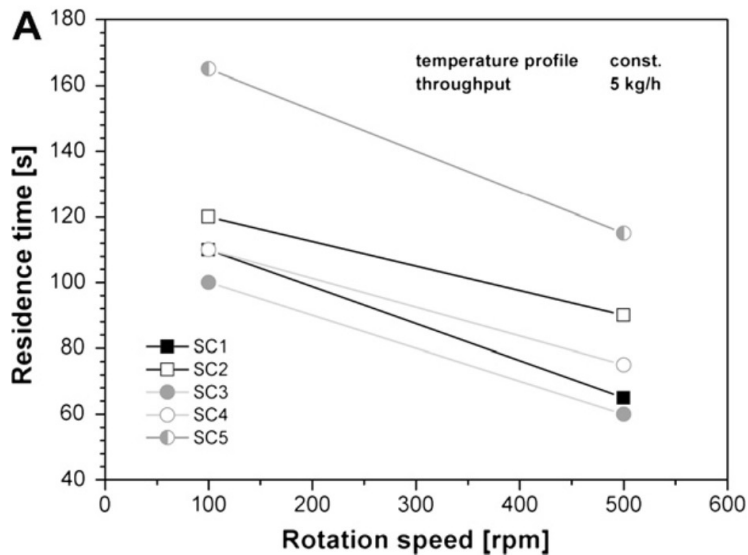


Very different residence times for different tools

Villmow et al., Comp Sc & Tech, 2010



Le Duc, Thèse, 2014



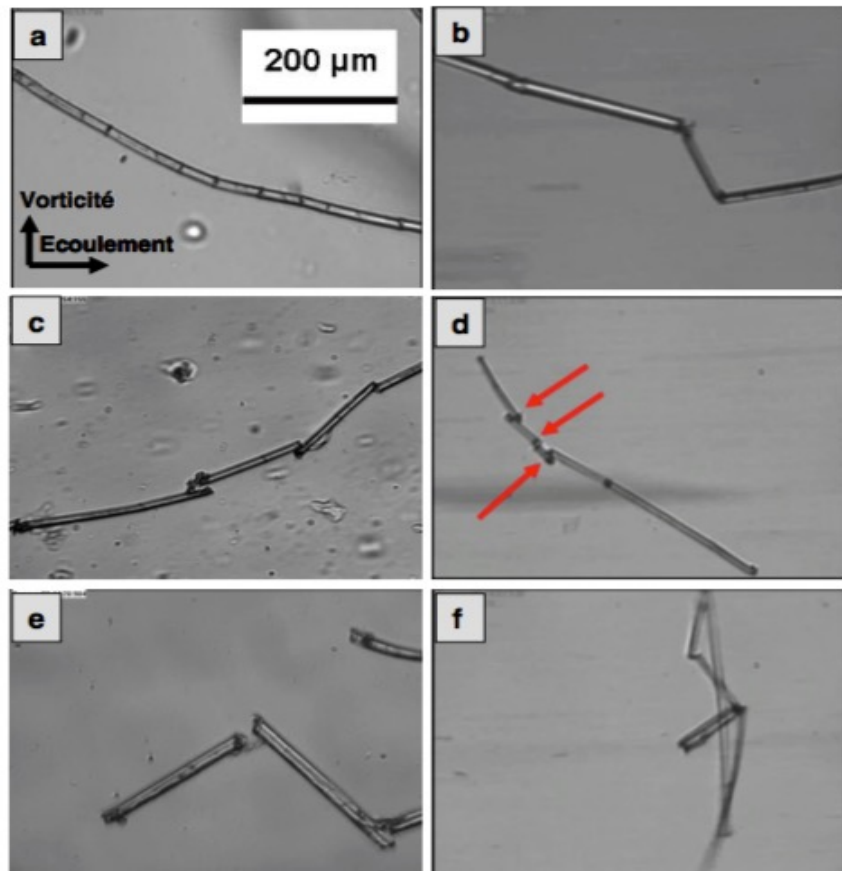
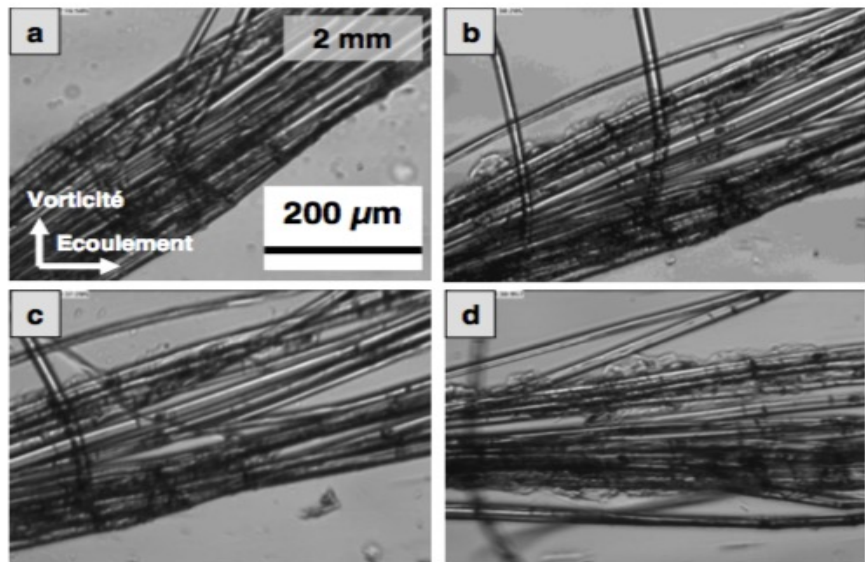
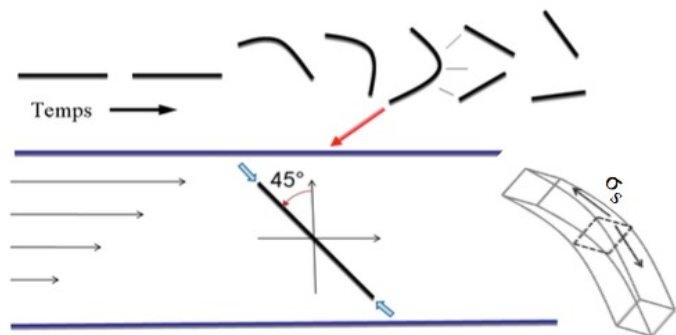
BUSS: 100 to 200 s

Twin screw: 10 to 180 s

Injection: 30 to 300 s

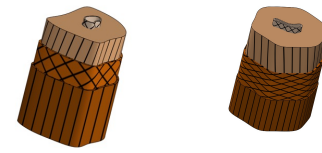


PROCESSING: MORPHOLOGICAL DAMAGES

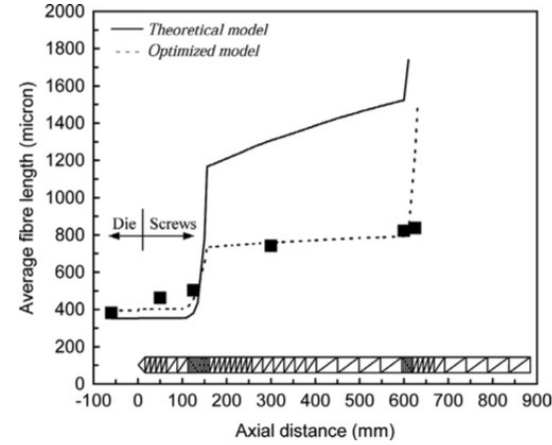
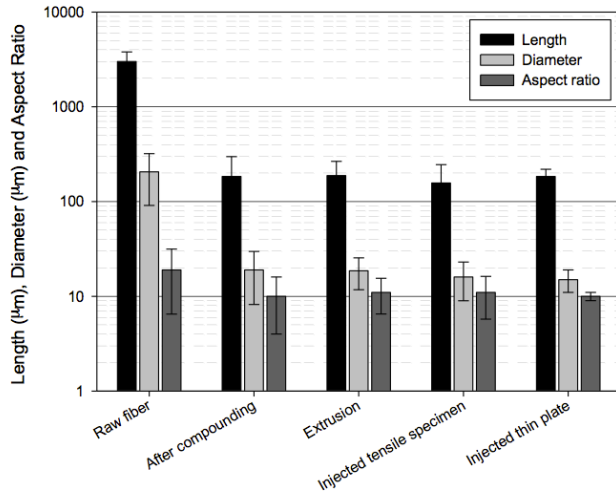
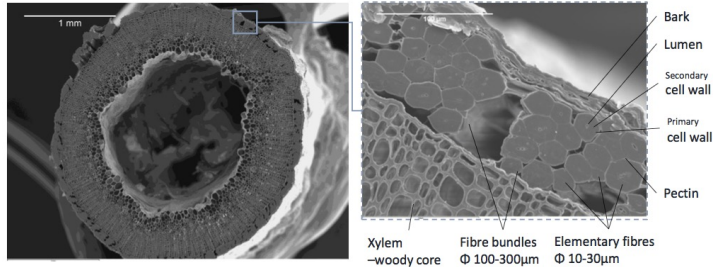




PROCESSING: MORPHOLOGICAL DAMAGES

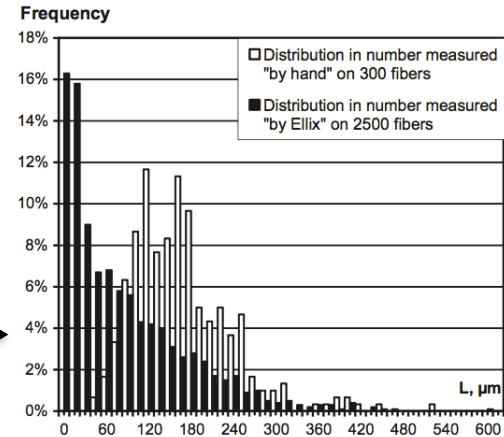


Zheng et al.,
Comp Part A,
2014



Berzin &
Beaugrand, Comp
Part A , 2014

Specificity of plant fibres: fibrillation and appearance of fine particles



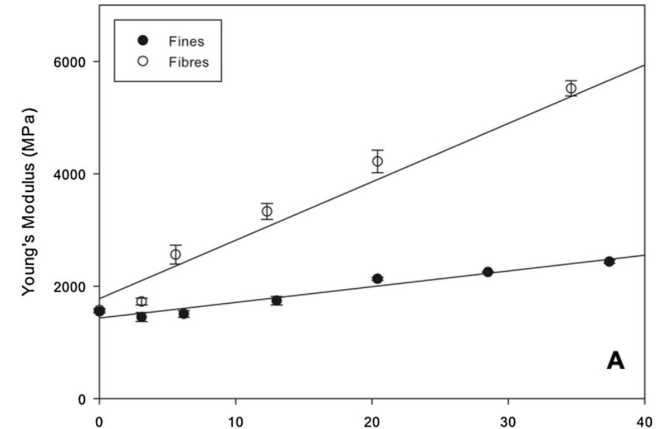
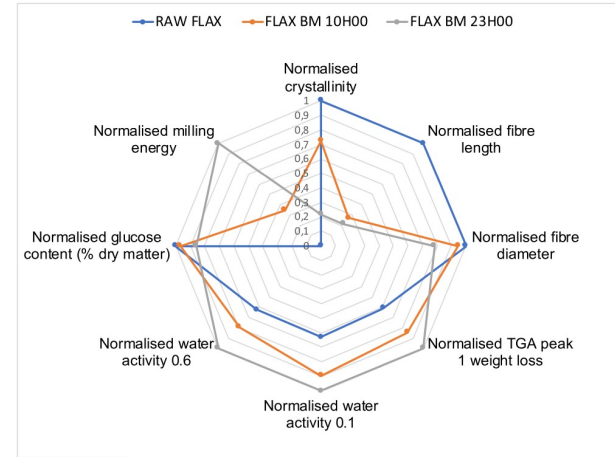
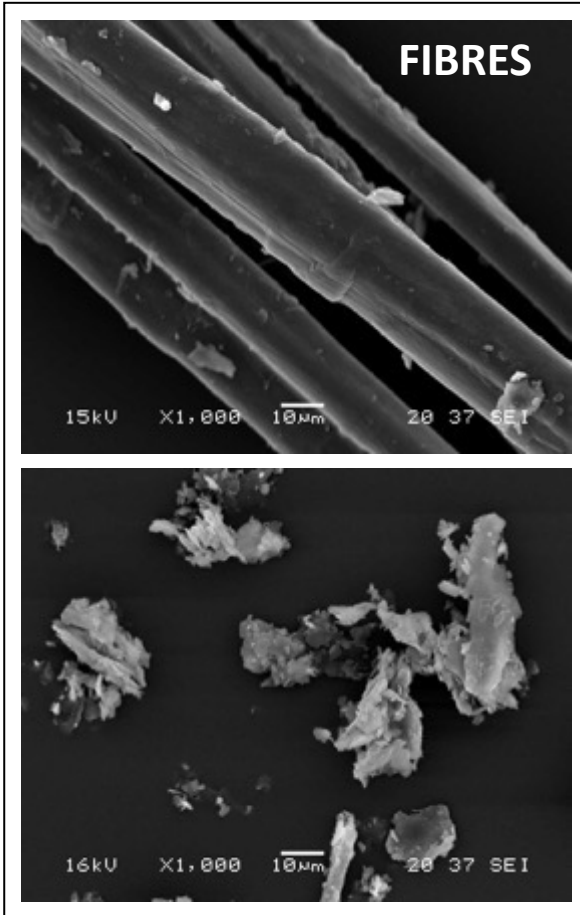
Le Moigne et
al., Comp Part A
, 2011



PROCESSING: MORPHOLOGICAL DAMAGES



Injection

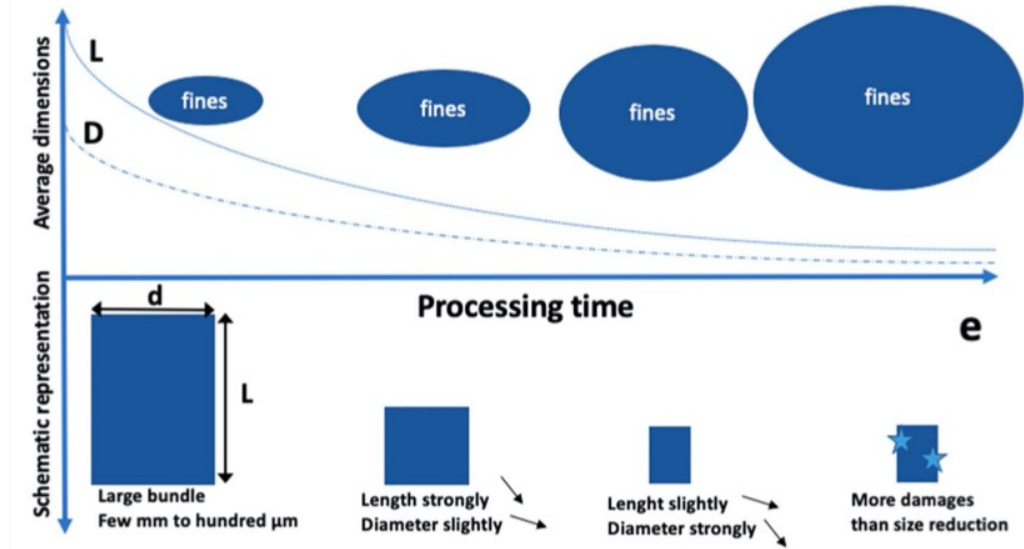
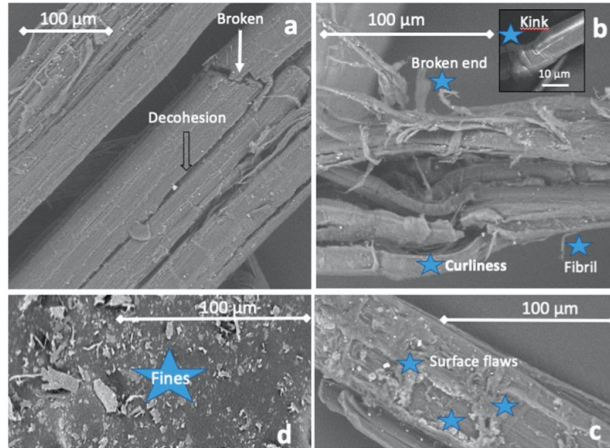
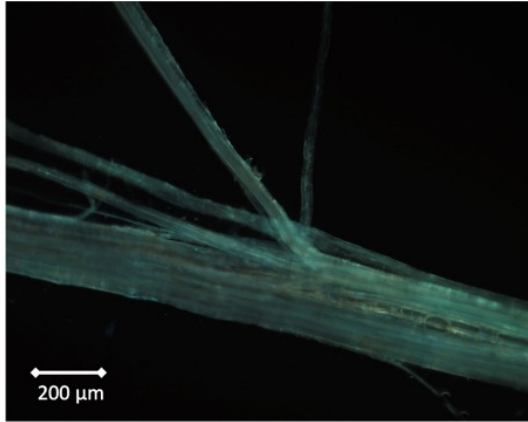




PROCESSING: MORPHOLOGICAL DAMAGES

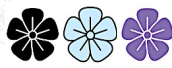


Injection



Plant fiber- breakage mechanism during a process cycle

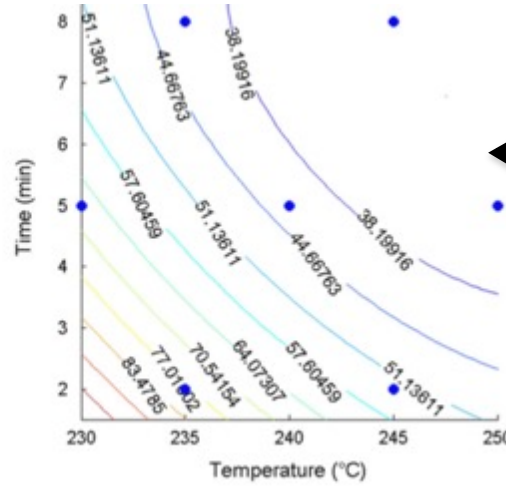
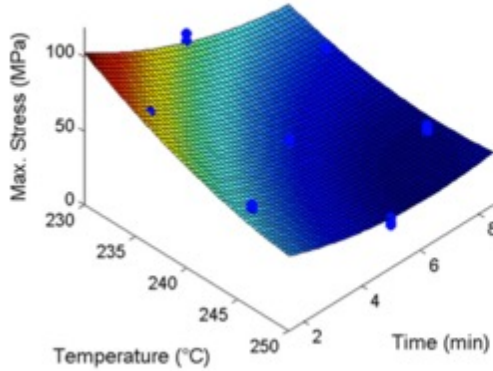
Similar behaviour after several process or recycling stages



IMPACT AT COMPOSITE SCALE



PA6-Flax



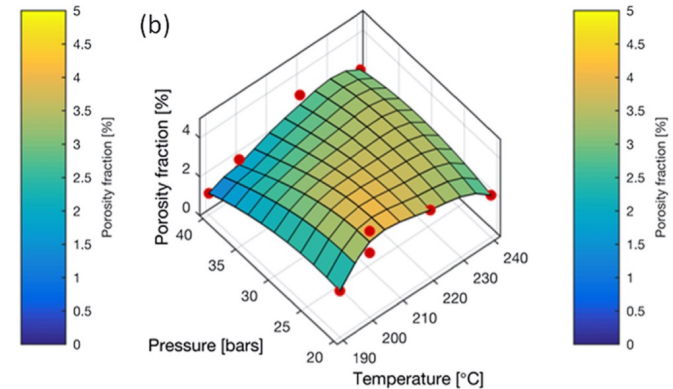
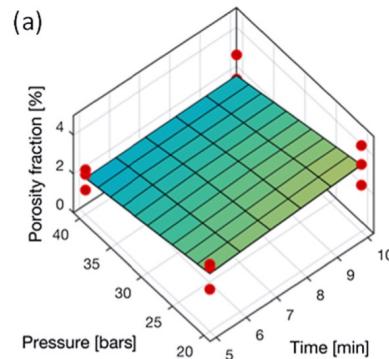
Time and temperature are impacting on composite properties: exemple of stress (Mpa)

Liang et al., J Mat Sc, 2015

Pressure has also a great impact on porosity fraction



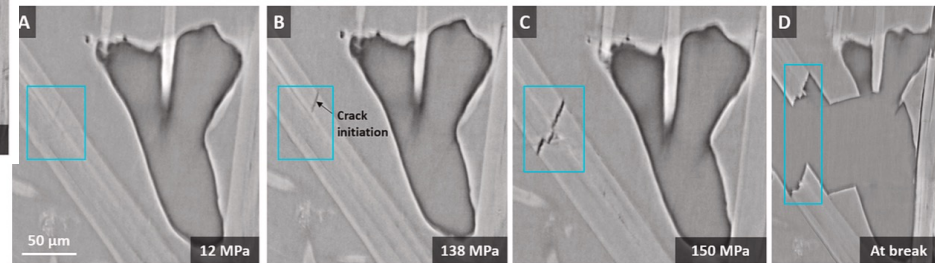
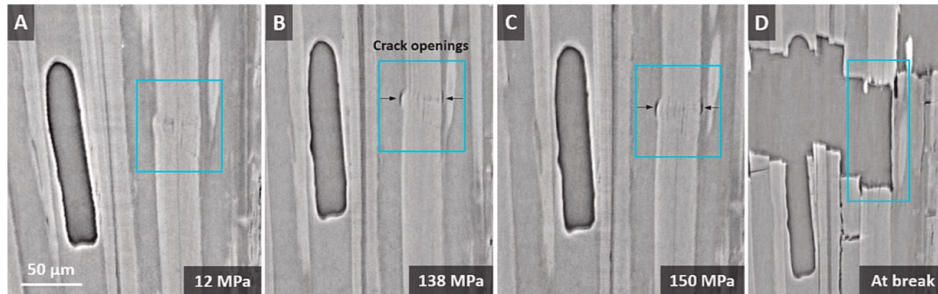
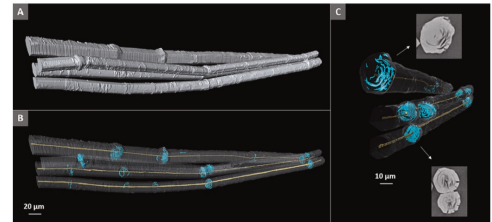
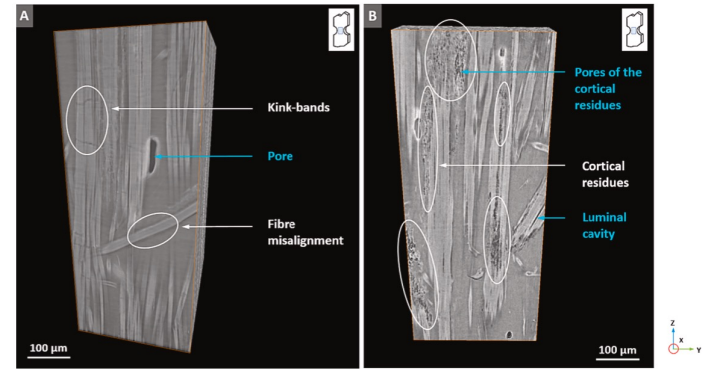
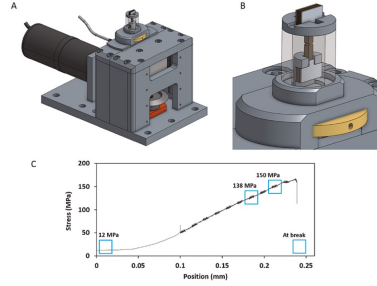
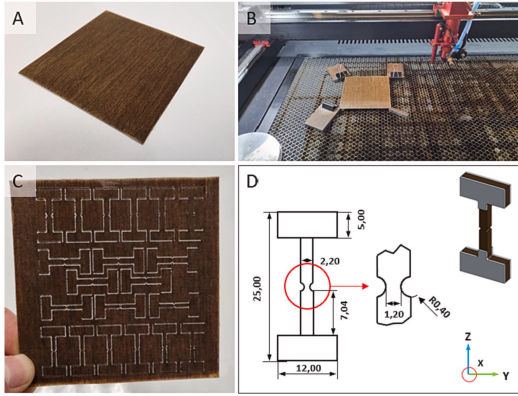
PP-Flax



Ramakrishnan, Le Moigne et al., Comp Part A, 2019

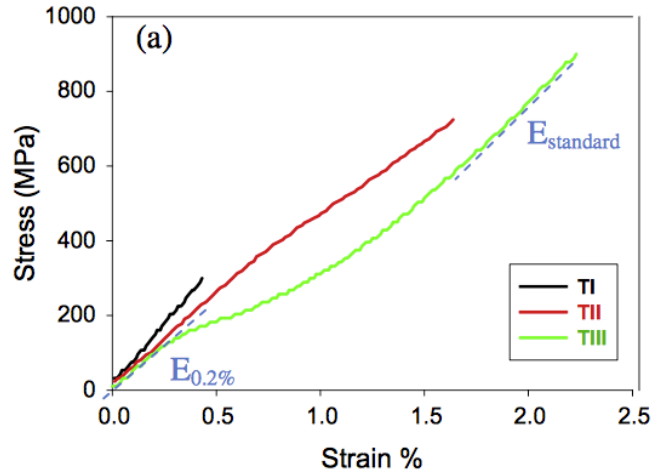
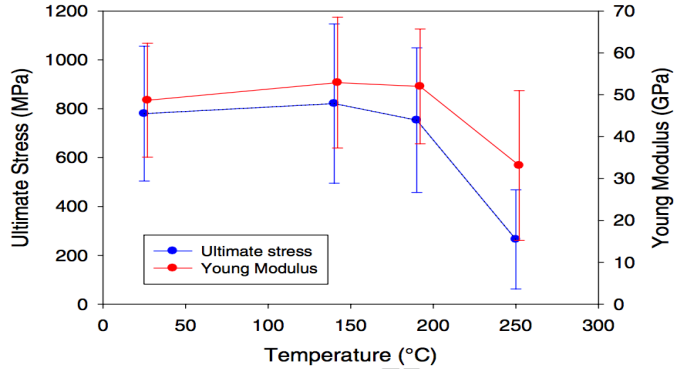


IMPACT AT COMPOSITE SCALE

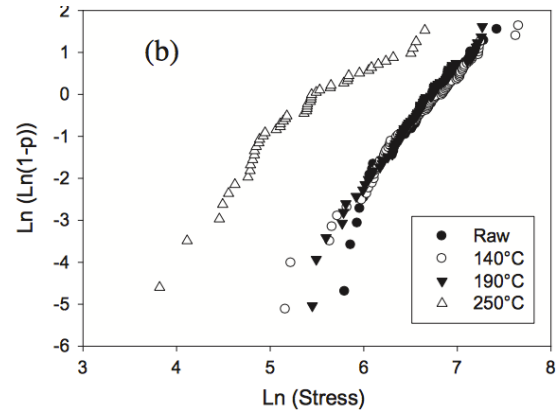
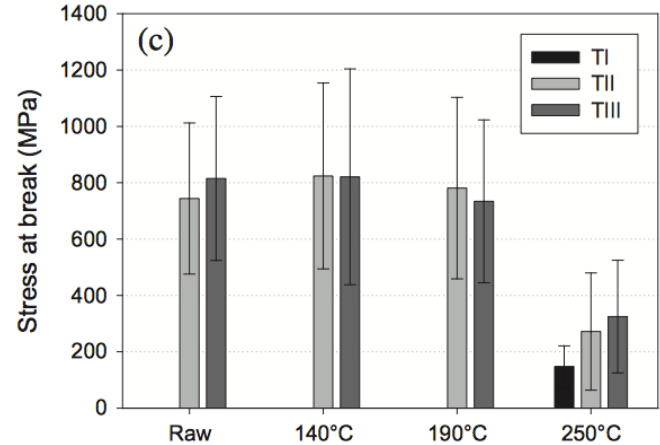




IMPACT AT FIBRE SCALE

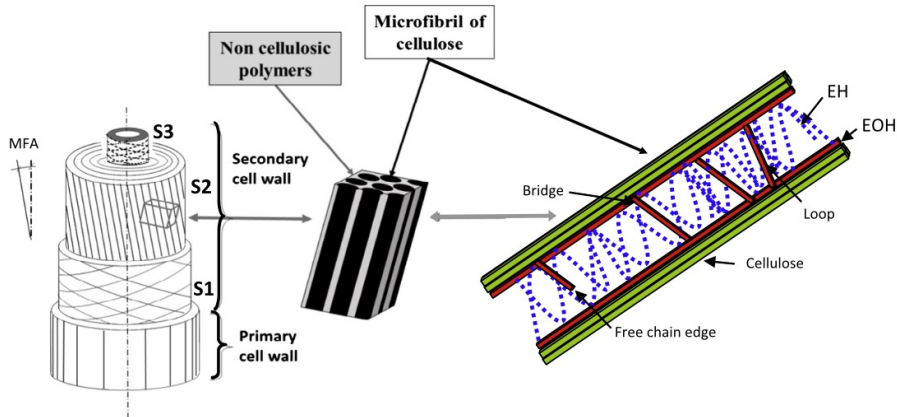
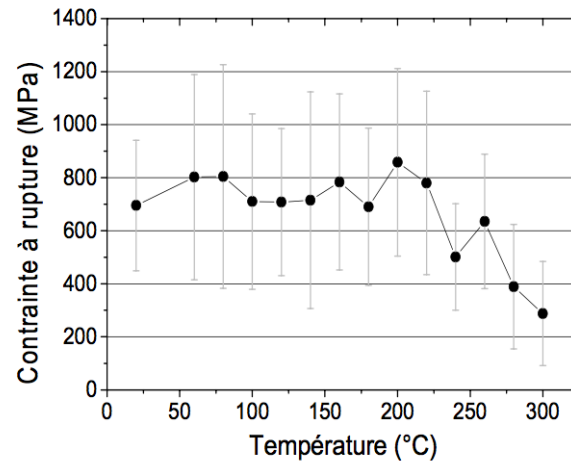
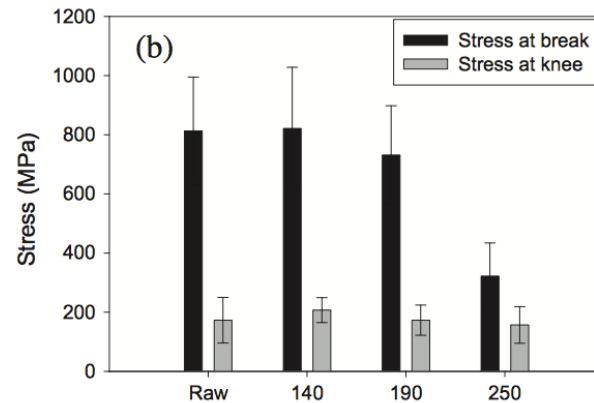
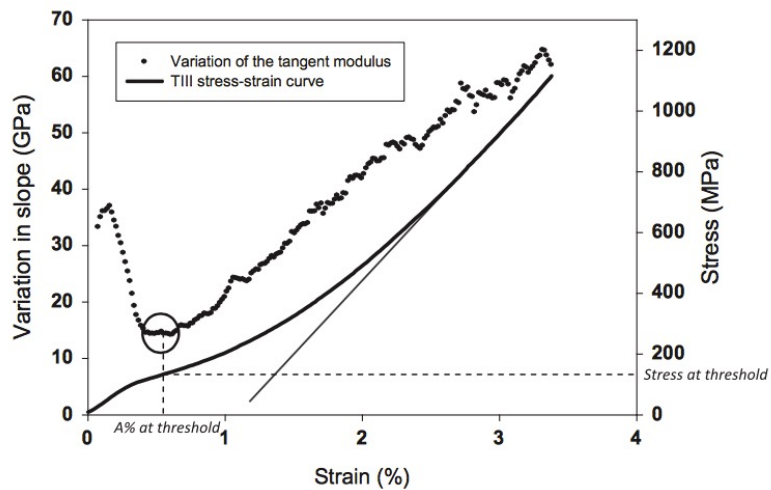


Irreversible damage to the fibre



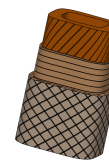


IMPACT AT FIBRE SCALE

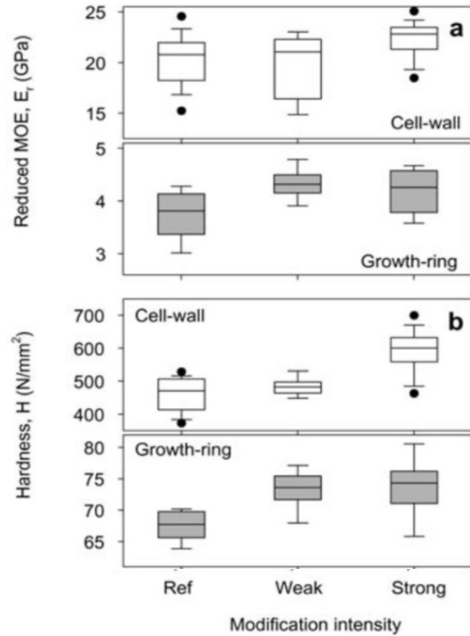




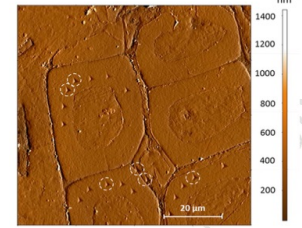
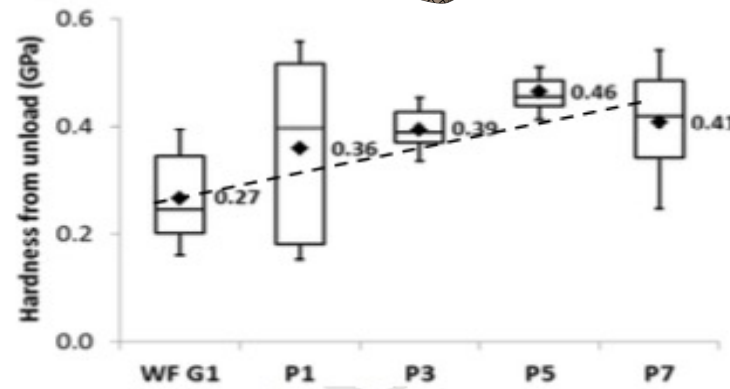
IMPACT AT CELL WALL SCALE - WOOD



Injection

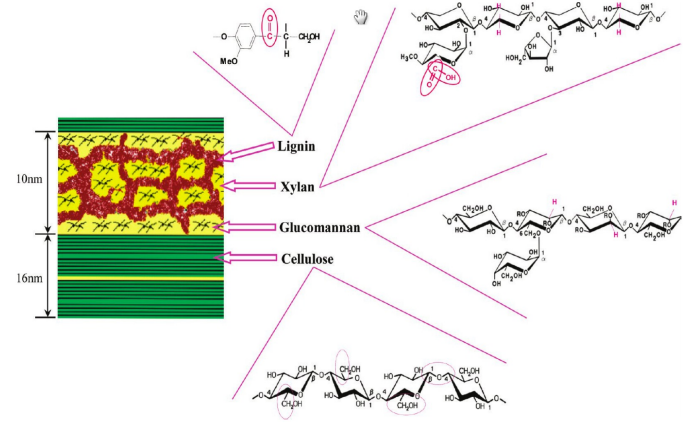


Stanzl-Tschegg et al. Holzforschung, 2009



Soccalingame et al. Pol Deg & Stab, 2015

Hypothesis: cross-linking of matrix components (cellulose-xylan-lignin bonds)



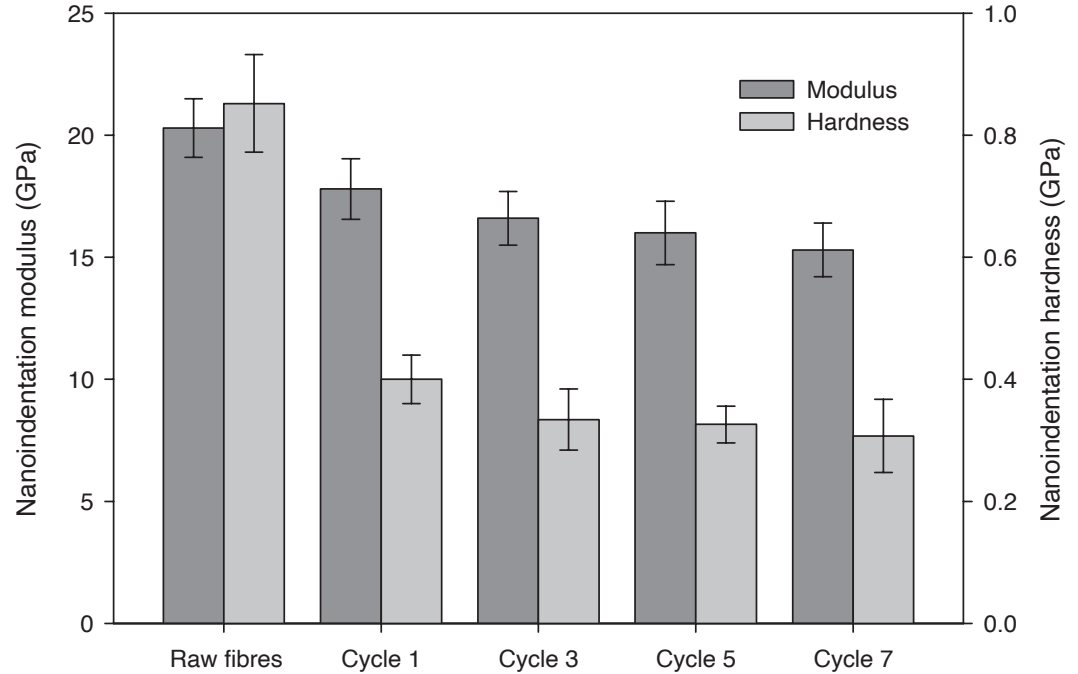
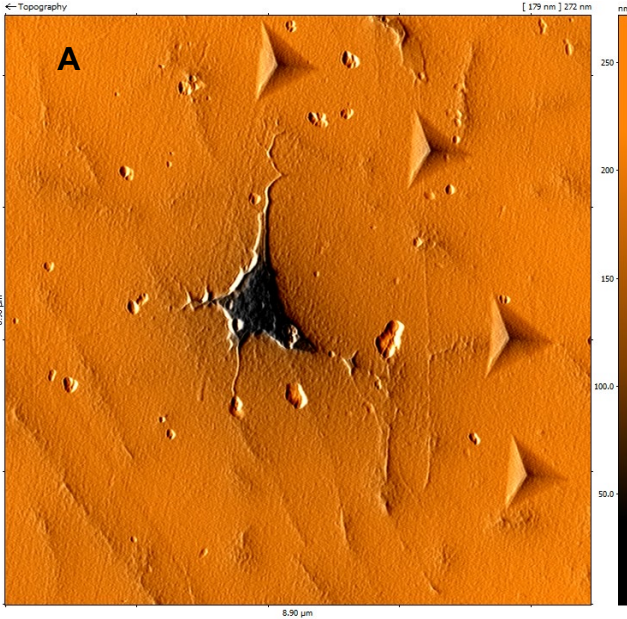
Yin et al. Biomacromolecules, 2011

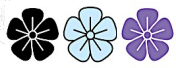


IMPACT AT CELL WALL SCALE - FLAX

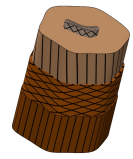
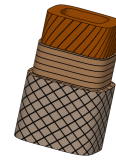


Flax plant walls after several injection cycles

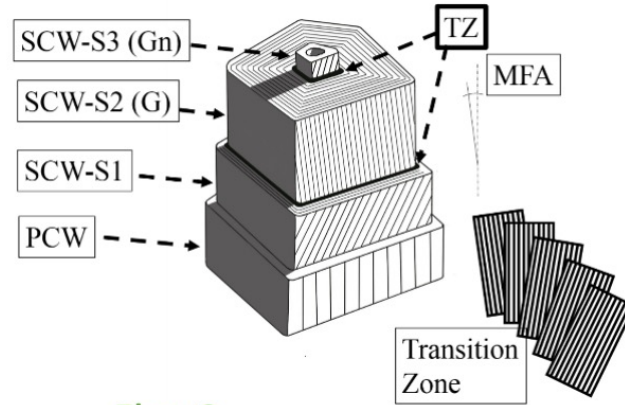
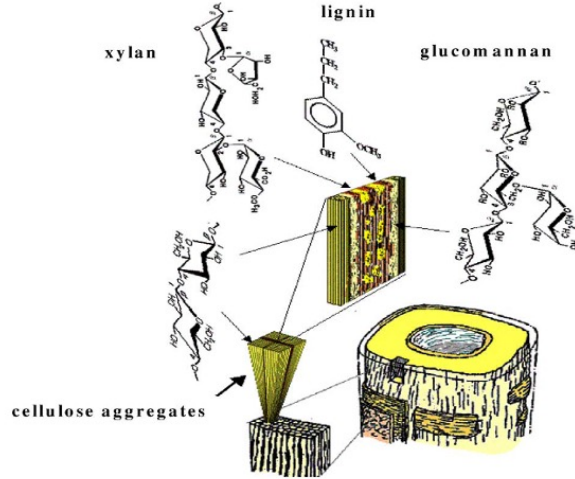




...BUT BIOCHEMICAL STRUCTURE IS DIFFERENT



Wood cell wall layer ultrastructure [Salmén, C.R. *Biologies*, 2004]



Wood S₂

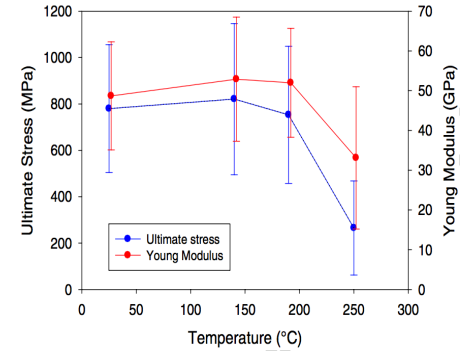
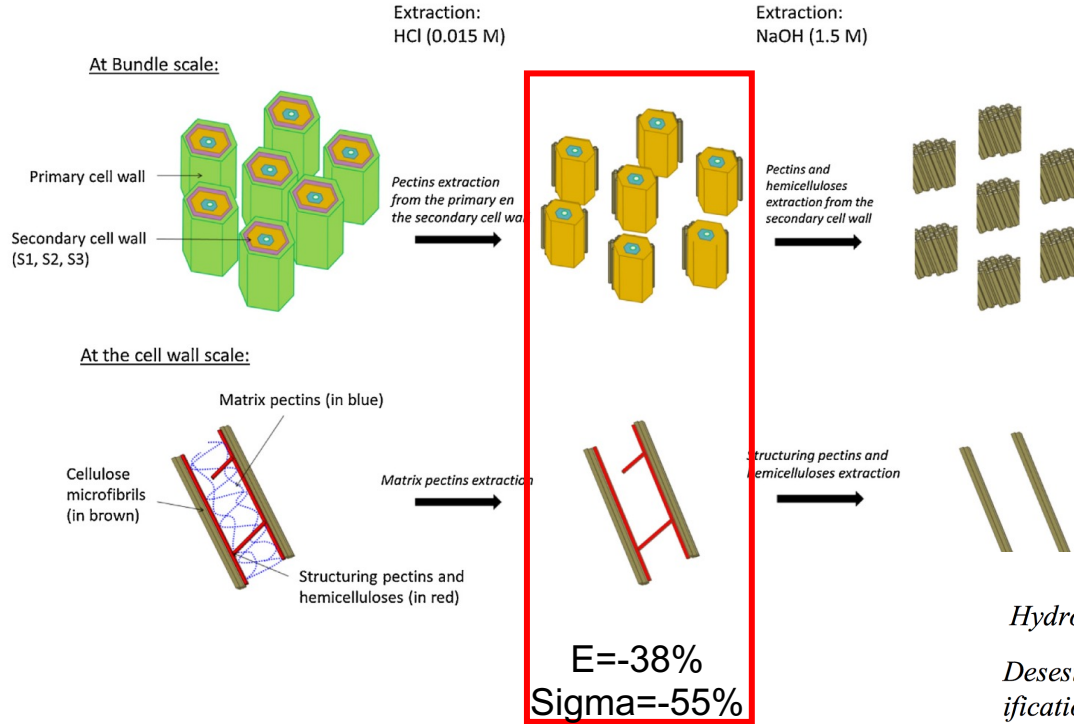
50% cellulose
 25% hemicelluloses - pectines
 25% lignin
 + extractives, ashes
 MFA = (0) 8-40°

Flax G

80% cellulose
 20% hemicelluloses - pectines
 ≈0% lignin
 + protein
 MFA = (0) 5-10°



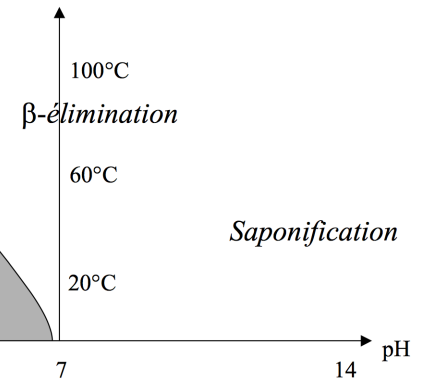
IMPACT OF NON CELLULOSIC POLYMERS ON PERFORMANCES



Gourier et al., Comp Part A, 2014

What about the effect of temperature on these polymers?

Hydrolyse
Desestérification



Renard, seminar INRA, 2010



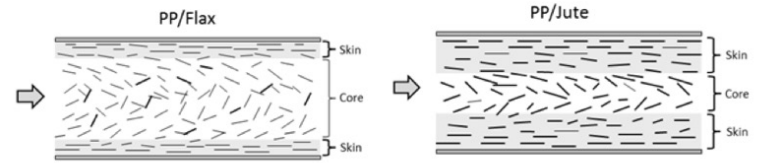
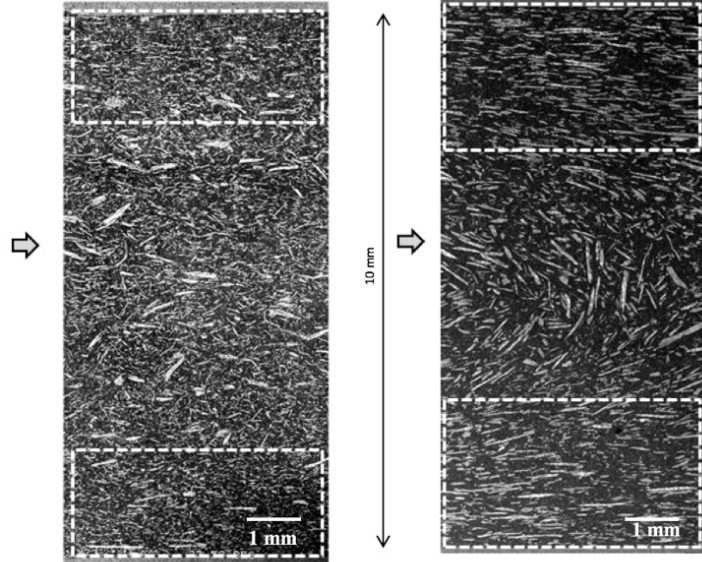
IMPACT ON COMPOSITE STRUCTURE



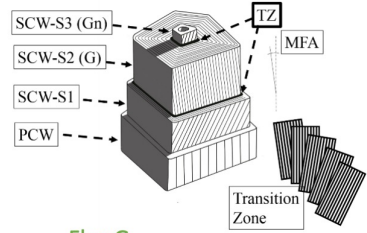
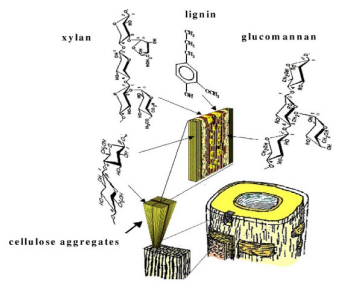
Injection



Flow direction →



Wood cell wall layer ultrastructure
[Salmén, C.R. *Biologies*, 2004]



Wood S₂
50% cellulose
25% hemicelluloses - pectines
25% lignin
+ extractives, ashes
MFA = (0) 8-40°

Flax G
80% cellulose
20% hemicelluloses - pectines
≈0% lignin
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MFA = (0) 5-10°



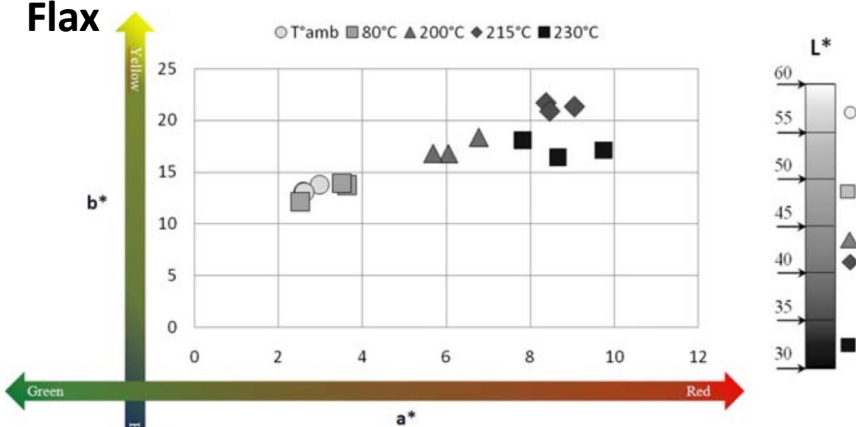
A strong influence of biochemical composition on orientation behaviour



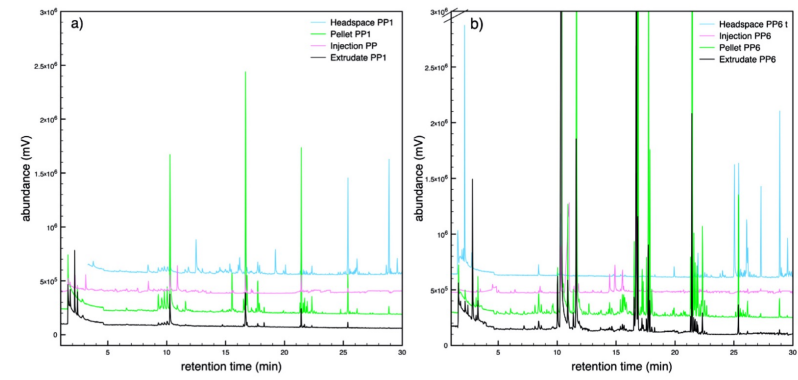
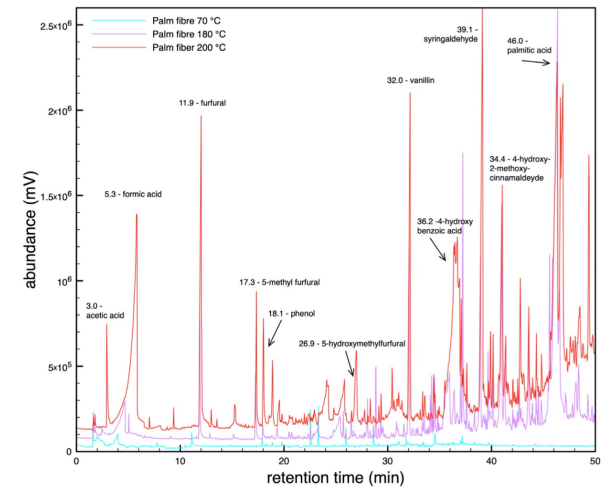
IMPACT ON VOC EMISSIONS



Flax



Palm

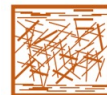




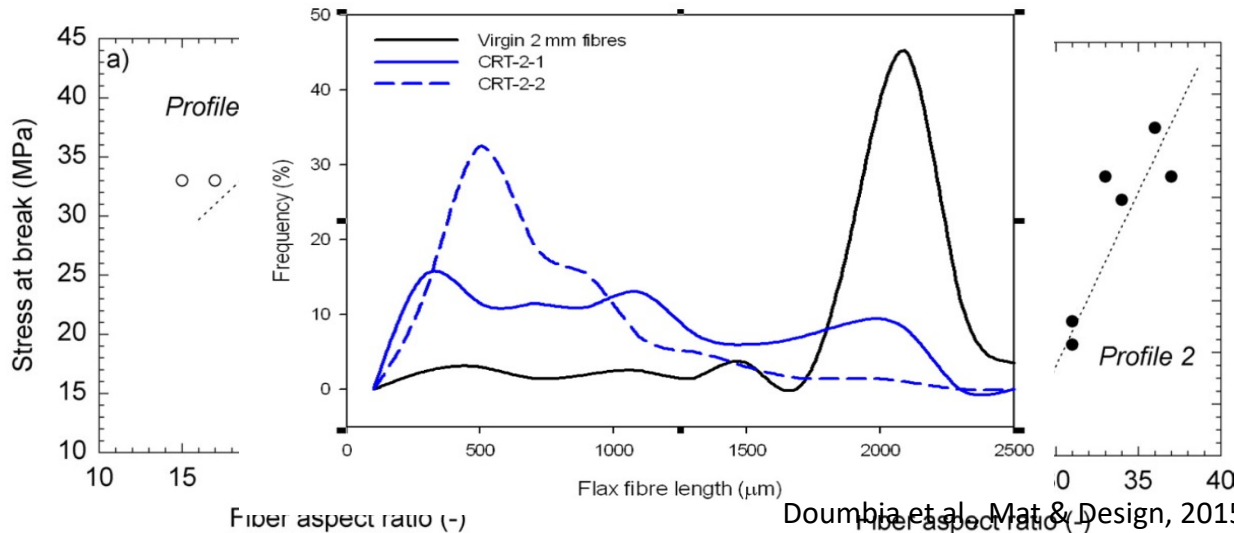
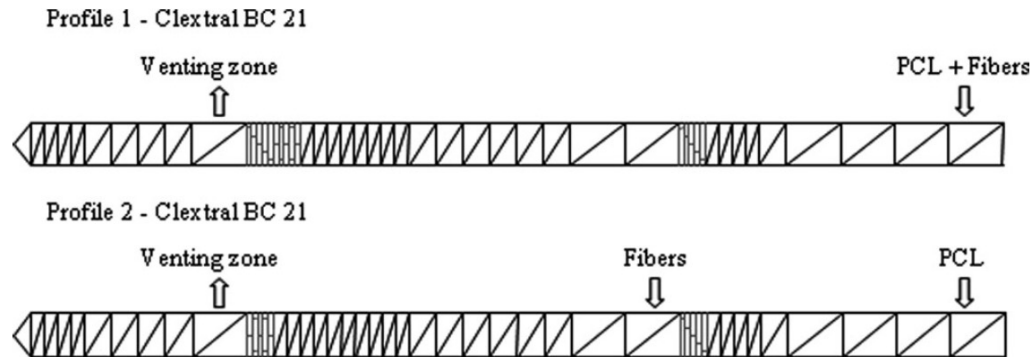
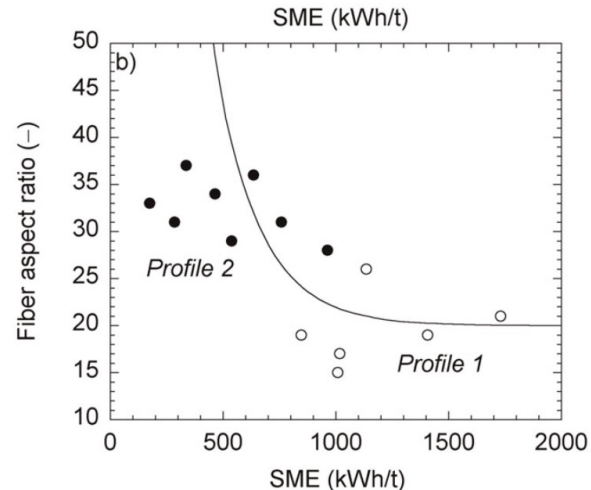
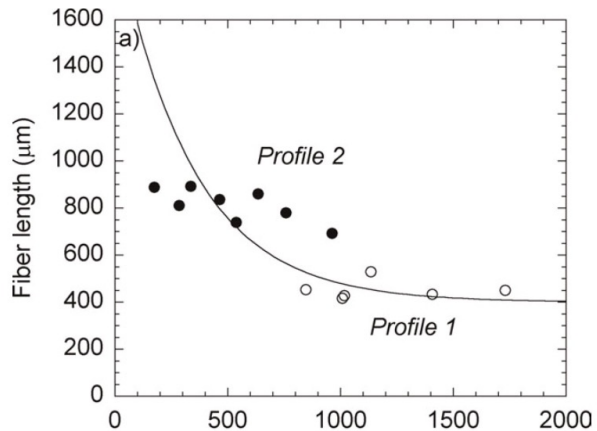
ADJUSTING THE PROCESS



Advantage: preserve fibre lengths



Injection



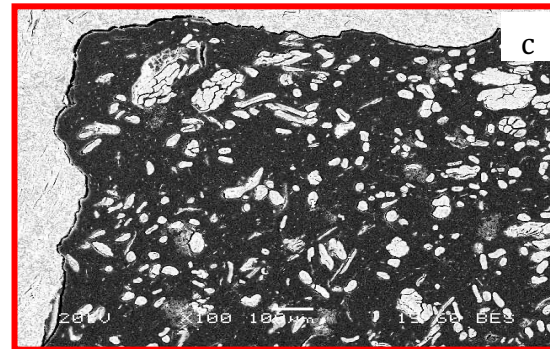
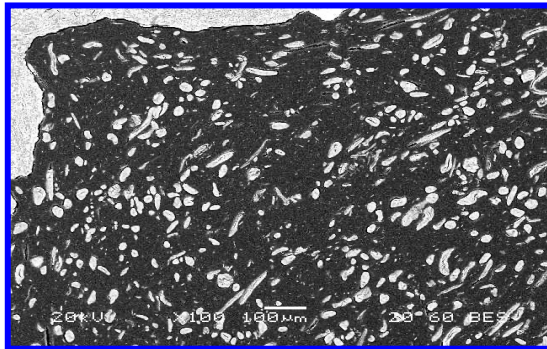


Compounding tool	Diameter (mm)	Dispersion factor R	Composite modulus (Mpa)	Composite strength (Mpa)
Single screw	28.6	0.94	3702±121	33.6±0.1
Twin screw	15.1	1.21	4405±133	37.4±0.1
Buss Hard	18.2	1.26	4419±157	38.5±0.5
Buss Soft	24.9	1.01	3924±196	35.6±0.3

Coroller, PhD Thesis, 2012

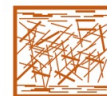
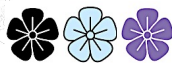
BUSS

TWIN-SCREW



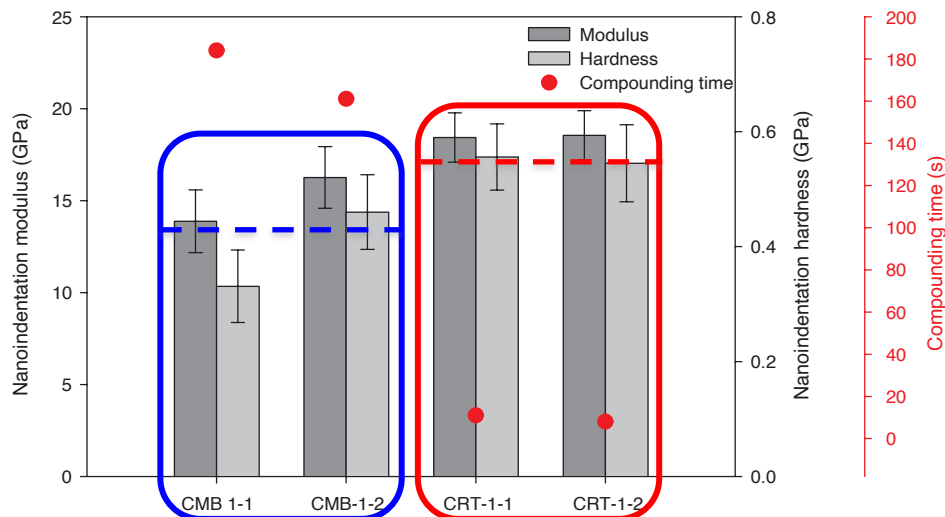
Injection

Higher composite stress for BUSS, similar stiffness



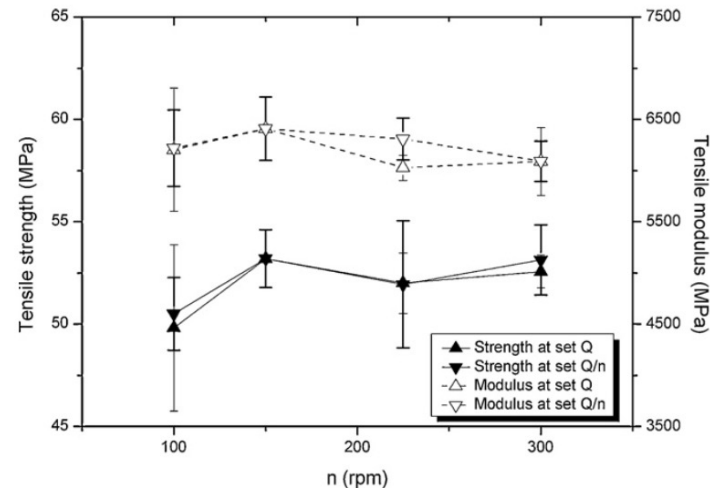
Injection

SHOULD RESIDENCE TIMES OR TEMPERATURE BE LIMITED?



BUSS TWIN SCREW

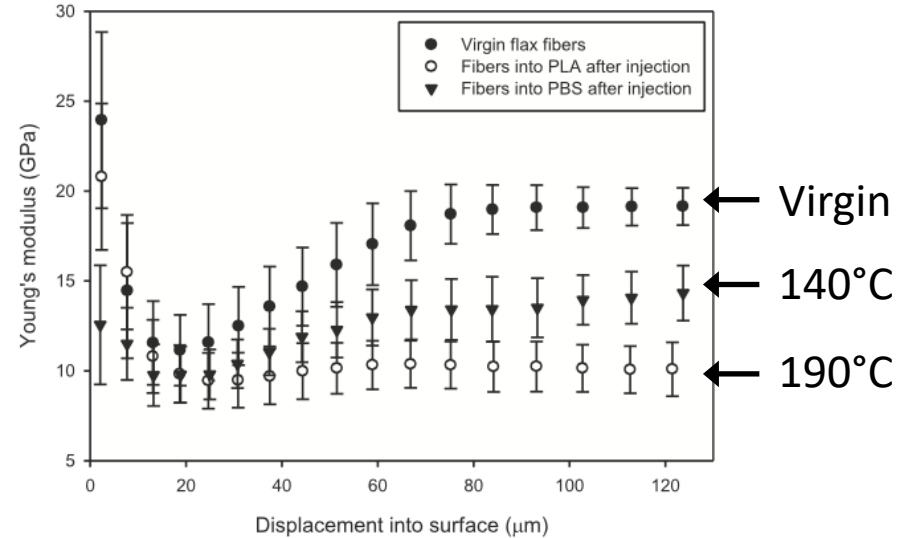
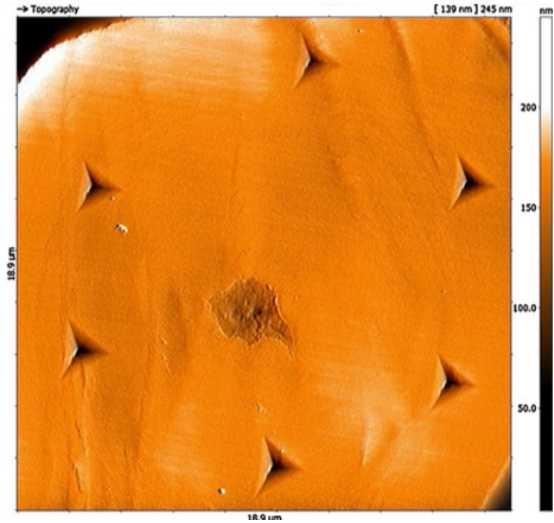
Doumbia et al., Mat & Design, 2015

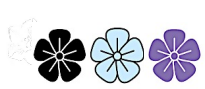


Gamon et al. Ind Crop & Prod, 2013

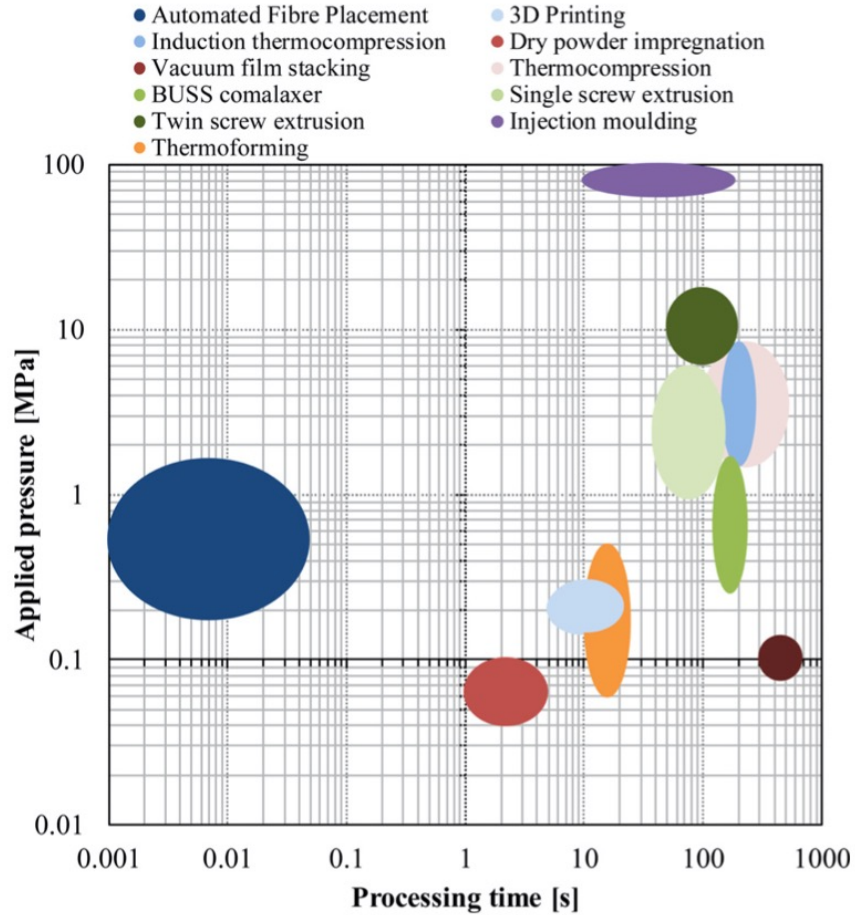


SHOULD RESIDENCE TIMES OR TEMPERATURE BE LIMITED?



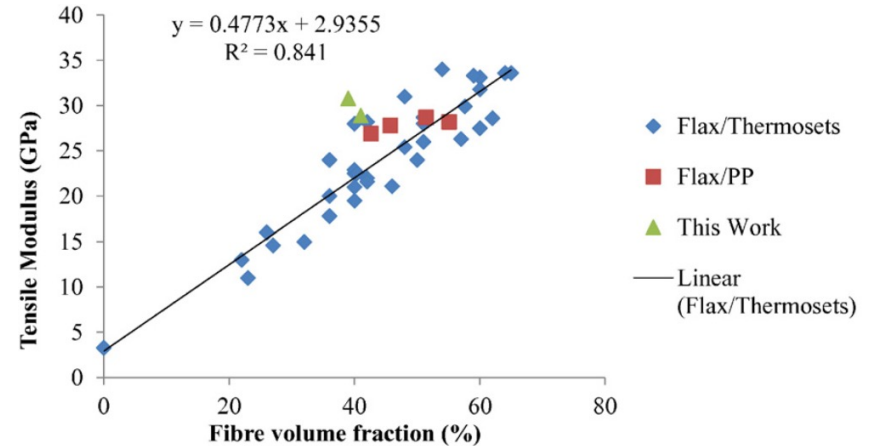
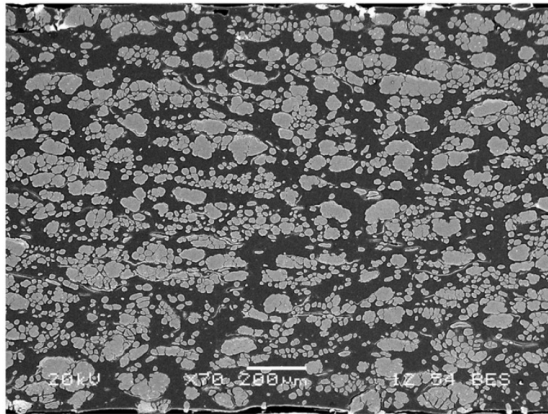
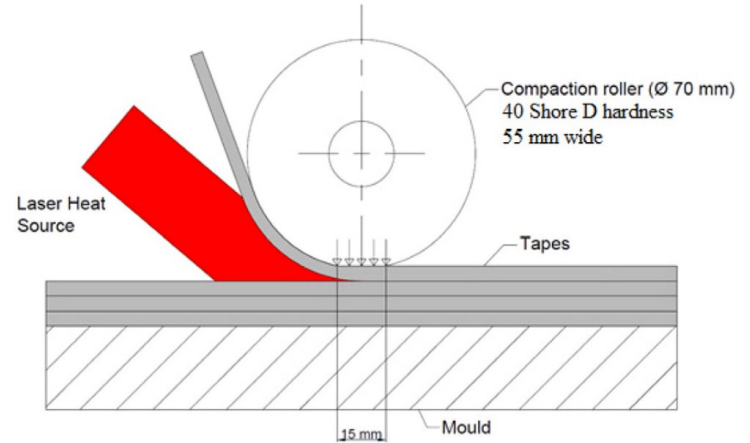
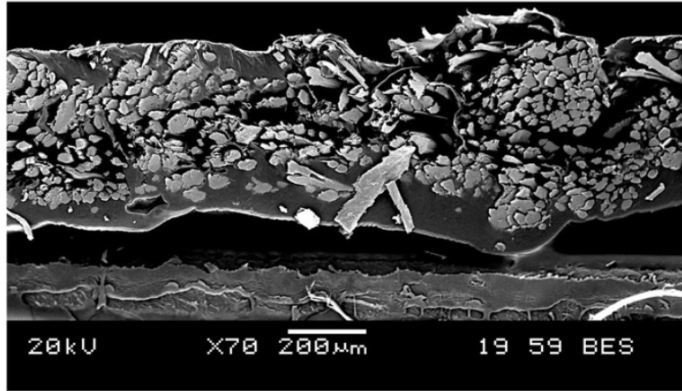


ADJUSTING THE PROCESS





ADJUSTING THE PROCESS – EXAMPLE OF AFP



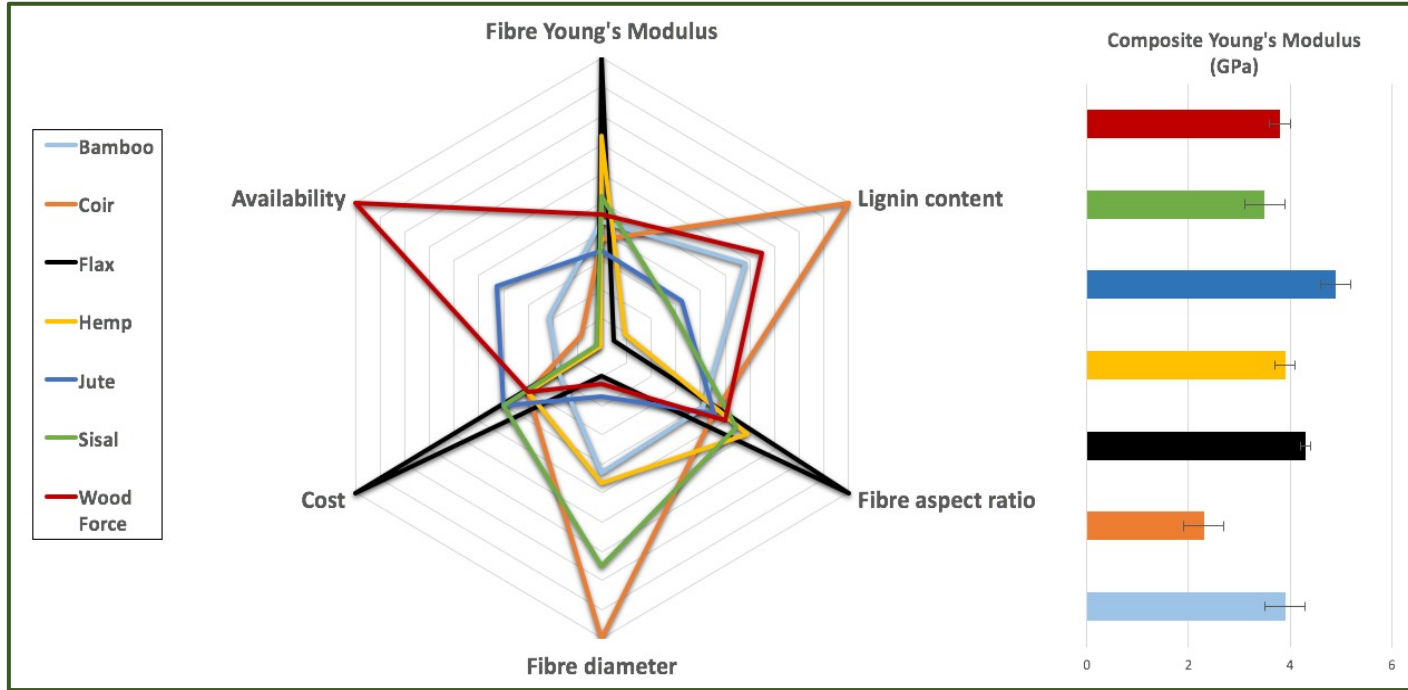


OR AN APPROPRIATE CHOICE OF PLANT FIBRES ?



Injection

Assesment / Injection



- Same matrix (PP)
- Same compounding way
- Same testing

It is necessary to take other parameters than fibre modulus into consideration